Potassium supplement upregulates the expression of renal kallikrein and bradykinin B₂ receptor in SHR

LAN JIN, LEE CHAO, AND JULIE CHAO
Department of Biochemistry and Molecular Biology, Medical University of South Carolina, Charleston, South Carolina 29425

Potassium supplement upregulates the expression of renal kallikrein and bradykinin B₂ receptor in SHR. Am. J. Physiol. 276 (Renal Physiol. 45): F476–F484, 1999.—High potassium intake is known to attenuate hypertension, glomerular lesion, ischemic damage, and stroke-associated death. Our recent studies showed that expression of recombinant kallikrein by somatic gene delivery reduced high blood pressure, cardiac hypertrophy, and renal injury in hypertensive animal models. The aim of this study is to explore the potential role of the tissue kallikrein-kinin system in blood pressure reduction and renal protection in spontaneously hypertensive rats (SHR) on a high-potassium diet. Young SHR were given drinking water with or without 1% potassium chloride for 6 wk. Systolic blood pressure was significantly reduced beginning at 1 wk, and the effect lasted for 6 wk in the potassium-supplemented group compared with that in the control group. Potassium supplement induced 70 and 40% increases in urinary kallikrein levels and renal bradykinin B₂ receptor density, respectively (P < 0.05), but did not change serum kininogen levels. Similarly, Northern blot analysis showed that renal kallikrein mRNA levels increased 2.7-fold, whereas hepatic kininogen mRNA levels remained unchanged in rats with high potassium intake. No difference was observed in β-actin mRNA levels in the kidney or liver of either group. Competitive RT-PCR showed a 1.7-fold increase in renal bradykinin B₂ receptor mRNA levels in rats with high potassium intake. Potassium supplement significantly increased water intake, urine excretion, urinary kinin, cAMP, and cGMP levels. This study suggests that upregulation of the tissue kallikrein-kinin system may be attributed, in part, to blood pressure-lowering and diuretic effects of high potassium intake.

Potassium supplement upregulates the expression of renal kallikrein and bradykinin B₂ receptor in SHR.

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ously hypertensive rats (SHR) caused sustained reduction of blood pressure for several weeks (7, 19, 46, 53). These findings demonstrated a direct linkage between tissue kallikrein gene expression and blood pressure regulation.

To explore the potential role of the tissue kallikrein-kinin system in blood pressure reduction after potassium supplement, we analyzed the expression of the system components in SHR. The results showed that a high potassium intake induced increases in the expression of renal kallikrein and bradykinin B2 receptor as well as increases in urine excretion, kinin, cGMP, and cAMP levels. Activation of the renal kallikrein-kinin system may be attributed to the blood pressure-lowering and diuretic effects of high potassium intake.

METHODS

Animal treatment. Young SHR (male, 4 wk old, 50–70 g) were purchased (Harlan Sprague Dawley, Indianapolis, IN). Rats were housed at a constant room temperature (25°C) with a 12:12-h light-dark cycle and had free access to rat chow and tap water. SHR were randomly divided into two groups with six animals in each group. The control group was given regular tap water and the experimental group was given 1% KCl in tap water. All procedures were in accordance with the Guide for the Care and Use of Laboratory Animals (National Institutes of Health, Bethesda, MD).

Blood pressure measurement. Systolic blood pressure of SHR was measured with a manometer-tachometer (Nastume KN 210; Nastume Seisakusho, Tokyo, Japan) using a tail-cuff method (46). Unanesthetized rats were placed in a plastic holder mounted on a thermostatically controlled warm plate that was maintained at 37°C during measurements. An average of 10 readings was taken for each animal after they became acclimated to their environment. Body weight and heart rate were recorded at the same time as blood pressure was monitored.

Urine collection. Twenty-four-hour urine was collected in metabolic cages 6 wk after potassium supplement. To eliminate the contamination of urine samples during urine collection period, rats were not given food but were given only tap water with or without 1% KCl solution in control and experimental groups. The urine samples were collected 24 h later and centrifuged at 1,000 g to remove particles. The urine volume was measured, and the supernatant was analyzed for kallikrein, kinin, cAMP, and cGMP levels.

