Chloroquine and inhibition of Toll-like receptor 9 protect from sepsis-induced acute kidney injury

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Yasuda H, Leelahavanichkul A, Tsunoda S, Dear JW, Takahashi Y, Ito S, Hu X, Zhou H, Doi K, Childs R, Klinman DM, Yuen PS, Star RA. Chloroquine and inhibition of Toll-like receptor 9 protect from sepsis-induced acute kidney injury. Am J Physiol Renal Physiol 294: F1050–F1058, 2008. First published February 27, 2008; doi:10.1152/ajprenal.00461.2007.—Mortality from sepsis has remained high despite recent advances in supportive and targeted therapies. Toll-like receptors (TLRs) sense bacterial products and stimulate pathogenic innate immune responses. Mice deficient in the common adapter protein MyD88, downstream from most TLRs, have reduced mortality and acute kidney injury (AKI) from polymicrobial sepsis. However, the identity of the TLR(s) responsible for the host response to polymicrobial sepsis is unknown. Here, we show that chloroquine, an inhibitor of endocytic TLRs (TLR3, 7, 8, 9), improves sepsis-induced mortality and AKI in a clinically relevant polymicrobial sepsis mouse model, even when administered 6 h after the septic insult. Chloroquine administration attenuated the decline in renal function, splenic apoptosis, serum markers of damage to other organs, and prototypical serum pro- and anti-inflammatory cytokines TNF-α and IL-10. An oligodeoxynucleotide inhibitor (H154) of TLR9 and TLR9-deficient mice mirror the actions of chloroquine in all functional parameters that we tested. In addition, chloroquine decreased TLR9 protein abundance in spleen, further suggesting that TLR9 signaling may be a major target for the protective actions of chloroquine. Our findings indicate that chloroquine improves survival by inhibiting multiple pathways leading to polymicrobial sepsis and that chloroquine and TLR9 inhibitors represent viable broad-spectrum and targeted therapeutic strategies, respectively, that are promising candidates for further clinical development.

multiple organ failure; innate immune system; bacterial products; cytokines

DESPITE ADVANCES IN SUPPORTIVE care and specific therapeutic interventions, the mortality rate of sepsis is still unacceptably high. Death frequently follows the development of multiple organ failure (42, 47). Invasion of bacteria elicits a complex interplay of local and systemic responses that seek to control the infection. The innate immune system recognizes the presence of invading bacteria and bacterial products and initiates a local host response by releasing cytokines that recruit inflammatory cells, enhance bacterial clearance, and prime adaptive immune cells to generate pathogen-specific antibodies (27). But if the infection worsens, the local host response becomes amplified, resulting in a systemic “counterregulatory” anti-inflammatory response syndrome that aids local recruitment of inflammatory cells and prevents it from spreading systemically. However, over time, this can lead to immune paralysis (29, 35) as evidenced by impaired leukocyte, T and B cell function, bacterial clearance (“hyporesponsiveness”), and lymphocyte apoptosis in GI tract, lymph nodes, and spleen (7, 9, 20, 23, 25) that together is thought to be a major contributor to the morbidity and mortality of sepsis (8). Indeed, blockade of proinflammatory pathways has not been effective clinically and may have increased mortality in some subpopulations. This immunosuppressive environment increases the susceptibility to subsequent sepsis episodes and other infections (45) and may account for the high mortality rate of sepsis.

Innate immunity is triggered by Toll-like receptors (TLRs), which recognize infection of microbial pathogens and initiate the innate response. Most TLR signals are activated via MyD88, which is an essential downstream adaptor molecule that activates signaling pathways such as mitogen-activated protein kinases and NF-κB with subsequent induction of cytokines. Weighardt et al. (44) demonstrated that MyD88-deficient mice were resistant to polymicrobial sepsis produced by a colon ascendsent stent peritonitis (CASP), although TLR2- and TLR4-deficient mice were not resistant. We recently showed that a cecal ligation and puncture (CLP)-induced acute kidney injury (AKI) was attenuated in MyD88-deficient mice but not in TLR4-deficient mice (11).

