Intrarenal angiotensin II and angiotensinogen augmentation in chronic angiotensin II-infused mice

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Gonzalez-Villalobos RA, Seth DM, Satou R, Horton H, Ohashi N, Miyata K, Katsurada A, Tran DV, Kobori H, Navar LG. Intrarenal angiotensin II and angiotensinogen augmentation in chronic angiotensin II-infused mice. Am J Physiol Renal Physiol 295: F772–F779, 2008. First published June 25, 2008; doi:10.1152/ajprenal.00019.2008.—The objectives of this study were to determine the effects of chronic angiotensin II (ANG II) infusions on ANG II content and angiotensinogen expression in the mouse kidney and the role of the angiotensin II type 1 receptor (AT1R) in mediating these changes. C57BL/6J male mice were subjected to ANG II infusions at doses of 400 or 1,000 ng·kg⁻¹·min⁻¹ either alone or with an AT1R blocker (olmesartan; 3 mg·kg⁻¹·day⁻¹) for 12 days. Systolic and mean arterial pressures were determined by tail-cuff plethysmography and radiotelemetry. On day 13, blood and kidneys were collected for ANG II determinations by radioimmunoanalysis and intrarenal angiotensinogen expression studies by quantitative RT-PCR, Western blotting, and immunohistochemistry. ANG II infusions at the low dose elicited progressive increases in systolic blood pressure (135 ± 2.5 mmHg). In contrast, the high dose induced a rapid increase (152 ± 2.5, P < 0.05 vs. controls, 109 ± 2.8). Renal ANG II content was increased by ANG II infusions at the low dose (1,203 ± 253 fmol/g) and the high dose (1,258 ± 173) vs. controls (499 ± 40, P < 0.05). Kidney angiotensinogen mRNA and protein were increased only by the low dose to 1.13 ± 0.02 and 1.26 ± 0.10, respectively, over controls (1.00, P < 0.05). These effects were not observed in mice infused at the high dose and those receiving olmesartan. The results indicate that chronic ANG II infusions augment mouse intrarenal ANG II content with AT1R-dependent uptake occurring at both doses, but only the low dose of infusion, which elicited a slow progressive response, causes an AT1R-dependent increase in intrarenal angiotensinogen expression.

METHODS

Animal Preparation and Sample Collection

All protocols were approved by the Tulane University Health Sciences Center Animal Care and Use Committee. Mice (C57BL/6J, 8- to 10-wk-old males) were purchased from Jackson Laboratories (Bar Harbor, ME) and maintained in a temperature-controlled room on a 12:12-h light-dark cycle with free access to food (Na⁺ content 0.4%) and water.

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Intrarenal RAS in Angiotensin II-Infused Mice

First series. In this series, mice were trained for systolic blood pressure (SBP) determinations by tail-cuff plethysmography before recording of initial pretreatment measurements. Following this period, mice were anesthetized with 1–2% isoflurane inhalation in 100% O2 to allow subcutaneous implantation of osmotic minipumps (Alzet 1002, Durect, Cupertino, CA). ANG II was infused at doses of 400 or 1,000 ng·kg⁻¹·min⁻¹ with a separate group of each series treated with olmesartan (an AT₁R blocker, 3 mg·kg⁻¹·day⁻¹) for 12 days before sample collection. Sham-operated animals were used as controls. SBP was recorded every 3 days using the Visitech BP2000 system (Visitech Systems, Apex, NC) as the average of 30 readings/animal taken between 1:00 and 5:00 P.M. On day 13, the mice were killed by conscious decapitation to collect blood and tissue samples. The right kidney was utilized for ANG II determinations. The left kidney cortex was utilized for mRNA and protein expression studies. The kidneys were processed as described previously (14, 30).