Tissue homogenate preparation and protein determination. Six weeks after receiving tap water or 1% KCl, rats were anesthetized intraperitoneally with pentobarbital sodium (50 mg/kg body wt). Five ml blood was withdrawn directly through the heart. The vena cava was cut and the heparin (100 U/rat) was injected into the left ventricle. The circulation was perfused with 60 ml normal saline until tissues appeared bloodless. The kidney was quickly removed, minced, and homogenized with a Polytron (Brinkmann Instruments, Westbury, NY) in PBS, pH 7.0. The homogenate was centrifuged at 600 g for 10 min. The supernatant was incubated in 0.5% sodium deoxycholate and then centrifuged at 10,000 g for 30 min. Protein concentration was determined by the method of Lowry et al. (21). The protein extracts were used to measure intrarenal kallikrein levels, and sera were used to measure kininogen levels.

Membrane protein preparation. Rat kidneys were rinsed in ice-cold saline and minced by scissors. Tissues were suspended in 50 mM Tris- HCl buffer, pH 7.4, with 5 mM EDTA with a hand-held glass homogenizer. The suspension was centrifuged at 500 g for 5 min, and the supernatant was centrifuged again at 40,000 g for 20 min to pellet membranes. Membrane proteins were aliquoted and stored at −80°C.

Kallikrein RIA. Urinary and intrarenal kallikrein levels were determined by a direct RIA (39). The iodogen method was used to label 5 µg of purified rat tissue kallikrein. A GF-5 column (Pierce, Rockford, IL) was used to separate the unlabeled and labeled kallikrein.125I-labeled kallikrein (100 µl; 10,000 cpm/100 µl), 100 µl tissue kallikrein antisera (at a 1:200,000 dilution), 100 µl sample, and 100 µl assay buffer containing 1% BSA in PBS, bringing to a final volume of 400 µl, were incubated at 4°C overnight. Separation of free kallikrein and antibody-bound kallikrein was performed by centrifugation at 3,500 g for 30 min after adding 400 µl of 1% bovine γ-globulin and 800 µl of 25% polyethylene glycol in PBS. The standard kallikrein used ranged from 80 pg to 10 ng.

Determination of kininogen levels. Kininogen levels in rat sera were measured as described previously (8). Sera (50 µl) were added to 450 µl 0.02 M Tris·HCl, pH 8.0, and boiled for 30 min to eliminate kininase activity. Forty micrograms N-tosyl-L-phenylalanine chloromethyl ketone-trypsin (Sigma, St. Louis, MO) in 400 µl of 0.02 M Tris·HCl, pH 8.0, was added to 100 µl supernatant of boiled sera after 5 min of microcentrifugation. Samples were incubated at 37°C for 10 min, and the reaction was stopped by boiling for 10 min. The aliquots were used in a kinin RIA as described (38). Briefly, 100 µl of 125I-labeled [Tyr0]bradykinin ([Tyr0]BK; 10,000 cpm/100 µl), 100 µl rabbit antisera against bradykinin (at a 1:100,000 dilution), 100 µl diluted sample, and 100 µl 0.1% assay buffer (0.1% egg albumin, 10 mM EDTA, 3 mM 1,10-phanthrolin in PBS, pH 7.0) in a final volume of 400 µl were incubated at 4°C overnight. After addition of 400 µl of 1% bovine γ-globulin and 800 µl of 25% polyethylene glycol in PBS to the reaction mixture, free and antibody-bound bradykinin were separated by centrifugation at 3,500 g for 30 min. The standard bradykinin used ranged from 4 to 500 pg. Kininogen levels were expressed as micrograms kinin equivalents per milliliter serum.