The TLRs that recognize bacterial and viral nucleic acids (3, 7–9) are found in the endosomal compartment (4, 26) and appear to be trafficking between endosomes and lysosomes (1, 14, 29). When trafficking and/or acidification is disrupted by chloroquine or bafilomycin A1, TLR signaling is inhibited (1, 14, 30, 32, 37). In addition to these studies in vitro, a few reports suggest that chloroquine can inhibit innate immune responses in vivo: in a two-hit model of hemorrhage then CLP (13), after CpG/LPS administration (18), and in a mouse cryptococcosis infection model (31).

We sought to determine the therapeutic potential of the clinically well-tolerated TLR inhibitor chloroquine, in CLP-induced polymicrobial sepsis in elderly mice treated with fluid and antibiotics, a clinically relevant model of sepsis and sepsis-induced AKI. Furthermore, we examined TLR9 as a potential target for chloroquine action by using an oligo-
Chloroquine and inhibition of TLR9 protect from sepsis.

Deoxynucleotide TLR9 inhibitor H154 and TLR9-deficient mice.

Materials and Methods

**Animals.** Animal care followed National Institutes of Health (NIH) criteria for the care and use of laboratory animals in research, under a protocol approved by the National Institute of Diabetes and Digestive and Kidney Diseases Animal Care and Use Committee. C57BL/6 mice (32–40 wk of age) were obtained from NIH (Frederick, MD). TLR9-deficient mice were obtained from S. Akira (Osaka University, Osaka, Japan) (17). The TLR9-deficient mice were backcrossed with C57BL/6 mice for >8 generations before TLR9-deficient mice were reestablished.

**Surgery.** The CLP procedure used for induction of septic peritonitis was described in detail previously (49).

**Drug treatments.** Chloroquine (50 mg/kg; Sigma, St. Louis, MO) or an equal volume of saline was administered orally at either 3 h before or 6 h after CLP surgery.

Phosphorothioate-containing oligodeoxynucleotides were synthesized as described previously (40, 48) and administered (3 mg/kg) intraperitoneally immediately after CLP surgery. A suppressive ODN H154 (5′-CTTACAGCTGTAGGGG-3′) acts specifically through TLR9 (48), whereas a control suppressive ODN A151 (5′-TTAGGTTAGGTTAGGTTAGG-3′) does not act through TLR9 (40).

**Survival study.** Survival was assessed every ~6–12 h after surgery. Antibiotic injection and fluid resuscitation were started 6 h after surgery by subcutaneous injection, and then repeated every 12 h for 4 days. Animals exhibiting extreme morbidity were euthanized.

**Renal pathology.** Tissue was fixed in 10% formalin and embedded in paraffin. Four-micrometer sections were stained with periodic acid Schiff (PAS) reagent. Tubular damage was assessed by counting vacuolized tubules at ×400 magnification using >100 randomly selected tubules from each animal.

**Splenic apoptosis.** Active caspase 3 staining, a marker of splenic apoptosis, was examined in >5 randomly chosen ×400 fields of white pulp and expressed as positive cells per high-power field as described previously (11).

**Measurements blood urea nitrogen, serum creatinine, and cytokines.** Blood urea nitrogen (BUN) and serum creatinine were measured as previously described (34, 50). Serum TNF-α and IL-10 were determined by ELISA (R&D Systems, Minneapolis, MN).

**Statistical analysis.** All the data are expressed as means ± SE. Differences between groups were examined for statistical significance by ANOVA with a multiple comparison correction or t-test (Prism 4, GraphPad Software, San Diego, CA or SigmaStat, Systat Software, Point Richmond, CA). A P value <0.05 was accepted as statistically significant.

Results

**Chloroquine improves survival and AKI after polymicrobial sepsis.** We determined whether chloroquine alters sepsis-induced mortality and renal dysfunction in aged mice treated with fluid and antibiotics. Chloroquine given 3 h before CLP surgery significantly improved survival: at 96 h after CLP, the survival was 15% for mice treated with vehicle, and 69% for mice treated with chloroquine (Fig. 1A). Delaying chloroquine administration to start at 6 h after CLP, when clinical symptoms first appear, also improved survival; at 96 h after CLP, the survival was 24% for mice treated with vehicle, and 55% for mice treated with chloroquine (Fig. 1B). These results suggest that chloroquine can function both as a preventative and as a therapeutic agent in polymicrobial sepsis.