Second series. In this series, mice were subjected to isoflurane anesthesia and a catheter connected to a radiotlemetry device was inserted in the left carotid artery to monitor heart rate and BP by telemetry in conscious, unrestrained conditions (3, 21) (model PA-C10, Data Sciences International, St. Paul, MN). After a recovery phase (7–14 days), basal mean arterial pressure (MAP) levels were established and animals were again subjected to general anesthesia for minipump implantation. Data were collected, stored, and analyzed using Dataquest A.R.T 4.0 software (Data Sciences International). ANG II was infused only at the lower dose (400 ng·kg⁻¹·min⁻¹) for 12 days before sample collection. On day 13, animals were given an isoflurane overdose and the kidneys were collected and fixed in zinc-saturated formalin and further processed for immunohistochemistry.

Determinations of ANG II, Plasma Renin Concentration, and Kidney Renin Content by RIA

For analysis of plasma ANG II, trunk blood from mice in the first series was collected in chilled tubes containing 5.0 mmol/L EDTA and a protease inhibitor cocktail. For intrarenal ANG II, the right kidneys were homogenized in methanol immediately after extraction. Later, plasma and kidney samples were processed as previously described (10, 16, 30). A rabbit anti-ANG II antiserum (Peninsula Laboratories, San Carlos, CA) was used for ANG II determinations by RIA (10, 16, 30). Plasma renin concentration (PRC) and kidney renin content (KRC) were also determined by RIA (GammaCoat, DiaSorin, Stillwater, MN) as the generation of ANG I after addition of excess renin substrate in samples collected without inhibitors.

Angiotensinogen Expression Studies

mRNA expression as determined by quantitative real-time RTPCR. RNA isolation. Five to eight milligrams of renal cortices were used to isolate total RNA using a commercial kit following the manufacturer’s recommendations (RNAlater Mini kit, Qiagen, Valencia, CA) that include DNase treatment. Total RNA was then tested for quality and quantity.

QUANTITATIVE RT-PCR. The sequence of oligonucleotide primers and Taqman probes specific for mouse angiotensinogen and mouse β-actin used were as follows: For angiotensinogen, forward 5'-CATGACCCAGTTCTTT-3', reverse 5'-AAGTGACACCTGTGTTGCAA-3', and probe 5'-6-AMC-gcttgccacagtgtgcgttg-fAM-3'. For β-actin, forward 5'-GTATCCATGCTGAGATGAAACCA-3', reverse 5'-GATGATTATGGGCAACTTGTG-3', and probe 5'-HEX-TCT-CTGCAACAACACATGTGGTTCCTGAGG-3'. All samples were amplified in triplicate in a Stratagene MX 3000P multiplex quantitative (q) RT-PCR system using Stratagene PCR master mix II (Stratagene, La Jolla, CA). Twenty nanograms of the sample were used as a template, and angiotensinogen and β-actin were coamplified in the same reaction. The conditions for the reaction were set as one individual cycle of 50°C (30 min) and 95°C (10 min) followed by 45 cycles of 95° (15 s), 65°C (60 s), and 72°C (60 s). Assay quality was determined based on standard curve values of R² ≥ 0.99, Y ≤ 3.8, and an efficiency of ≥0.80 and ≤110%. Quantitative values were extrapolated from individual standard curves for angiotensinogen and β-actin whose expression was not affected by the experiments. Results were normalized to β-actin expression.

Angiotensinogen protein expression as determined by Western blotting and immunohistochemistry. PROTEIN ISOLATION. Renal cortices were thawed and homogenized in 500 μl of lysis buffer. After that, samples were sonicated and centrifuged to collect the supernatant. Protein concentration was determined using the Bradford method (Bio-Rad protein assay, Bio-Rad, Hercules, CA).

WESTERN BLOTTING. Five micrograms of protein of each sample were subjected to gel electrophoresis and transferred to nitrocellulose membranes following protocols described elsewhere (16).

The membranes were subjected to successive probing with a polyclonal primary rabbit anti-mouse/rat angiotensinogen antibody in a 1:200 dilution overnight at room temperature (code no. 99666, IBL, Gmuma, Japan) and a secondary fluorescent-tagged goat anti-rabbit IgG antibody (IRDye 800 CW, Licor Biosciences). β-Actin detection was performed with a mouse monoclonal antibody (Abcam, 1:5,000) and a secondary fluorescent-tagged anti-mouse IgG antibody (IRDye 680 goat anti-mouse, Licor Biosciences). Fluorescent intensity was collected in two separate channels using an infrared Odyssey system (Licor Biosciences). The intensity of each angiotensinogen band was normalized to β-actin detected in the same membrane.