Bradykinin B2 receptor binding assay. Synthetic [Tyr0]BK was used as radioligand for bradykinin B2 receptor binding studies. [Tyr0]BK was labeled as described previously (19). For saturation studies, aliquots of the membrane extract (100 µg protein) were incubated in duplicate for 2 h at 25°C in the binding buffer consisting of 1 mM 1,10-phanthrolin, 140 µg/ml bacitracin, 1 mM SQ-14225 (captopril), 1 mM DTT, and 0.1% BSA in 25 mM TES, pH 6.8, in the presence of increasing amounts of 125I-[Tyr0]BK. Specific binding was calculated by subtracting nonspecific binding obtained in the presence of excess unlabeled bradykinin (0.1 mM) from total binding obtained in the absence of unlabeled peptide. The final assay volume was 0.5 ml. At the end of the incubation, 4 ml of washing buffer (0.1% BSA in 25 mM TES buffer, pH 6.8) was added. The reaction mixture was filtered on a Whatman GF/C glass fiber filter (1.2 µm) previously soaked for at least 2 h in 0.1% polyethyleneimine. The filter was washed four additional times with 4 ml of washing buffer. The filter-bound radioactivity was detected in a gamma counter. Results were calculated by Scatchard transformation of binding data using the Kinetic Radiolig computerized program (27) and expressed as means ± SE of three independent experiments conducted with three different membrane preparations from each group of rats.

RIA of urinary cAMP. Urinary cAMP levels were determined by a RIA as previously described (9). cAMP (5 µg) was labeled with 1 mCi of 125I-jodide and incubated with chlo-
mine-T (Sigma) for 30 s at room temperature followed by addition of 50 µl 25% acetic acid. Iodinated cAMP in 50 mmol/l potassium phosphate buffer, pH 7.0, was separated on a reversed-phase C-18 HPLC column in an acetonitrile gradient (10% solution A containing 0.1% trifluoroacetic acid and 90% solution B containing 100% acetonitrile in 0.1% trifluoroacetic acid). Samples (100 µl) were acetylated with 5 µl acetylation agent of triethylamine and acetic anhydride in 2:1 ratio and were then added to 900 µl 50 mM sodium acetate buffer, pH 6.0. The reaction mixture, containing 100 µl 125I-labeled cAMP (12,000 cpm/100 µl), 100 µl cAMP antiserum (at a 1:20,000 dilution) in assay buffer (1% BSA in 50 mM sodium acetate buffer, pH 6.0), and 100 µl sample in a final volume of 300 µl, was incubated at 4°C overnight. Free and antibody-bound cAMP were separated by centrifugation at 1,500 g for 30 min after incubation for 20 min with 50 µl 1% bovine γ-globulin and 500 µl of 25% polyethylene glycol in PBS. The standard cAMP ranged from 0.8 to 200 pg.

RIA of urinary cGMP. Urinary cGMP levels were determined by a RIA as described (9). cGMP (5 µg) was labeled with 1 mCi of [125I]iodide using the same method as labeling cAMP. The reaction mixture, containing 25 µl 125I-labeled cAMP (15,000 cpm/25 µl), 25 µl cGMP antiserum (at a 1:1,400 dilution), 25 µl assay buffer (1% BSA in 50 mM sodium acetate buffer, pH 4.75) in a final volume of 100 µl was incubated at 4°C overnight. Free and antibody-bound cGMP were separated by centrifugation at 1,400 g for 20 min after incubation for 1 h with 50 µl 5% diluted human plasma (50 mM sodium acetate buffer, pH 4.75) and 1 ml 12% polyethylene glycol (50 mM sodium acetate buffer, pH 6.2). The standard cGMP ranged from 20 pM to 10 nM.

RNA preparation. Total RNA was extracted from fresh tissues by the guanidine isothiocyanate-cesium chloride gradient ultracentrifugation method (35). The extracted RNA was dissolved in diethyl pyrocarbonate-treated water. The concentration of RNA was determined by the absorbancy at 260 nm. The RNA was stored at −80°C until use.

