Comparison of protective effects of trimetazidine against experimental warm ischemia of different durations: early and long-term effects in a pig kidney model

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Comparison of protective effects of trimetazidine against experimental warm ischemia of different durations: early and long-term effects in a pig kidney model. Am J Physiol Renal Physiol 292: F1082–F1093, 2007; doi:10.1152/ajprenal.00338.2006.—Acute renal failure (ARF) is often the consequence of an ischemia-reperfusion injury (IRI) and associated with high mortality. Warm ischemia (WI) is a crucial factor of tissue damage, and tissue destruction led by ischemia-reperfusion (I/R) can impact the early and long-term functional outcome. Trimetazidine (TMZ) is an anti-ischemic drug. Previously, we already verified its protective effect on a cold-ischemic pig kidney model by directly adding TMZ into the preservation solution (Faure JP, Baumert H, Han Z, Goujon JM, Favreau F, Dutheil D, Petit I, Barriere M, Tallineau C, Tillement JP, Carretier M, Mauco G, Papadopoulos V, Hauet T. Biochem Pharmacol 66: 2241–2250, 2003; Faure JP, Petit I, Zhang K, Dutheil D, Doucet C, Favreau F, Eugene M, Goujon JM, Tillement JP, Mauco G, Vandewalle A, Hauet T. Am J Transplant 4: 495–504, 2004). In this study, we aimed to study the potential effect of TMZ pretreatment (5 mg/kg iv 24 h before WI) on the injury caused by WI for 45, 60, and 90 min and reperfusion in a WI pig kidney model. Compared with sham-operated (control) and uninephrectomized animals (UNX), TMZ pretreatment significantly reduced deleterious effects after 45 min, and particularly 60 and 90 min of WI by improving the recovery of renal function and minimizing the inflammatory response commonly prevalent in ischemic kidney injury. Compared with controls (control group and UNX group), it was observed that 1) hypoxia-inducible factor-1 (HIF-1α) expression occurred earlier and with a higher intensity in the TMZ-treated groups; 2) the reduction of IRI during the first week following reperfusion was correlated with an earlier and greater expression of stathmin, which is involved in the process of tubular repair; and 3) the tubulointerstitial fibrosis was reduced, particularly after 60 and 90 min of WI. In conclusion, TMZ made the warm-ischemic kidneys more resistant to the deleterious impact of a single episode of I/R and reduced early and long-term subsequent damage.

mitochondrial damage; ischemia-reperfusion injury; stathmin; renal regeneration

ACUTE RENAL FAILURE (ARF) is a clinical syndrome defined as an acute loss of renal function and is associated with high morbidity and mortality (27, 34). Postischemic renal failure is a severe outcome mostly observed after cadaveric renal transplantation, hemorrhagic shock, or sepsis. Ischemia-reperfusion injury (IRI) is a multifactorial process that has both immediate and long-term effects. Mitochondria take a critical part in it due to their role in tubule cell metabolism, generation of reactive oxygen species (ROS), and the initiation of apoptosis. Consequently, agents which could act on this organite are interesting therapeutic tools (18, 22). Moreover, the vascular endothelium is also a key site of injury from ROS and endothelial cell dysfunction is central to the pathogenesis of chronic allograft nephropathy (2, 3). We have previously evidenced the protective effect of TMZ on kidney grafts from cold preservation and reperfusion, showing that TMZ was efficient when added to various currently used preservation solutions (11, 12). Several reports have described different pharmacological effects of TMZ which are relevant in the prevention of organ damage from IRI: partial inhibition of fatty acid β-oxidation (20), reduction of ionic imbalance during ischemia-reperfusion (I/R) (9), increase in phospholipid uptake turnover (33), and membrane protection against reoxygenation-induced stress (32, 28). Other studies have showed that TMZ reduces intracellular acidosis, preserves ATP synthesis, limits inflammatory processes, and therefore ROS generation, and prevents Ca2+ overload (for a review, see Ref. 23). A recent study has identified the TMZ protective effect against different oxidizing systems, namely, Cu, Fe/ascorbate, and met-myoglobin/H2O2 (34). It was suggested that TMZ could act as a metal chelator of the permeability transition (34) and that TMZ inhibits mPTP opening and protects the rabbit heart from prolonged ischemia-reperfusion injury (1). Collectively, these studies have demonstrated the protective effect of TMZ when the kidney is exposed to warm ischemia (WI)-reperfusion. It was also shown that TMZ is efficient in protecting the liver from IRI in a rat model (10). These effects were well correlated with an inhibition of mitochondrial damage, since TMZ restored ATP synthesis, NAD(P)H levels, and mitochondrial membrane potential and generation of mitochondrial permeability transition (10). Altogether, these data outline the central role of mitochondria and the correlation between its damage during WI and cell dysfunction during reperfusion. However, data available from these reports are limited to the early time of reperfusion.

