Novel sandwich ELISA for human angiotensinogen

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Katsurada A, Hagiwara Y, Miyashita K, Satou R, Miyata K, Ohashi N, Navar LG, Kobori H. Novel sandwich ELISA for human angiotensinogen. Am J Physiol Renal Physiol 293: F956–F960, 2007.—We recently reported that urinary excretion rates of angiotensinogen (UAGT) provide a specific index of intrarenal renin-angiotensin (ANG) system (RAS) status in ANG II-dependent hypertensive rats. When this is shown to be applicable to human subjects, a diagnostic test to identify those hypertensive patients most likely to respond to an RAS blockade could provide useful information to allow a mechanistic rationale for selection of an optimized approach to treatment of hypertensive patients. However, simple and accurate methods to measure human angiotensinogen (hAGT) are unavailable. For future studies of human subjects, we developed antibodies and a sensitive and specific quantification system for hAGT using a sandwich ELISA. We raised two antibodies against hAGT: a mouse monoclonal antibody and a rabbit polyclonal antibody. The standard curve of this ELISA exhibited a high linearity (0.31–20 ng/ml). The correlation coefficient was >0.99. Plasma angiotensinogen concentrations of healthy volunteers ranged from 28 to 71 μg/ml (n = 10). The ratio of UAGT to urinary creatinine concentration ranged from 5.0 to 30 μg/g (n = 7). Intra- and interassay coefficients of variation ranged from 4.4 to 5.5% and from 4.3 to 7.0%, respectively. This ELISA system had no cross-reactivity with major proteins in proteinuric urine samples, such as human albumin, immunoglobulin, or transferrin. Moreover, the cross-reactivity of the system with angiotensin peptides was also negligible. This hAGT ELISA will be a useful tool to investigate the relationship of UAGT and reactivity to antihypertensive drugs in hypertensive patients.

MATERIALS AND METHODS

Protocol. The experimental protocol of this study was approved by the Institutional Review Board of Tulane University. All samples were obtained from healthy volunteers, who provided written informed consent.

Preparation of expression vectors for human angiotensinogen. The full length, except for the signal peptide (1-33 aa), of the human angiotensinogen (34-485 aa) gene (gene identification no. 73622269) was amplified by PCR using a high-fidelity Pyrococcus furiosus DNA polymerase (Promega) with sense (5’-CGG GAT CCG ACC GGG TGT ACA TAC ACC CC-3’) and antisense (5’-CGG TAT CCG ACC GGG GAC TCG TGT CCG CAC ACC CC-3’) primers from the human adult liver cDNA library (Clontech). Then this fragment was inserted into pGEX4T1 expression vectors (Promega) with the glutathione S-transferase (GST) tag at the 5’-end by the restriction enzymes BamH I and Sal I.

Preparation of recombinant proteins for human angiotensinogen. The recombinant constructs were transformed to a high-efficiency expression bacterial strain (Takara). Large-scale bacterial cultures were induced with isopropyl-β-D-thiogalactopyranoside (Takara) and harvested for protein purification. GST-tagged proteins were purified using glutathione beads (Upstate) in native conditions.

Antibody preparation. We raised two antibodies for human angiotensinogen: a mouse monoclonal and a rabbit polyclonal antibody.

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The monoclonal antibody was raised in mouse against recombinant protein of human angiotensinogen (see above). The polyclonal antibody was raised in rabbit against synthetic oligopeptide corresponding to human angiotensinogen (72-89 aa). Both antibodies were affinity purified.

Western blot. Western blot analysis was performed as previously described (12, 20, 22) using the LI-COR Odyssey infrared imaging system.

Epitope mapping. The epitopes of these antibodies were determined by Western blot analysis using a variety of lengths of recombinant proteins of human angiotensinogen (34-124, 34-223, 34-303, 34-393, and 34-485 aa).

Plate preparation. The ELISA plates were coated with the polyclonal antibody against human angiotensinogen (100 μl/well in 100 mmol/l carbonate buffer, pH 9.5) at 4°C overnight. The plates were stored at 2–8°C.