Northern blot analysis. Nick-translated cDNA probes of rat tissue kallikrein-kininogen and α-actin were used for Northern blot analysis. Total kidney RNA (20 µg) and liver RNA (10 µg) from rats were separated by electrophoresis on a 1.5% agarose gel containing 0.66 M formaldehyde. The RNAs were transferred to Immobulin-N membranes in 20 mM sodium citrate-sodium phosphate–EDTA solution overnight. After crosslinking, the membrane was prehybridized in buffer (5× sodium chloride-sodium phosphate–EDTA (SSPE), 10× Denhardt, 0.5% SDS, and 100 µg/ml herring sperm DNA) at 60°C for at least 4 h. Nick translation for labeling the cDNA probes was performed using α-32P DATP (New England Nuclear Research Products, Boston, MA), according to the instructions of the manufacturer (Bethesda Research Laboratories, Bethesda, MD). A G-50 spin column was used to remove unincorporated components. The specific activity of the probe was ~2 × 108 cpm/µg DNA. After hybridization at 60°C for 16–18 h, the membrane was washed with 2× SSPE and 0.1% SDS at 60°C and exposed to X-ray film at −80°C. The blot was stripped and reprobed with the β-actin cDNA probe. The films were scanned to Adobe Photoshop 4.0 with a Hewlett Packard ScanJet and Image 1.47 computer software package. The ratio between the intensities of the competitor and target PCR products was plotted against the concentration of competitor cDNA added to the samples. The quantity of bradykinin B2 receptor mRNA was taken as the absorbance value that corresponded to a ratio of 1 on the ordinate axis.

Statistical analysis. Data were analyzed using standard statistical methods. Repeated blood pressure measurements were taken for comparison between control and experimental groups at each time point with the use of unpaired Student’s t-test. Group data are expressed as means ± SE. Values were considered significantly different at a value of P < 0.05.

RESULTS

Effect of high potassium intake on blood pressure in SHR. The effects of high potassium intake (1% KCl in drinking water) on blood pressure of young SHR were monitored weekly from 1 to 6 wk postsupplement. The basal blood pressure was 130 mmHg in both groups before potassium supplement. Potassium supplement caused a significant delay of blood pressure rise beginning on week 1 and the effect lasted for 6 wk (Fig. 1). At 1 wk postsupplement, the rise of systolic blood pressure of SHR given KCl in drinking water was significantly reduced compared with that in the control rats (131.9 ± 2.2 vs. 141.1 ± 2.3 mmHg, n = 6, P < 0.05). A maximal blood pressure reduction was observed 3 wk postsupplement (131.9 ± 2.5 mmHg, n = 6, P < 0.001). Significant reduction of blood pressure was observed from 2 to 6 wk in SHR given potassium supplement compared with that of control rats.

Physiological parameters after potassium supplement. Table 1 shows the physiological parameters in SHR 6 wk postsupplement. The systolic blood pressure was significantly reduced in SHR given 1% KCl in drinking water compared with that in control rats given tap water (173 ± 6.2 vs. 185.1 ± 2.5 mmHg, n = 6, P < 0.05). No significant differences in body weight or heart rate of both groups were observed. However, there were significant increases in sodium intake (18.6 ± 1.4 vs. 8.8 ± 0.7 ml·100 g body wt·1·day−1, n = 6, P < 0.001) and urine excretion.
Fig. 1. Blood pressure profiles of young spontaneously hypertensive rats (SHR) given 1% KCl in drinking water or tap water. Systolic blood pressure is expressed as means ± SE (n = 6). Bars represent SE. *P < 0.05 between control rats given tap water and rats given 1% KCl in drinking water. Statistical analysis was performed by unpaired Student’s t-test.

(15.3 ± 1.7 vs. 6.3 ± 0.5 ml · 100 g body wt −1 · day −1, n = 6, P < 0.001) between potassium-supplemented and control rats.

Table 1. Physiological parameters after potassium supplement

<table>
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<th>Variables</th>
<th>Control</th>
<th>1% KCl</th>
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</thead>
<tbody>
<tr>
<td>Blood pressure, mmHg</td>
<td>185.1 ± 2.5</td>
<td>173.6 ± 1.4*</td>
</tr>
<tr>
<td>Body wt, g</td>
<td>274.4 ± 8.6</td>
<td>270.9 ± 5.0</td>
</tr>
<tr>
<td>Heart rate, beats/min</td>
<td>370.0 ± 1.0</td>
<td>361.8 ± 2.4</td>
</tr>
<tr>
<td>Urine excretion, ml · 100 g body wt −1 · day −1</td>
<td>6.3 ± 0.5</td>
<td>15.3 ± 1.7†</td>
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<tr>
<td>Water intake, ml · 100 g body wt −1 · day −1</td>
<td>8.8 ± 0.7</td>
<td>18.6 ± 1.4‡</td>
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Values for each group are means ± SE (n = 6). Four-week-old spontaneously hypertensive rats (SHR) received 1% KCl in their drinking water, and physiological measurements were performed at 6 wk after potassium supplement compared with control rats receiving tap water. Statistical significance between the 2 groups was determined by unpaired Student's t-test; *P < 0.05 and †P < 0.001.