Chloroquine administration, either 3 h before or 6 h after CLP, significantly improved both sepsis-induced increases in serum creatinine and BUN (Fig. 2A) and tubular damage in both the outer stripe of the outer medulla and the cortex [Fig. 2B, Supplement Fig. 1, A and B (The online version of this article contains supplemental data)]. To assess injury to other organs, ALT, AST, amylase, CK, and LDH were measured in serum. As previously reported (34), all of these enzymes were significantly elevated 24 h after CLP. Treatment with chloroquine 3 h before CLP resulted in a significant decrease in each of these parameters. Treatment with chloroquine 6 h after CLP also improved each parameter; however, only ALT was significantly decreased (Fig. 3).

**Chloroquine administration reduces splenic apoptosis after polymicrobial sepsis.** Apoptosis of splenocytes has been shown to worsen the outcome of sepsis and contribute to immune depression (19, 38, 46). Therefore, we evaluated the effects of chloroquine administration on splenic apoptosis. Splenic apoptosis, as detected by active caspase3, occurred mostly in the white pulp. CLP-induced sepsis profoundly increased the number of active caspase3-positive cells in the white pulp of the spleen at 24 h after CLP (Fig. 4A, Supplement Fig. 2), as previously described (11). Chloroquine, administered either 3 h before or 6 h after CLP, significantly decreased the number of active caspase3-positive cells in the spleen at 24 h after CLP (Fig. 4A, Supplement Fig. 2). Chloroquine also significantly reduced bacterial counts in blood but had no effect on bacterial counts in peritoneal fluid (Supplement Fig. 3).
Effect of TLR9 deficiency and inhibition. Because of its sensitivity to chloroquine and its predominant expression in the spleen, TLR9 is a good candidate for mediating the beneficial effects of chloroquine. We used TLR9-deficient mice to determine whether the absence of TLR9 could mimic the effects of chloroquine. Survival at 96 h after CLP was significantly improved from 23% in wild-type mice to 70% in TLR9-deficient mice (Fig. 5A). TLR9-deficient mice also had significantly reduced BUN and serum creatinine levels 24 h after CLP compared with wild-type mice (Fig. 5C). Splenic apoptosis, as measured by active caspase3 staining, was also significantly decreased in TLR9-deficient mice compared with wild-type mice (Fig. 5B, Supplement Fig. 4). Increases in proinflammatory and anti-inflammatory cytokines TNF-α and IL-10, respectively, were blunted in TLR9-deficient mice compared with wild-type mice (Fig. 5E).

To complement the studies on TLR9-deficient mice, we used a selective TLR9 oligodeoxynucleotide inhibitor H154 (28) and a noninhibitory oligodeoxynucleotide A151. H154, but not A151, significantly inhibited the increase of BUN and serum creatinine after CLP (Fig. 6A), significantly inhibited the increase in serum AST, ALT, amylase (not significant), CK, and LDH (Fig. 6, B and C), significantly inhibited the increase in splenic active caspase3 staining (Fig. 6D, Supplement Fig. 5), and significantly inhibited the increase in TNF-α and IL-10 (Fig. 6E), confirming the results from TLR9-deficient mice. By all of these parameters, TLR9 deficiency mirrored chloroquine treatment, implicating TLR9 as a substantial target that may
account for much of the protective effects of chloroquine. However, H154 did not significantly reduce bacterial counts in blood (Supplement Fig. 3), which may reflect a TLR9-independent component of chloroquine action on immune function.

Finally, we examined the effect of chloroquine and TLR9 inhibition on TLR9 protein abundance. In SLE patients and a SLE animal model, TLR9 expression in renal tubules was upregulated (5), but we were unable to detect TLR9 in kidney after CLP (data not shown). By Western blot both oligonucleotides induced moderate increases in splenic TLR9. Chloroquine, administered either 3 h before or 6 h after CLP, decreased splenic TLR9 to undetectable levels (Supplement Fig. 6). This provides a mechanism by which chloroquine could interfere with TLR9 signaling.