IMMUNOHISTOCHEMISTRY. From each paraffin-embedded kidney, 10 consecutive 3-μm tissue slices were obtained, of which one slice per mouse was selected randomly for staining. After antigen activation, a Dako Envision plus system-HRP (Dakocytomation, Carpinteria, CA) was used to detect the binding of rabbit primary antibodies against mouse/rat angiotensinogen (IBL, see above) in the slice. Each tissue slice was probed at a concentration of 0.25 μg/ml for 15 min. Samples were also counterstained with hematoxylin before analysis. The staining protocol was performed to completion in an Autostainer Plus (Dako). Immunoreactivity was semiquantitatively evaluated in a blind manner as described elsewhere (16). Briefly, 20 microscopic fields/slide were selected at random for evaluation. Experiments were performed using a microscope with ×200 magnification (Olympus BX51-TRF) and an integrated digital camera system (Magnafire SP). In every field, the sum of intensities of positive areas was calculated and averaged by the field area using Image Pro-plus software (Media Cybernetics, Bethesda, MD). The field area was calculated as the total field area minus the areas corresponding to glomeruli, blood vessels, and empty spots in the field. The average of intensities of the 20 fields represented the total intensity per slide and therefore per mouse.

Statistical Analyses

All data are presented as means ± SE. Two-way ANOVA with Bonferroni’s posttest were used when blood pressure changes were analyzed. A paired t-test was applied when changes in body weight were studied. For the rest of the data, one-way ANOVA with Bonferroni’s posttest were applied. A value of P < 0.05 was regarded as significant.

RESULTS

Body and Kidney Weights

Control mice gained weight during the period of the study (23.4 ± 0.6 before vs. 24.5 ± 0.3 g at the completion of the study, n = 6). Mice infused at a low dose of ANG II (400 ng·kg⁻¹·min⁻¹) showed a similar gain (24.1 ± 0.3 before vs. 25.1 ± 0.3 g after, n = 8, P < 0.05). Mice subjected to a high dose of ANG II (1,000 ng·kg⁻¹·min⁻¹) failed to show such progression (23.1 ± 0.4 before vs. 22.9 ± 0.5 g after treatment,
Olmesartan treatment did not affect the weight gain in controls and mice infused at a low dose (mice treated with 400 ng·kg\(^{-1}\)·min\(^{-1}\) plus olmesartan weight was 22.6 ± 0.4 vs. 24 ± 0.6 g after treatment, \(n = 5\), \(P < 0.05\)). Furthermore, when given to mice infused at 1,000 ng·kg\(^{-1}\)·min\(^{-1}\), olmesartan restored normal weight gain in this group (22.8 ± 0.6 vs. 24.4 ± 0.6 g after treatment, \(n = 5\), \(P < 0.05\)).

ANG II infusions caused a significant reduction of kidney weight (KW) only when infused at the high dose (139.6 ± 3.5 mg, \(n = 11\), \(P < 0.05\)) compared with controls (156.6 ± 4.7 mg, \(n = 7\)). This reduction was prevented by cotreatment with olmesartan (ANG II 1,000 ng·kg\(^{-1}\)·min\(^{-1}\)+olmesartan, 160.6 ± 4.9 mg, \(n = 5\), NS). ANG II infusions at the low dose either alone (150 ± 3.0 mg, \(n = 6\)) or in combination with olmesartan (ANG II 400 ng·kg\(^{-1}\)·min\(^{-1}\)+olmesartan, \(n = 5\), 163.6 ± 6.4 mg, NS) did not change KW significantly. Finally, olmesartan treatment did not change KW in controls (161.5 ± 7.5 mg, \(n = 4\), NS).