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(a few hours to a few days) and the long-term effects of TMZ remain to be clarified.

The present preclinical study was designed to study early and long-term effects of TMZ against different WI times followed by reperfusion, mimicking different clinical situations such as aortic and renal surgery. To clarify the impact of TMZ, we have focused on new pathophysiological factors that have recently emerged in IRI. Hypoxia-inducible factor 1 (HIF-1α) in acute hypoxic injury is known to upregulate factors that have been shown to be “cytoprotective.” In addition, strategies that limit HIF prolyl-hydroxylation or HIF proteolysis may be powerful in improving IRI. Consequently, we have compared HIF expression in reperfused tissue exposed or not exposed to TMZ treatment. Acute tubular necrosis (ATN) is also characterized by a regeneration phase, which involves a sequence of events, including epithelial cell dedifferentiation and proliferation (2). Stathmin is an important molecule that is known to play a critical role in the process of mitosis and the regulation of the microtubule cytoskeleton. In ATN, recent reports have shown that stathmin is a marker of dedifferentiated, mitotically active epithelial cells that may contribute to tubular regeneration after ATN (44). Interestingly, in stathmin-deficient mice subjected to IRI, aberrant regulation of cell cycle progression, which impaired or at least delayed the process of tubular repair and recovery after acute renal injury, has been described (45). The other relevant aspect, the role of the inflammatory process in damaged kidneys, has been studied in in vitro and in vivo models, which have evidenced that inflammation plays a major role in the pathophysiology of ARF resulting from ischemia. Given that T lymphocytes are shown to be important pathogenic modulators in allo-antigen- independent injury to the kidney, leading to long-term dysfunction (5, 6), we have completed our experimental protocol with an analysis of cell populations involved in the inflammatory reaction within damaged tissue, adhesion molecule expression, and fibrosis development.

**METHODS**

**Surgical procedures and experimental groups.** Large white male pigs (INRA, J. C. Caritez, Le Magneraud, Surgères, France) were prepared as previously described (14). WI was induced by clamping the left renal pedicle for 45 (WI-45, n = 7), 60 (WI-60, n = 8), and 90 min (WI-90, n = 8) with a vascular nontraumatic clamp, followed by a right nephrectomy. TMZ was administered (5 mg/kg iv) 24 h post-WI (WI-45-TMZ, n = 7), 60 (WI-60-TMZ, n = 8), and 90-TMZ, n = 8). These experimental groups were compared with the uninunephectomized group (UNX; n = 8, matched for weight, nephrihonic mass, and age) and the control group (sham-operated; n = 6, matched for weight and age). All experiments were carried out in accordance with the Guidelines of the French Agricultural Office and legislation governing animal studies. Animals were followed up at 16 wk after WI.

**Functional parameters.** Endogenous creatinine clearance (C\text{Cr}; expressed in ml/min) and urinary proteins were measured as previously described (14). Pigs were placed in cages to allow specific 24-h urine collections. Plasma and urinary creatinine (C\text{r}) was measured enzymatically (Jaffé method, rate blanked and compensated, Roche Diagnostic, Mannheim, Germany), and the glomerular filtration rate was calculated by determination of C\text{Cr}, calculated using the following formula: urinary flow rate (UFR) × urinary C\text{r}/plasma C\text{r}. Urinary proteins were determined using a colorimetric method (Roche Diagnostic).

Supplementary animals were submitted to the same experimental conditions and killed at 3 h (n = 2) day 1 (n = 2), day 3 (n = 2), and day 7 (n = 2) for TR-PCR and Western blot studies in renal tissue.

**Real-time PCR using SYBR green for stathmin.** Between days 1 and 7, stathmin mRNA in the cortex and outer medulla was measured to assess the influence of TMZ treatment on renal regeneration. Real-time quantitative PCR amplification reactions were carried out using the Smart Cycler detection system (Cepheid, Sunnyvale, CA) in a 25-μl volume. The reaction mixture consisted of 1× QuantiTect SYBR Green PCR Master Mix (Qiagen, Courtabœuf, France) and an additional 0.5 mM MgCl₂, 0.5 and 1.25 U of AmpliTag Gold DNA polymerase (Roche, Meylan, France), and 0.4 μM of each primer. One microliter of prepared cDNA template was added to each reaction. To quantify stathmin, 18S RNA was used as an internal control. The RT-PCR for stathmin and 18S rRNA were performed on the same cDNA samples in separate tubes. The primer sequences for PCR were as follows: stathmin (GenBank accession no. AY644720), forward 5'-CAGGTGAARGARCTNGARAARC-3', reverse, 5'-ATTAGT-CAGClyCTCGTYTCTRCNGC-3'; and 18S rRNA forward, 5'-CGTCGCCGCCCCCTTGCCTC-3'; reverse 5'-GCTTTCGCTCTG-GTCCGTCTT-3'.