Fig. 3. Chloroquine inhibits multiple organ damage after polymicrobial sepsis. Serum chemistry 24 h after sham surgery (white bars, n = 5), CLP (black bars, n = 6–9), or CLP after chloroquine treatment (gray bars, n = 6–9). Chloroquine treatment 3 h before CLP is shown on the left and chloroquine treatment 6 h after CLP is shown on the right. A: alanine aminotransferase (ALT) and aspartate aminotransferase (AST). B: amylase. C: creatine kinase (CK) and lactate dehydrogenase (LDH). Values are means ± SE. *P < 0.05 vs. CLP given vehicle.
DISCUSSION

In a clinically relevant model of polymicrobial sepsis, we found that 1) chloroquine improves mortality and reduces renal injury, even when delayed for 6 h; 2) chloroquine reduces systemic inflammation and multiple organ damage, but perhaps more critically, it improves splenic apoptosis, suggestive of a mechanism that includes reducing immune paralysis; and 3) TLR9 deficiency or TLR9 inhibition has the same effect as chloroquine, making TLR9 a likely target for the actions of chloroquine.

Chloroquine has preventative and therapeutic actions. Chloroquine improved survival, renal injury, and renal function, whether administered 3 h before surgery or 6 h after surgery, when the animals first become symptomatic. The degree of protection is similar to that seen with ethyl pyruvate (34, 41) or simvastatin (49). Chloroquine has also been shown to improve survival after CpG/LPS administration (18), in a 2-hit model of hemorrhage followed by CLP (13), and in a mouse cryptococcosis infection model (31). Hence, chloroquine might be useful for treating newly diagnosed sepsis, rather than just as a preventative agent.

Effect of chloroquine on kidney function. Chloroquine treatment either 3 h before or 6 h after CLP had a significant benefit on serum creatinine, BUN, and both cortical and OSOM histology scores. As chloroquine can have diuretic and natriuretic effects under normal conditions (3, 36), it is not known whether these phenomena contribute to the protective effect of chloroquine on kidney function during sepsis; we also cannot rule out an effect of chloroquine on sepsis-associated hypotension. We used serum enzyme markers as surrogates for multiple organ damage, as well as examined splenic apoptosis and reduction of circulating bacteria as surrogates for immune paralysis to assess whether chloroquine simply decreased the overall severity of sepsis, or whether there was any selectivity toward the kidney. Chloroquine treatment 3 h before CLP showed a broad protective effect over every marker, including renal and splenic damage. However, chloroquine administration 6 h after CLP, a more clinically relevant treatment, resulted in significant decreases in kidney and spleen injury markers, and ALT, but not AST, amylase, CK, or LDH. The differences between preventative (–3 h) vs. therapeutic (6 h) chloroquine on organ function are not consistent with prevention causing a uniform but more intense dampening of the overall extent of sepsis vs. delayed chloroquine. While not conclusive, the tighter association between mortality and both kidney and splenic injury (compared with other organs) suggests a close mechanistic link between kidney and splenic injury that may significantly contribute to mortality. Alterna-

Fig. 4. Chloroquine inhibits spleen apoptosis and serum cytokines after polymicrobial sepsis. A: number of active caspase-3-positive cells in the spleen at 24 h after CLP (n = 5 per group). B: serum TNF-α and IL-10 at 24 h after CLP (n = 6–12 per group). Values are means ± SE. *P < 0.05 vs. CLP given vehicle.
tively, chloroquine may act through a different mechanism during earlier vs. later stages of sepsis.

We conclude that chloroquine does not act exclusively through the kidney, and any connection between splenic injury and kidney injury, while consistent with our data, is only speculative at this point. Disruption of one or more mediators of this putative organ-selective injury sequence would be needed to establish such a mechanism.