Fig. 2. Representative saturation binding curve and Scatchard plot (inset) for kidney membrane proteins from control rats and rats with potassium supplement. Tyr-BK, [Tyr0]bradykinin; B/F, bound/free.

Urinic levels were not altered (27.3 ± 1.9 vs. 28.0 ± 2.7 ng/mg protein, n = 6) between the two groups. Similar kininogen levels in serum (3.3 ± 0.3 vs. 3.4 ± 0.3 μg kinin/ml serum, n = 4) were observed between experimental and control groups.

Table 2. Immunoreactive tissue kallikrein, kininogen, and renal bradykinin B2 receptor levels in rats with or without potassium supplement

<table>
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<th>Variables</th>
<th>Control</th>
<th>1% KCl</th>
</tr>
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<tr>
<td>Urinary kallikrein, μg/day</td>
<td>35.4 ± 2.0</td>
<td>58.8 ± 6.3†</td>
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<tr>
<td>Intrarenal kallikrein, ng/mg protein</td>
<td>28.0 ± 2.7</td>
<td>27.3 ± 1.9</td>
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<tr>
<td>Total kininogen, μg kinin equivalent/ml serum</td>
<td>3.4 ± 0.3</td>
<td>3.3 ± 0.3</td>
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<tr>
<td>Renal B2 receptor density, pM/mg protein</td>
<td>59.2 ± 5.2</td>
<td>81.1 ± 0.8*</td>
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</table>

Values are means ± SE. Immunoreactive tissue kallikrein, kininogen, and renal bradykinin B2 receptor measurements were performed at 6 wk after potassium supplement. Tissue kallikrein levels in the kidney and urine were measured by direct kallikrein RIA. Total kininogen levels expressed as μg kinin equivalent/ml serum were determined by RIA measuring amount of released kinin after trypsin treatment. Renal bradykinin B2 receptor densities were measured by receptor binding assay. *P < 0.05 and †P < 0.001, respectively, between control and potassium group. Statistical analysis was performed by unpaired Student’s t-test.
control group, whereas intrarenal kallikrein levels remained the same after 6 wk of potassium loading. Our results showed that potassium supplement caused increased expression of renal kallikrein. Because urinary kallikrein is mainly originated from the kidney, these results indicate that rapid secretion of renal kallikrein into the urine may be attributed to the unchanged renal kallikrein content. In the liver (Fig. 3B), no significant change of kininogen mRNA was observed between potassium and control groups (115.7 ± 4.7 vs. 102.3 ± 8.3 densitometric units, n = 3). No difference was observed in β-actin mRNA levels in the liver of either group.

Effect of potassium supplement on bradykinin B₂ receptor mRNA levels. The efficiency of amplification of the competitor to that of bradykinin B₂ was first tested by kinetic analysis. We found that the same molar quantity of a competitor and a target yielded a similar amount of products after various PCR cycles. Figure 4A shows electrophoretic profiles of the bradykinin B₂ receptor (target) and its competitor in the kidney of control (Fig. 4A, left) and potassium-loaded (Fig. 4A, right) rats. The competitor concentration used in PCR ranged from 0.01 to 5.00 pM. The ratio between the competitor and target PCR products was plotted against the amount of added competitor cDNA. The quantity of B₂ mRNA was taken as the abscissa value that corresponded to a ratio of 1 on the ordinate axis using linear regression analysis to fit the data. Figure 4B shows that bradykinin B₂ receptor mRNA levels in the kidney were significantly higher in potassium-supplemented rats than that in control rats (0.46 ± 0.10 vs. 0.27 ± 0.10 pM, n = 6, P, 0.05, respectively).