**Western blotting.** Pieces of snap-frozen kidney tissues (cortex and outer medulla) were disrupted in a mortar and then homogenized with a polytron at 0°C in Trition X-100 lysis buffer (50 mmol/l Tris, pH 7.6, 150 mmol/l NaCl, 1% Triton X-100, 0.1% SDS) containing 1% aprotinin, 2 mmol/l ε-aminocaproic acid, and 0.5 mmol/l phenylmethylsulfonyl fluoride. The homogenate was centrifuged at 2,000 g for 5 min. The supernatant was then centrifuged at 12,000 g for 5 min and retained. Protein concentrations were determined, and 50 μg of protein extracts, denatured in Laemmli buffer and separated on 12% SDS-PAGE, were electrotransferred to a nitrocellulose membrane. The membrane, blocked for 2 h at room temperature with TBS, 0.05% Tween 20, and 5% nonfat dry milk, was incubated overnight at 4°C with a 1:1,000-diluted rabbit anti-stathmin peptide antiserum against an amino acid sequence conserved across species as previously described (STC, Prof. A. Sobel, INSERM U.440, Paris, France) and an amino acid sequence conserved across species as previously described (STC, Prof. A. Sobel, INSERM U.440, Paris, France) and 1:1,000-diluted rabbit anti-stathmin peptide antiserum against an amino acid sequence conserved across species as previously described (STC, Prof. A. Sobel, INSERM U.440, Paris, France). The membrane was washed with immunoblotting buffer and probed with a 1:1,000-diluted rabbit anti-stathmin antiserum against an amino acid sequence conserved across species as previously described (STC, Prof. A. Sobel, INSERM U.440, Paris, France) and 1:1,000-diluted rabbit anti-stathmin antiserum against an amino acid sequence conserved across species as previously described (STC, Prof. A. Sobel, INSERM U.440, Paris, France).

**Table 1. List of antibodies used**

<table>
<thead>
<tr>
<th>Antibody</th>
<th>Ref. No.</th>
<th>Origin</th>
<th>Isotype</th>
<th>Specificity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mouse anti-human CD106</td>
<td>MCA907B</td>
<td>Serotec products (Oxford, UK)</td>
<td>IgG1</td>
<td>VCAM-I</td>
</tr>
<tr>
<td>Mouse anti-porcine MHC class II</td>
<td>MCA1335</td>
<td>Serotec products</td>
<td>IgG2b</td>
<td>MHC class II</td>
</tr>
<tr>
<td>Mouse anti-porcine macrophage and monocyte</td>
<td>MCA1218</td>
<td>Serotec products</td>
<td>IgG2b</td>
<td>Monocytes/macrophages</td>
</tr>
<tr>
<td>Mouse anti-porcine CD4⁵</td>
<td>MCA1749</td>
<td>Serotec products</td>
<td>IgG2b</td>
<td>T helper lymphocytes</td>
</tr>
<tr>
<td>Mouse anti-porcine CD8⁴</td>
<td>MCA1223</td>
<td>Serotec products</td>
<td>IgG2a</td>
<td>T cytotoxic-suppressor lymphocytes</td>
</tr>
</tbody>
</table>

The anti-human antibody used cross-reacted with the pig species.

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metric analysis of the immunoreactive protein bands was performed using OptiQuant image-analysis software (Packard BioScience, Meriden, CT).

**Histopathological studies.** Biopsies of tissue samples from the deep cortex-outer medulla region were collected at days 1, 3, 5, 7, and 14, and at 4–5, 10–12, and 16 wk after every WI followed by the reperfusion process. Conventional stains were applied (hematoxylin and eosin, periodic acid-Schiff). Different histological parameters (tubular necrosis, brush-border integrity, and intratubular detachment) were studied. Histological lesions were expressed in a percentage of kidney samples using a previously described semiquantitative scale (13): 0, no abnormality; 1, mild lesions affecting <10% of kidney samples; 2, lesions affecting 10–25% of kidney samples; 3, lesions affecting 25–50% of kidney samples; 4, lesions affecting 50–75%; and 5, lesions affecting >75% of kidney samples. Mitochondrial integrity was also measured using electromicroscopy. Tubulointerstitial injury was defined as inflammatory cell infiltrates, tubular atrophy, or interstitial fibrosis. To estimate the level of tubulointerstitial fibrosis, tissue sections were also labeled with Picro Sirius for collagen identification (collagen I and III) as recommended (19). The amount of interstitial fibrosis was determined in Picro Sirius-stained sections by a semiquantitative imaging technique. Image analysis was performed by either a technician or a pathologist blinded to the clinical source of the sample. The slides were examined with a microscope, and the images were acquired using a ×40 objective. The percentage of Picro Sirius-stained surface was determined by a computer-aided image-analysis program. In each representative slide, staining in renal tissue was semiautomatically quantified in 15 fields by the computer program in each experimental condition and expressed as percentage of the total surface area examined. Tubular atrophy was semiquantitatively scored on a scale of 0–4 by two pathologists blinded to the experimental conditions (0, normal; 1, small changes affecting 5–10%; 2, changes affecting 10–25% of specimen area; 3, changes affecting 25–50% of specimen area; 4, changes affect-