**SBP**

All groups displayed the same level of BP (107–110 mmHg) at the beginning of the study (Fig. 1). SBP determined by tail-cuff plethysmography (Fig. 1A) remained stable in controls during the study, with SBP averaging 109 ± 2.8 mmHg by day 12 (\(n = 13\)). ANG II infused at the low dose caused a slowly progressive increase in BP that reached significance (as determined by 2-way ANOVA) after 9 days of infusion; by day 12, the average SBP of this group was 135 ± 4.0 mmHg (\(n = 16\)). In contrast, ANG II at the high dose caused a rapid and sustained increase in BP that reached significance by day 3 and onwards.

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**Fig. 1.** Effects of chronic ANG II and olmesartan infusions on systolic arterial pressure (SAP) as determined by tail-cuff plethysmography (A) and telemetry (B) and mean arterial pressure (MAP; C). Telemetry recordings (B) obtained at the same hours as the tail-cuff measurements (A) (between 1 and 5 P.M.) were averaged to allow comparison between these methods. C: 3-h MAP averages separated in 24-h intervals. Olm, olmesartan (3 mg·kg\(^{-1}\)·day\(^{-1}\)). *\(P < 0.05\) for daily averages vs. controls. **\(P < 0.05\) for daily averages vs. controls in olmesartan-treated animals.
remained elevated throughout the experiment; by day 12 the SBP was 152 ± 2.5 mmHg (n = 11).

Olmesartan treatment markedly reduced SBP in control mice and prevented the development of hypertension in mice treated with both doses of ANG II [ANG II 400 ng·kg⁻¹·min⁻¹ + olmesartan (n = 5), 77 ± 2.9 mmHg; ANG II 1,000 ng·kg⁻¹·min⁻¹ + olmesartan (n = 5), 78 ± 3.4 mmHg]. These changes in SBP were similar to those observed in controls treated only with olmesartan [82 ± 3.4 mmHg (n = 5)].

Because mice infused with 400 ng·kg⁻¹·min⁻¹ of ANG II displayed a SBP profile by tail-cuff plethysmography that resembled the slow pressure response observed in other models of ANG II-dependent hypertension (2, 13, 39), the effects of this dose of ANG II on BP were further assessed by telemetry (Fig. 1, B and C). Systolic arterial pressure telemetric recordings between 1 and 5 P.M. (the same time of the day when tail-cuff measurements were taken) (Fig. 1B) showed similar values for control animals that also remained stable during the experiment (109 ± 4.3 mmHg by day 12, n = 6). Meanwhile, ANG II-infused mice exhibited progressive increases in systolic arterial pressure recordings that reached higher values than those recorded by the tail-cuff method (146 ± 6.4 mmHg by day 12, n = 10). Hence, a higher difference was found between these two groups by telemetry (36 mmHg) than by plethysmography (26 mmHg).

MAP

Both groups, controls and ANG II-infused mice, displayed a circadian pattern in MAP with higher values during night hours (Fig. 1C) than during day hours as previously described (34). However, there was a slow response pattern toward higher BP in ANG II-infused mice that reached significance by day 9 as determined by two-way ANOVA, and these mice displayed higher BP values at all times during the 24-h cycles. By day 12, control animals had a MAP of 107 ± 1.9 mmHg (n = 6) vs. ANG II-infused mice, which had a MAP of 139 ± 4.4 (n = 10) (P < 0.05).

Plasma Levels of ANG II

ANG II infusions caused significant changes in plasma ANG II levels (Fig. 2A, P < 0.05 by one-way ANOVA overall). Compared with controls (282 ± 46 fmol/ml, n = 10), plasma ANG II in mice infused with the low dose, although higher, was not statistically different (497 ± 68 fmol/ml, n = 15, NS). In turn, the high dose induced a threefold increase in plasma ANG II that was significant (784 ± 271 fmol/ml, n = 5, P < 0.05).