Effect of potassium supplement on urinary kinin, cAMP, and cGMP levels. Figures 5 and 6 show urinary kinin, cAMP, and cGMP levels measured by their respective RIAs. After 6 wk of potassium supplement there were significant increases in urinary kinin levels (2.7 ± 0.4 vs. 1.5 ± 0.3 ng·100 g body wt⁻¹·day⁻¹, n = 6, P < 0.05) (Fig. 5), cAMP levels (152.5 ± 10.3 vs. 106.6 ± 9.7 nmol·100 g body wt⁻¹·day⁻¹, n = 6, P < 0.05), and cGMP levels (31.6 ± 2.4 vs. 22.7 ± 1.6 nmol·100 g body wt⁻¹·day⁻¹, n = 6, P < 0.05) in rats receiving potassium supplement compared with those in control rats receiving tap water (Fig. 6).

DISCUSSION

The present study demonstrated that high potassium intake attenuated the rise of blood pressure in SHR.
that was accompanied by upregulation of the expression of the renal kallikrein-kinin system. Potassium supplement induced increases in water intake, urine excretion, urinary kinin, cAMP, and cGMP levels. Previous studies employing transgenic and somatic gene delivery strategies showed that the expression of the tissue kallikrein transgene could induce prolonged reduction of blood pressure and attenuation of renal injury in various animal models (6, 19, 40, 48, 53). Together, these combined results suggest that the blood pressure-lowering and diuretic effects exerted by high potassium intake could be, in part, due to the activation of renal kallikrein-kinin-bradykinin receptor system components.

It has been reported that dietary KCl supplement blunted blood pressure rise in SHR but had no effect on blood pressure in normotensive Wistar-Kyoto (WKY) rats (24, 45). Furthermore, KCl in drinking fluid has been shown to cause increases in urine excretion and fluid intake, urinary potassium, and sodium excretion as well as urinary kallikrein excretion in normotensive Sprague-Dawley rats (32). These findings indicate that dietary KCl supplement affects urinary excretion and renal kallikrein excretion in both hypertensive and normotensive rats. A previous report showed that both serum K and aldosterone levels were increased after potassium supplement in rats (30, 54). In addition, aldosterone has been shown to stimulate kallikrein release and increase kallikrein protein/activity, without affecting kallikrein mRNA transcription (14, 16). Similarly, acute administration of aldosterone did not induce the synthesis of renal kallikrein (25). Our present study is the first one to demonstrate increases in both renal kallikrein protein and mRNA levels after potassium supplement.

Both potassium and thiazide diuretics induce increased kallikrein excretion and have diuretic and blood pressure-lowering effects (33). However, their mechanisms of action may not be the same. Thiazide diuretics act on the cortical diluting segment of the renal tubule and increase salt and water excretion primarily by inhibition of sodium and water reabsorption, whereas potassium intake produces effects similar to osmotic diuresis to increase potassium excretion (3). One to three weeks of thiazide treatment has minimal effect on blood pressure in SHR (31). Twenty-six weeks of thiazide therapy did decrease blood pressure in SHR (18). Our study showed that potassium supplement for 1 wk had a significant effect on blood pressure in SHR. Therefore, the effect of high potassium intake on early blood pressure reduction in SHR is not, for the most part, due to its diuretic actions but may be due to other related mechanisms. In this study, we did not observe body weight changes after potassium supplement. In agreement with our study, Barden and coworkers (2) also showed no change of body weight in rats with or without supplement with 1% KCl for 5 wk (2). The reason for the lack of body weight loss from marked diuresis with 1% KCl may be due to a similar magnitude of increased water drinking, which may offset the loss of body weight.