![Image of Fig. 1. Influence of trimetazidine (TMZ) on warm ischemia (WI) time on creatinine clearance (A) and protein excretion (B). Renal function was measured in the different experimental groups after different WI times and compared with control group and uninephrectomized group. Values are means ± SE. *P < 0.05, **P < 0.01 WI-45 vs. WI-45-TMZ. *P < 0.05; **P < 0.01 WI-60 vs. WI-60-TMZ. *P < 0.05 WI-90 vs. WI-90-TMZ.](http://ajprenal.physiology.org/)

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ing 50–75% of specimen area; and 5, changes affecting 75–100% of specimen area).

Immunohistochemical studies. Frozen and paraffin-embedded kidney biopsy sections (5 μm) were processed for indirect immunohistochemistry using several mouse monoclonal antibodies (mAbs; dilution 1:20) as previously described (13) by using the antibodies listed in Table 1. Indirect immunofluorescence was performed by using an anti-human VCAM-1 antibody (see Table 1). All sections were stained with anti-CD8, CD8-APC, CD4, MHC class II, and VCAM-1. The number of MCA1218-, CD4+-, CD8+-, and MHC class II and VCAM-1 staining was semiquantitatively estimated.

Indirect immunofluorescence was performed by using an anti-human VCAM-1 antibody (see Table 1). All sections were stained with anti-CD8, CD8-APC, CD4, MHC class II, and VCAM-1. The number of MCA1218-, CD4+-, CD8+-, and MHC class II and VCAM-1 staining was semiquantitatively estimated.

Anti-CD8, anti-CD8-APC, anti-CD4, anti-MHC class II, and anti-VCAM-1 staining was semiquantitatively estimated.

Evolution of cellular integrity of postreperfused kidneys after varying intervals of WI with and without TMZ. Histological analysis of biopsy samples from WI kidneys revealed different differences in the cellular integrity after 40 min of reperfusion and at day 7 postreperfusion. TMZ treatment in WI-30 groups exhibited a very low level of difference (data not shown). Kidneys from the WI-45 group exhibited reduced damage compared with WI-60 and particularly WI-90 groups (Table 2). The degree of initial acute proximal tubule cell injury appeared closely related to the duration of WI, particularly after 60 min of WI (Table 2). The WI-60 and WI-90 groups exhibited an intense proximal tubule necrosis with a significant loss of brush border and proteinaceous cast. TMZ reduced the injury score after WI-45, and particularly after 60 and 90 min of WI. The protective effect of TMZ was more pronounced on brush-border integrity and mitochondria (Table 2 and Fig. 2). Interestingly, TMZ-pretreated groups presented a more reduced injury score at day 7 compared with WI groups particularly after 60 and 90 min of WI, which is related to the repair process.

Effect of varying intervals of renal ischemia with and without TMZ on stathmin mRNA and stathmin and HIF-1α protein expression in pig kidneys. Our results indicate that the expression of stathmin mRNA increases in all IRI samples and that the highest increase was found in the RNA extracted from kidneys subjected to TMZ treatment (Fig. 3) and in pig kidneys treated with TMZ after 3-h reperfusion. At day 7, stathmin mRNA was significantly increased in groups without TMZ compared with TMZ-treated groups, suggesting that the process of tubular repair and recovery after acute renal injury was not delayed in TMZ-treated groups. Stathmin protein increased and reached its peak levels by 72 h after the induction of ischemia. Stathmin expression was increased in TMZ-perfused pig kidneys at day 1 (Fig. 4A). At day 7, stathmin expression decreased in experimental groups with TMZ (particularly WI-45-TMZ and WI-60-TMZ groups) compared with nontreated groups (Fig. 4, A and B, bottom). HIF-1α was not detected in the WI-60 and particularly in WI-90 groups. When TMZ was administered before WI, proteinuria was reduced for WI of 45, 60, and 90 min, WI-60-TMZ, and WI-90-TMZ. As assessed in the two tests (Cr and proteinuria), TMZ (5 mg/kg iv) administered 24 h before WI was always effective, but their improvements were only partial.