Similarities between chloroquine treatment and inhibition of TLR9. We previously demonstrated that MyD88-deficient mice were resistant to sepsis and sepsis-induced AKI (11), consistent with a central role of TLR signaling. As TLR4-deficient mice had an intact sepsis-induced AKI response (11), and chloroquine inhibited sepsis-induced AKI, our data support the hypothesis that one or more of the endosomal class of TLR: TLR3, TLR7/8, TLR9, was primarily involved in the development of sepsis-induced AKI. To test this more directly, we initially focused on TLR9 by using 1) TLR9-deficient mice and 2) a selective inhibitor of TLR9, the phosphorothioate oligodeoxynucleotide H154 and its corresponding control A151. TLR9 deficiency improved CLP sepsis-induced mortality and AKI, with improvements in renal function and histology, downstream systemic pro- and anti-inflammatory cytokines, and splenic apoptosis. The TLR9 inhibitor H154 improved renal function, reduced multiple organ damage, including splenic apoptosis, and dampened systemic pro- and anti-inflammatory cytokine responses. Chloroquine also decreased TLR9 protein levels in spleen to undetectable levels, analogous to TLR9 deficiency, further supporting a TLR9-centric mechanism for chloroquine. Previous studies showed that survival after sepsis was improved in mice lacking the common TLR adapter protein MyD88 in a CASP model (44). However, survival was not improved in mice lacking TLR2 or TLR4 in a CASP model (44), and mice lacking TLR4 were either not protected (11) or only marginally protected (12) against CLP sepsis-induced AKI. Our findings further support TLR9 as a key component of
the host response to polymicrobial sepsis. We cannot eliminate significant roles of other TLRs, but if they are also important in polymicrobial sepsis, they at least require functional TLR9 signaling.

For each of the parameters tested, chloroquine administration qualitatively had the same effect as TLR9 deficiency. However, chloroquine may have effects in addition to inhibiting TLR9. Other TLRs (TLR3, 7, and 8) are internalized and function through an endosomal pathway (10, 14–16, 30) and could be inhibited by chloroquine (13, 18). In support of this view, 1) chloroquine was somewhat more effective than TLR9 deficiency to inhibit AKI, and 2) chloroquine significantly decreased blood bacterial counts, whereas reduction by the TRL9 inhibitor H154 was not significant. Chloroquine has also been reported to interfere with ERK-mediated TNF-α upregulation in vitro (43), but the relative contribution of this pathway to sepsis in vivo is not known. Chloroquine also may have direct renal effects, including increases in urine flow, sodium excretion, and glomerular filtration rate (2, 3), that might contribute to protection of tubular damage induced by CLP; whether these are related to TLRs or other actions is unknown. Nevertheless, the concordance between TLR9 deficiency and chloroquine administration on almost all measured outcomes (survival, renal injury, cytokine levels) suggests that chloro-

Fig. 6. TLR9 inhibition by phosphorothioate oligodeoxynucleotides improves acute kidney injury, multiple organ damage, and serum cytokines. Mice were subjected to sham surgery (white bars, n = 5) or CLP (black bars, n = 8), and CLP was treated with control (A151, hatched bars, n = 8) or TLR9-selective (H154, gray bars, n = 8) phosphorothioate oligodeoxynucleotides and evaluated at 24 h for kidney function (A), ALT and AST (B), amylase, CK, LDH (C), number of active caspase-3-positive cells in the spleen (D), TNF-α and IL-10 (E). Values are means ± SE. * P < 0.05 vs. WT.
Chloroquine is functioning in this model in large part via inhibition of splenic TLR9.

Chloroquine as a therapeutic agent. The spleen has become an attractive target for new therapeutic strategies for sepsis, particularly with regard to splenic apoptosis (22). Chloroquine administration compares favorably with previously studied methods to improve spleen function: caspase inhibitors (19, 24), Akt overexpression (6), Tat-BH4 or Tat-Bcl-XL peptides (21), anti-CD-40 antibody (39), caspase 8 or fas siRNA administration (46), or administration of exosomes from immature dendritic cells (33). Because of its low toxicity and administration (46), or administration of exosomes from immature dendritic cells (33), Chloroquine is functioning in this model in large part via inhibition of spleen function and vasopressin secretion: a nitric oxide-dependent effect. J Immunol Exp Ther 305: 123–130, 2003.

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