Sample collections. Peripheral blood samples from healthy volunteers were collected into EDTA-containing tubes, and plasma samples were separated after centrifugation. First-morning-urine samples were also collected from healthy volunteers.

Development of sandwich ELISA. Highly purified angiotensinogen from human plasma was used as the standard. Human angiotensinogen standards (0.31–20 ng/ml diluted in ELISA buffer), plasma (1:8,000 dilution in ELISA buffer), and urine (1:8 dilution in ELISA buffer) samples (100 μl/well) were added to each well of the plates and incubated at 37°C for 1 h. Then the plates were washed a total of seven times with a washing buffer (PBS containing 0.05% Tween 20,
pH 7.5). After the plates were incubated with horseradish peroxidase-labeled monoclonal antibody against human angiotensinogen (100 μl/well, 1:30 dilution in antibody solution) at 37°C for 30 min, they were washed a total of nine times with the washing buffer. Then the plates were incubated with 3,3'-5,5'-tetramethylbenzidine solution (100 μl/well) under light-protected conditions at room temperature for 30 min. The reaction was stopped by treatment of the plates with sulfuric acid (100 μl/well, 0.5 mol/l). The absorbance values were measured at 450 nm.

Measurement of urinary creatinine. The urinary creatinine concentrations were measured by an automated machine (model DCA 2000+, Bayer) with microalbumin/creatinine reagent kits (Bayer). The urinary angiotensinogen concentrations were normalized to urinary creatinine concentrations.

Statistical analysis. Statistical analysis was performed using a one-way factorial ANOVA with post hoc Scheffe’s F test. Values are means ± SE. P < 0.05 was considered significant.

RESULTS AND DISCUSSION

Epitope mapping of antibodies. An equal amount (10 ng) of various lengths of recombinant proteins of human angiotensinogen (34-124, 34-223, 34-303, 34-393, and 34-485 aa) was blotted on nitrocellulose membranes, and epitope mapping was done by Western blot. The polyclonal antibody against human angiotensinogen recognized GST-tagged recombinant proteins of human angiotensinogen (34-124, 34-223, 34-303, 34-393, and 34-485 aa), but not mock protein or GST protein (Fig. 1A). The monoclonal antibody against human angiotensinogen recognized GST-tagged recombinant proteins of human angiotensinogen (34-485 aa), but not GST-tagged recombinant proteins of human angiotensinogen (34-124, 34-223, 34-303, and 34-393 aa), mock protein, or GST protein. An antibody against GST protein recognized GST-tagged recombinant proteins of human angiotensinogen (34-124, 34-223, 34-303, 34-393, and 34-485 aa), as well as GST protein, but not mock protein. These data clearly indicate that the epitope for the polyclonal antibody against human angiotensinogen is located on the NH2 terminus of human angiotensinogen (34-124 aa) and that the epitope for the monoclonal antibody against human angiotensinogen is on the COOH terminus of human angiotensinogen (394-485 aa).

Plate stability. To address their stability, we incubated the polyclonal antibody-coated ELISA plates at 37°C for 1, 8, or
15 days. Then different concentrations of human angiotensino-
gen standards were applied to the plates and processed, and the absorbance values were obtained. Each concentration of hu-
gen standards exhibited similar absorbance values beyond 15 days (Fig. 1B). These data clearly indicate that the polyclonal antibody-coated ELISA plates are stable ≥15 days after antibody coating.

Standard curves. The corresponding standard curve exhibited a high linearity (0.31–20 ng/ml), with absorbance values ranging from 0.03 to 2.34 (n = 6 at each concentration; Fig. 1C). The correlation coefficient was >0.99.

Detection limit. The lowest sensitivity for this system (0.31
/ng/ml) was determined using the guidelines of the National
Committee for Clinical Laboratory Standards Evaluation Pro-
tocols.