Table 2. Degree of proximal tubule cell lesions and mitochondrial damage after different periods of warm ischemia

<table>
<thead>
<tr>
<th>Injury Type/Reperfusion Time</th>
<th>WI-45-TMZ/WI-45</th>
<th>WI-60-TMZ/WI-60</th>
<th>WI-90-TMZ/WI-90</th>
</tr>
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<tbody>
<tr>
<td>Brush-border integrity</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>30–45 min</td>
<td>3.6±0.2/3.4±0.2</td>
<td>3.0±0.3/2.7±0.2</td>
<td>2.5±0.2/2.2±0.2</td>
</tr>
<tr>
<td>day 7</td>
<td>4.2±0.2/3.6±0.2*</td>
<td>3.7±0.2/3.1±0.1†</td>
<td>3.3±0.2/2.5±0.2‡</td>
</tr>
<tr>
<td>Tubular necrosis</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>30–45 min</td>
<td>2.4±0.2/2.7±0.3</td>
<td>2.9±0.3/3.2±0.2</td>
<td>3.5±0.2/3.8±0.2</td>
</tr>
<tr>
<td>day 7</td>
<td>1.7±0.1/1.6±0.2*</td>
<td>2.0±0.2/2.5±0.2‡</td>
<td>2.3±0.2/3.0±0.2‡</td>
</tr>
<tr>
<td>Mitochondrial injury</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>30–45 min</td>
<td>2.1±0.2/2.4±0.2</td>
<td>2.7±0.2/3.5±0.2</td>
<td>3.6±0.3/4.1±0.3</td>
</tr>
<tr>
<td>day 7</td>
<td>1.2±0.2/1.8±0.2*</td>
<td>1.7±0.2/3.3±0.2‡</td>
<td>2.5±0.2/3.2±0.2‡</td>
</tr>
</tbody>
</table>

Values are means ± SE for all parameters analyzed. The histological lesions from early (30–40 min) and late (45–90 min) reperfusion. TMZ treatment in WI-30 groups exhibited an intense proximal tubule necrosis with a significant loss of brush border and proteinaceous cast. TMZ reduced the injury score after WI-45, and particularly after 60 and 90 min of WI. The protective effect of TMZ was more pronounced on brush-border integrity and mitochondria (Table 2 and Fig. 2). Interestingly, TMZ-pretreated groups presented a more reduced injury score at day 7 compared with WI groups particularly after 60 and 90 min of WI, which is related to the repair process.

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detected at day 0 (basal level). The HIF-1α increase occurred within 3 h after reperfusion in TMZ-treated groups and in the WI-45 group and peaked at day 7. In turn, the HIF-1α protein increase was delayed in groups without TMZ pretreatment (Fig. 4B, bottom). This data support indicated that the process of tubular repair and recovery after acute renal injury was delayed in non-TMZ-treated groups. In addition, the production of HIF-1α, which was correlated with the excretion of cytoprotective factors such as VEGF or erythropoietin, was delayed in non-TMZ-treated kidneys.

Effect of varying intervals of renal ischemia with and without TMZ on MHC class II expression in pig kidneys.

The expression of MHC class II molecules was analyzed after WI (Table 3 and Fig. 5). The topology of immunostaining mainly involved the peritubular capillary endothelial cells and surrounding inflammatory cells. The WI-60 and WI-90 groups exhibited a more diffuse and intense immunolabeling in these cells. Proximal tubule, thick ascending limb of loop of Henle, and collecting duct epithelial cells were always negative. Rarefied MHC class II vascular endothelial cells were observed in the W45 groups. TMZ reduced MHC class II expression in all WI groups and particularly in the WI-60 and WI-90 groups.

Effect of WI on cellular infiltration and VCAM-1 expression in pig kidneys after various WI periods with and without TMZ.

In accordance with previous studies, the major inflammatory cell population detected by immunohistochemical studies was CD4+ T lymphocytes. No infiltrating cell was detected in control, UNX, and WI-10 (data not shown). Almost no infiltrating CD4+ and CD8+ cells were detected in the remaining kidney from the W130 group (data not shown). In contrast, the

Fig. 2. Top and bottom: effect of WI on brush-border and mitochondrial integrity, respectively, in postreperfused kidneys. WI-45 and WI-TMZ-45, WI-60 and WI-TMZ-60, and WI-90 and WI-TMZ-90 (45, 60, and 90 min of WI, respectively) kidney samples are presented after reperfusion and compared with control and uninephrectomized groups. bb, Brush border; tl, tubular lumen. Brush-border integrity and mitochondrial density in different groups are represented by arrows. mt, Mitochondria. Mitochondria density was reduced inWI-60 and particularly WI-90. Biopsy samples from WI-30 are not presented because the absence of damage is sufficiently remarkable. Middle: effect of WI on tissue integrity in postreperfused kidneys. WI-45 and WI-TMZ-45, WI-60 and WI-TMZ-60, and WI-90 and WI-TMZ-90 kidney samples are presented after reperfusion. WI-30 biopsy samples are not presented because the absence of damage is sufficiently remarkable. Brush-border preservation is represented by arrows.