A proposed scheme for the action of the tissue kallikrein-kinin system in blood pressure reduction after...
derived relaxing factor, which in turn induces the
binding of kinin to bradykinin B2 receptor stimulating phospholipase A2 (PLA2) with increased prostacyclin formation. Increased urinary prostacyclin and its metabolites, such as 6-keto-PGF1α and PGF2α after potassium loading were found in several other studies and were implicated to be a consequence of elevated kallikrein activity and local kinin formation (2, 26, 30). Binding of prostacyclin to its receptor may result in stimulation of adenylate cyclase and increased cAMP levels. Our results that urinary cAMP levels were significantly elevated after potassium supplement suggest that increased cAMP may be involved in upregulation of the expression of tissue kallikrein and bradykinin B2 receptor genes due to a positive feedback mechanism. cAMP has been shown to enhance the synthesis and expression of bradykinin B2 receptor in cultured arterial smooth muscle cells (10). We also found that the mRNA levels of both bradykinin B2 receptor and tissue kallikrein were significantly increased by adding cAMP to primary cultured human renal proximal tubule cells (unpublished results). cAMP response elements were identified in the 5′-flanking region of the bradykinin B2 receptor and tissue kallikrein genes (29, 47). Collectively, these results support the notion that cAMP may upregulate the expression of renal kallikrein and bradykinin B2 receptor genes via positive feedback mechanisms.

Alternatively, activation of bradykinin B2 receptor may stimulate phospholipase C, which triggers nitric oxide formation (Fig. 7). Increased nitric oxide formation may result in stimulation of guanylate cyclase and increased cGMP levels (20). It has been shown that bradykinin stimulates the release of endothelium-derived relaxing factor, which in turn induces the production of cGMP via activation of bradykinin B2 receptors in cultured porcine arterial endothelial cells (36). Nitric oxide has been shown to be the epithelium-derived relaxing factor released by bradykinin in the guinea pig trachea (15). Bradykinin and the angiotensin-converting enzyme inhibitor (ramiprilat) enhance the levels of cytosolic calcium, prostacyclin, and nitric oxide in porcine brain capillary endothelial cells (49). Activation of B1 and B2 bradykinin receptors produces cGMP in cultured bovine aortic endothelial cells (50). Our results suggest that elevated kinin levels after potassium supplement may result in an increase in urinary cGMP formation. Elevated cGMP and cAMP levels have been shown to correlate with relaxation and antiproliferation of smooth muscle cells (20) and may induce vascular smooth muscle relaxation and thus account for the blood pressure-lowering effect in hypertensive rats after high potassium intake.

The long-term effects of dietary potassium on the renal end-organ damage were investigated in WKY rats and SHR. Albumin excretion rate (AER) was higher in SHR than in WKY rats. AER rose further with high sodium intake and was ameliorated by an addition of equimolar potassium in SHR. The graded histopathologic injury correlated well with measured AER. Major improvement in hypertensive renal lesions occurred in SHR with potassium supplement and salt loading. Potassium supplement has been shown to attenuate renal injury in SHR without affecting the blood pressure (12). These results show that potassium protected against renal lesions induced by salt loading independent of blood pressure effect in SHR. Also, a previous study suggested that low renal kallikrein levels may contribute to hypertension and renal disease (37). Reduced urinary or renal kallikrein levels have also been observed in a number of genetically hypertensive rats (1, 22). Long-term infusion of purified tissue kallikrein attenuated glomerular sclerotic lesions and tubular injury in hypertensive Dahl salt-sensitive rats without causing an apparent blood pressure reduction (44). Our recent study showed that adenoviral-mediated kallikrein gene delivery into Dahl salt-sensitive rats attenuated hypertension and renal injury induced by a high-salt diet (6). Taken together, these results demonstrated a direct linkage between tissue kallikrein expression and renal protection in hypertensive rats. The present studies show that high potassium intake upregulated tissue kallikrein and bradykinin B2 receptor gene expression in hypertensive rats, and elevated renal kallikrein-kinin system components may attribute, in part, to the protective effects of potassium against renal injury and hypertension.

This work was supported by National Heart, Lung, and Blood Institute Grants HL-29397 and HL-52196.

Address for reprint requests and other correspondence: J. Chao, Dept. of Biochemistry and Molecular Biology, Medical Univ. of South Carolina, 171 Ashley Ave., Charleston, SC 29425 (E-mail: Chaomnusc.edu).

Received 24 July 1998; accepted in final form 17 November 1998.

Fig. 7. Proposed scheme depicting potential roles of tissue kallikrein-kinin system in mediating effects of potassium.
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