Plasma and urinary angiotensinogen concentrations. An-
giotensinogen concentrations in plasma obtained from 10
healthy volunteers ranged from 28 to 71 μg/ml (Fig. 2A). Angiotensinogen concentrations in urine obtained from seven healthy volunteers ranged from 7.1 to 35 ng/ml (Fig. 2B). Urinary creatinine concentrations of seven healthy volunteers were 0.93–2.2 mg/l. The ratio of urinary angiotensinogen concentration to urinary creatinine concentration was 5.0–30 μg/g (Fig. 2C).

Addition and recovery test. Different known concentrations of human angiotensinogen standards were added to a human plasma sample or a human urine sample, and the added angiotensinogen concentrations were calculated as the difference between angiotensinogen concentrations in samples with and without added angiotensinogen. Recovery rates were >82.9% for plasma and >87.6% for urine (Table 1).

Sample stability. To address stability of samples, we performed two tests: a freeze-and-thaw test and an accelerated test. In a freeze-and-thaw test, one plasma sample was frozen and thawed 0, 1, 2, and 3 times, and plasma angiotensinogen concentrations were measured by the ELISA system. No sig-
nificant differences were observed (Fig. 3A). In an accelerated test, one plasma sample was incubated at 37°C for 0, 1, 3, and 7 days, and plasma angiotensinogen concentrations were mea-
sured by the ELISA system. No differences were observed (Fig. 3B). These data clearly indicate that angiotensinogen in plasma samples is stable without preservatives or protease inhibitors, except EDTA.

Intra-assay test. Intra-assay tests were performed using three different concentrations of human angiotensinogen standards. Coefficients of variation among the wells ranged from 4.4 to 5.5% (Table 2).

Interassay test. Interassay tests were performed using three different concentrations of human angiotensinogen standards. Coefficients of variation among the plates ranged from 4.3 to 7.0% (Table 3).

Specificity. To confirm cross-reactivity with other proteins, several human proteins (20 μg/ml) were applied to the ELISA system and processed. Cross-reactivity for other human pro-
teins in this ELISA system was negligible (Table 4). In addition, downstream products of angiotensinogen (20 μg/ml) were also applied to the ELISA system. The cross-reactivity for angiotensin peptides in this ELISA system was also negligible (Table 4).

Effect of pH of test samples on sensitivity of ELISA. As described above, plasma samples were diluted 1:8,000 and urine samples 1:8 in PBS-based dilution buffer (pH 7.0); therefore, pH of diluted plasma and urine samples should be ≥7.0. To address this issue, seven urine samples were prepared (pH 5.5–7.0, 6.3 ± 0.6). After the samples were diluted with the dilution buffer, pH values of the samples were integrated (pH 6.7–7.0, 6.8 ± 0.2). Moreover, diluted urine samples worked well with ELISA (15.9 ± 6.4 ng/ml). These data clearly indicate that the change of urine sample pH does not affect the sensitivity of ELISA.

Conclusion. We developed antibodies and a sensitive and specific quantification system for human angiotensinogen using a sandwich ELISA. This human angiotensinogen ELISA will be a useful tool to investigate the relationship between urinary angiotensinogen excretion rate and reactivity to anti-
hypertensive drugs in hypertensive subjects.

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GRANTS

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REFERENCES

1. Baltatu O, Silva JA Jr, Ganten D, Bader M. The brain renin-angiotensin system modulates angiotensin II-induced hypertension and cardiac hyper-


4. Dell’Italia LJ, Meng QC, Balecels E, Wei CC, Palmer R, Hageman GR, Durand J, Hanks GH, Oparil S. Compartmentalization of angio-
tensin II generation in the dog heart: evidence for independent mecha-

Table 4. Specificity

<table>
<thead>
<tr>
<th>Human Proteins</th>
<th>Cross-Reactivity, %</th>
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<tr>
<td>Human angiotensinogen</td>
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<td>Human tenasin C</td>
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<tr>
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<tr>
<td>Human growth-regulated oncogene-β</td>
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