Fig. 3. Expression of stathmin mRNA after various durations of ischemia followed by reperfusion in cortex and outer medulla. The stathmin mRNAs were standardized against 18S rRNA and expressed as ratios of the value of standardized stathmin in sham-operated kidneys as described in METHODS. Values are means ± SE from 3 independent experiments. *P < 0.05, **P < 0.01 vs. sham-operated and control groups (0 h).
Fig. 4. Western blot analysis of stathmin and hypoxia-inducible factor-1 (HIF-1α). A and B: expression of stathmin and HIF-1α, respectively, was examined by Western blotting in cortex and outer medulla at days 0, 3, and 7 after reperfusion. C: representative immunoblots for stathmin, HIF-1α, and control actin after various durations of ischemia (45, 60, and 90 min) followed by reperfusion and stathmin and HIF-1α expression in normal kidney (control) and uninephrectomized animals. Induction of stathmin and HIF-1α was observed at 72 h of reperfusion, particularly in the outer medulla in TMZ-treated groups, and this induction was delayed in no TMZ-treated groups. Values are means ± SE from 3 independent experiments. *P < 0.05 TMZ-treated groups vs. no TMZ-treated group and sham-operated and uninephrectomized groups (0 h). **P < 0.01 TMZ-treated groups vs. no TMZ-treated group and sham-operated and control groups (0 h).
number of CD4\(^+\) cells increased proportionally to the WI periods for 4 wk after reperfusion in the WI-45 and particularly WI-60 and WI-90 groups (Fig. 6A). After 4 wk, CD4\(^+\) cells slowed to a plateau up to 16 wk following reperfusion in the WI-45 group and slightly increased in the WI-60 and WI-90 groups. The number of CD8\(^+\) cells increased also with the WI time, particularly in the WI-60 and WI-90 groups (Fig. 6B). However, the number of CD8\(^+\) cells was much lower and gradually increased with longer WI periods. The number of macrophages/monocytes increased from day 1 to days 5–7 according to WI time and decreased between day 7 and week 2, particularly for the WI-45 group. In the WI-60 and WI-90 groups, monocytes/macrophages increased between day 1 and day 7, then slightly decreased between day 7 and day 14, increased again, and then remained unchanged after 10-wk follow-up (Fig. 6C). Unlike cold-ischemic injury, the biphasic process of inflammatory cell infiltration in interstitial areas was less prominent. TMZ reduced the inflammatory response in all groups and particularly in the WI-60 and WI-90 groups. VCAM-1 demonstrated a mild staining in tubulointerstitial areas in the WI-45 group. VCAM-1 expression increased with WI times in the WI-60 and WI-90 groups.

<table>
<thead>
<tr>
<th>Table 3. Semiquantitative analysis of MHC class II expression</th>
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<tr>
<td><strong>Week 1</strong></td>
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<tr>
<td>WI-45/WI-45-TMZ</td>
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<tr>
<td>Endothelial cells</td>
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<tr>
<td>Tubular cells</td>
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<tr>
<td>Inflammatory cells</td>
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<td>Tubular cells</td>
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<td>Inflammatory cells</td>
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Semiquantitative graded score of the immunostaining was used (0, no staining to 4+, dense). For details, see METHODS.

Fig. 5. Effect of WI time on MHC class II distribution in postreperfused kidneys. Biopsy samples were processed for immunohistochemistry using MCA 1335 anti-porcine MHC class II-DQ. The most intense staining (arrows) was noted in WI-45, WI-60, and WI-90 (left) compared with WI-TMZ-45, WI-TMZ-60, and WI-TMZ-90 (right). Semiquantitative graded score of immunostaining was used (0: no staining to 4+: dense).
exhibited a more intense VCAM-1 staining in tubular, peritubular capillary, and inflammatory cells (Fig. 7 and Table 4). TMZ-treated groups exhibited a reduced expression of VCAM-1 during the 18-wk follow-up.

**Effect of WI on the onset of interstitial fibrosis after various WI periods with and without TMZ.** The evolution of interstitial fibrosis is presented in Fig. 8A and B1 to B6. In our hands, fibrosis was correlated with the duration of WI, particularly
after 60 min and 90 min of WI (Fig. 7, A and C). The percentage of interstitial fibrosis area was also related to the progression of CCR as observed to the WI-45 group compared with WI-60 and WI-90 groups. These changes were accompanied by an atrophy of the surrounding tubules and significant interstitial fibrosis. The lesions in WI-60 and WI-90 groups exhibited a focal-radial character and are characterized by patchy destruction of renal structures (tubuli, glomeruli and vessels). Within kidneys from TMZ-treated groups, fibrosis development was reduced (Fig. 8 A to B).

**DISCUSSION**

ARF caused by IRI is among the major causes of morbidity in hospitalized patients and indicates poor prognosis in patients with multisystem organ failure. The reduction in regional blood flow to the outer medulla occurring during IRI is more pronounced than that to the cortical capillaries in the postischemic kidney (36). In the present study, we have demonstrated a strong correlation between CCr evolution and WI times as previously suggested (26). TMZ improved CCr in all groups, particularly in WI-90-TMZ 2 wk after reperfusion. These functional data were associated with the improvement of mitochondrial function itself. This was clearly observed particularly at day 7 compared with the one observed after 40 min of reperfusion. Recently, it was also demonstrated that TMZ improved mitochondrial functions of the myocardium subjected to several I/R in an experimental canine model (25). The mitochondrial morphological changes to “rod-like” seen in the TMZ-treated group indicate that mitochondria require a larger membrane surface area to produce ATP. In addition, TMZ in this model also increased mitochondrial turnover. Recently, TMZ was suggested to inhibit mitochondrial permeability transition pores and prevent lethal IRI after coronary artery occlusion (1). These data demonstrate that TMZ could limit apoptotic cardiomyocyte death and support recent findings by Yin et al. (40), who showed a similar antiapoptotic effect in cultured astrocytes subjected to hypoxia-reoxygenation. Collectively, these data suggest that TMZ protects mitochondria from IRI.

As to the effect of TMZ on renal regeneration, this drug improved the repair process level, as supported by its effect on stathmin expression. Stathmin, an 18-kDa protein, is expressed in cells that have reentered the cell cycle (30). Stathmin expression was found to be positively correlated with cellular proliferation in different cell lines (31). Interestingly, stathmin expression was shown to decrease on induction of differentiation and cessation of proliferation in undifferentiated cell lines. It regulates microtubule dynamics by promoting depolymerization of microtubules and/or preventing polymerization of tubulin heterodimers and can be regulated by phosphorylation. Recent studies have demonstrated that stathmin can have two distinct activities. Stathmin is capable of binding both polymerized and unpolymerized tubulin and can prevent polymerization of heterodimers under some conditions and/or promote depolymerization of microtubules under others (for a review, see Ref. 31). Previous studies have demonstrated that the expression of stathmin increases in the proximal tubules of kidneys subjected to IRI (44). Stathmin was expressed by dedifferentiated and actively proliferating proximal tubule cells in the corticomedullary junction and in the S3 segment of the proximal tubule (39). Recently, it was suggested that in stathmin-deficient mice subjected to IRI (44). Stathmin was expressed by dedifferentiated and actively proliferating proximal tubule cells in the corticomedullary junction and in the S3 segment of the proximal tubule (39). Recently, it was suggested that in stathmin-deficient mice subjected to IRI, the aberrant regulation of cell cycle progression impairs or at least delays the process of tubular repair and recovery after acute renal injury (45). In our study, the protective effect of TMZ in renal tissue of pretreated groups was correlated with an earlier and higher expression of stathmin than in renal tissue without TMZ treatment. This effect on stathmin, which is involved in the regulation of mitosis and proposed to be a relay integrating diverse intracellular signaling pathways controlling cell proliferation, differentiation, and functions, could be related to a recently suggested effect against dephosphorylation of nucleotides during ischemia (8). In addition, these results suggest that stathmin is a critical protein in the regulation of cell mitosis in damaged
Limitation of fibrosis was correlated with increases in CCr and delaying fibrosis development within tubulointerstitial tissue. On the other hand, the persistence of stathmin expression within kidneys from untreated animals could be related to prolonged cell stress. Such results are in accordance with previous data which have demonstrated that stathmin mRNA and protein reached a peak at 24-h posthepatectomy, then decreased until 96 h after hepatectomy (20). Taken together, these results reveal an innovative aspect of the protective role of TMZ in regulatory pathways of cell proliferation and differentiation during regeneration.

Regarding the effect of TMZ on HIF-1α, its expression was delayed in nontreated groups particularly after 90 min of WI. As a global regulator of hypoxic adaptation, HIF is involved in the regulation of mitochondrial signaling, hypoxic cell death, and recovery from IRI. HIF-1α is known to upregulate factors that have been shown to be cytoprotective. TMZ is involved in an earlier and more prolonged HIF-1α expression correlated with limitation of damage. The response of kidneys acutely damaged by ischemia or toxins is dominated by epithelial destruction and regeneration. Recent data have suggested that a reduction in the peritubular capillary density after ARF results in persistent reduction of renal PO2 and that hypoxia may play an important role in the progression of chronic disease after ARF (6). In our model, the influence on renal recovery with a protective drug such as TMZ was efficient after ARF (6).

The other important finding was the impact of a single injection of TMZ 24 h before WI on inflammation processes. Recent years have seen the emergence of the role of inflammation in IRI. Leukocyte adhesion/infiltration and endothelial cell activation in response to IRI is a well-known phenomenon (3, 17, 41). In the current study, we have studied the late phase of inflammation. TMZ modulated monocyte/macrophage infiltration, which served at this late phase as a crucial interface between the innate and adaptive immune systems. T lymphocytes were also reduced by TMZ. Evidence from different laboratories supports a role for T cells as modulators of renal IRI, particularly in rodent models (for a review, see Ref. 5). Using specific antibodies for inflammatory cells, the limitation of WI’s deleterious effects by TMZ was correlated with the limitation of inflammation process after 45, 60, and particularly 90 min of WI. This effect could be related to an effect of TMZ on the regulation of mitochondrial signaling, hypoxic cell death, and recovery from IRI. HIF-1α is known to upregulate factors that have been shown to be cytoprotective. TMZ is involved in a dynamic process involving vascular network and tubules in a complex interaction. Our data support previous findings on the counteracting role of TMZ in fibrosis development in different diseases. In a rat model of alkali burns of the esophagus, TMZ reduced the degree of fibrosis and limited histopathological damage (2). Given that a single episode of renal IRI is associated with increased chronic allograft nephropathy, which decreased long-term allograft nephropathic function, TMZ could be useful in the limitation of renal damage.

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Table 4. Semiquantitative analysis of VCAM-1 expression

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<tr>
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<th>Week 1</th>
<th>Week 2</th>
<th>Weeks 4–6</th>
<th>Weeks 8–12</th>
<th>Week 16</th>
</tr>
</thead>
<tbody>
<tr>
<td>WI-45/WI-45-TMZ</td>
<td>0 to +/0</td>
<td>+/0 to +</td>
<td>+/0 to +</td>
<td>+/0 to +</td>
<td>+/0 to +</td>
</tr>
<tr>
<td>WI-60/WI-60-TMZ</td>
<td>+/0 to +</td>
<td>+/0 to +</td>
<td>+/+</td>
<td>+/+</td>
<td>+/+</td>
</tr>
<tr>
<td>WI-90/WI-90-TMZ</td>
<td>+/+</td>
<td>+/+</td>
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Semiquantitative graded score of immunostaining was used (0, no staining to 4+, dense). For details, see METHODS.

Fig. 8. Influence of WI on the onset of tubulointerstitial fibrosis. A: percentage of kidney areas displaying interstitial fibrosis stained with Picro Sirius was measured over several weeks following WI within kidneys between W8 and W16. Bottom: tubulointerstitial fibrosis determined using Picro Sirius red coloration at W16. Values are means ± SE. *P < 0.05, **P < 0.01 WI-45 vs. WI-45-TMZ. °P < 0.05. °°P < 0.01 WI-60 vs. WI-60-TMZ. *P < 0.05 WI-90 vs. WI-90-TMZ.
on IRI. In addition, ROS, generated during IRI, can also activate T cells (5, 42), and the role of TMZ against ROS production and mitochondrial damage could indirectly explain this effect (24, 38). Otherwise, TMZ reduced VCAM-1 expression, which could also participate in the limitation of inflammation via endothelial cells. VCAM-1 is transcriptionally induced on endothelial cells, and its engagement induces a signal in the endothelial cells that trigger changes in shape and allow leukocyte emigration. TMZ treatment influenced MHC class II expression. The expression of MHC class II antigens by the injured tubular epithelium, picked up by and presented by recruited dendritic cells to CD4+ T cells, represents the re-active cellular immunity important in the discrimination between self and nonself (7). These data indicated that when ischemic damage was severe, there was a progressive loss of tolerance to parenchymal self, inducing a persistent immune response (14, 29). This effect paralleled the fibrosis development within damaged tissue and indicated that the protective effect of TMZ is a global effect, which improved the regeneration of tissue and limitation of the long-term deleterious effect of IRI.

Consequently, the interest of TMZ in this model was supported by its role in the early and late phases of IRI. We have previously demonstrated the positive effect of TMZ in cold preservation (14–16). This effect was also remarkable in the WI pig kidney model, suggesting that TMZ could be useful in clinical transplantation where the permanent shortage of organs had led to attempts to expand the donor pool. In addition, our model allows designing a study with a long-term follow-up compared with most experimental studies, which focused on a short-term effect in the first days following an ischemic insult. TMZ could be particularly efficient in “controlled donors” (type III and IV from the Maastricht group) as an early single perfusion could limit fibrosis development (4, 16). It would be useful to address the best regimen of TMZ to be used and to check whether sequential administration to the recipient of the graft could restore renal function.

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