Mice treated with olmesartan displayed significant increases in ANG II plasma levels compared with controls (sham-operated) as ANG II values in the olmesartan-only group were 715 ± 257 fmol/ml (n = 5, P < 0.05). In mice infused with the low dose of ANG II plus olmesartan, plasma ANG II values were 627 ± 141 fmol/ml (n = 5), and those mice infused with the high dose plus olmesartan showed values of 637 ± 107 fmol/ml (n = 5). These changes were not significant compared with mice infused only with ANG II at the same doses.

Intrarenal Levels of ANG II

ANG II infusions caused significant increases in intrarenal ANG II at low dose (1,203 ± 253 fmol/g, n = 7) and high dose (1,258 ± 173 fmol/g, n = 11). These changes represent a 2.5-fold increase compared with controls (499 ± 40 fmol/g, n = 8) (Fig. 2B). In all groups, ANG II intrarenal levels were higher than plasma values.

Olmesartan treatment prevented the increases in intrarenal ANG II observed during chronic ANG II infusions at both doses (ANG II 400 ng·kg⁻¹·min⁻¹ plus olmesartan, 352 ± 44 fmol/g, n = 5, ANG II 1,000 ng·kg⁻¹·min⁻¹ plus olmesartan 205 ± 28 fmol/g, n = 5).

PRC and KRC

Figure 3A shows that chronic ANG II infusions caused a marked suppression of PRC as control animals displayed a PRC of 427.80 ± 43.14 ng ANG I·ml⁻¹·h⁻¹ (n = 9) vs. ANG II-infused animals that had a PRC of 4.90 ± 4.62 ng ANG I·ml⁻¹·h⁻¹ when infused with the low dose (n = 3) and 10.81 ± 0.87 ng ANG I·ml⁻¹·h⁻¹ when infused with the high dose (n = 9). In contrast, compared with controls KRC (Fig. 3B) (4,428 ± 323.40 ng ANG I·mg⁻¹·h⁻¹·l⁻¹, n = 9), no reduction was observed in ANG II-infused mice with the low dose (4,367 ± 408.40 ng ANG I·mg⁻¹·h⁻¹·l⁻¹, n = 4) and only a 50% reduction in KRC was achieved during ANG II infusion with the high dose (2,385 ± 337.80 ng ANG I·mg⁻¹·h⁻¹·l⁻¹, n = 9).

*Fig. 2. Effects of chronic infusions of ANG II either alone or with olmesartan on ANG II plasma (A) and intrarenal levels (B). *P < 0.05 vs. controls. *P < 0.05 within group by 1-way ANOVA and Bonferroni’s posttest.

Fig. 3. Effects of chronic infusions of ANG II either alone or with olmesartan on PRC and KRC.
Expression of Intrarenal Angiotensinogen

Figure 4 displays the effects of ANG II infusions on angiotensinogen mRNA expression (Fig. 4A) and protein (Fig. 4B) in the renal cortex as determined by qRT-PCR and Western blotting from tissue homogenates. Compared with angiotensinogen mRNA in controls (normalized expression of angiotensinogen/β-actin as 1.00 ± 0.04, n = 11), the low dose of ANG II infusion caused a significant increase in angiotensinogen mRNA levels (1.13 ± 0.03, n = 13). In contrast, the high dose failed to cause significant changes (1.00 ± 0.03, n = 12).

When angiotensinogen protein expression was compared (Fig. 4B), only mice infused at the low dose of ANG II showed a significant increase in angiotensinogen protein expression (1.26 ± 0.13, n = 11, P < 0.05) vs. controls (1.00 ± 0.11, n = 11). The high dose did not cause any significant changes (1.00 ± 0.03, n = 12). Olmesartan treatment prevented the effects of ANG II to stimulate angiotensinogen mRNA and protein expression. Angiotensinogen mRNA expression in the 400 ng·kg⁻¹·min⁻¹ ANG II plus olmesartan group was 1.07 ± 0.02, (n = 5, NS vs. controls). In the same group, angiotensinogen protein expression was 0.75 ± 0.06. In controls, olmesartan treatment also reduced the expression of angiotensinogen mRNA (0.86 ± 0.03, n = 5) but not protein. No significant changes were observed in the group treated with ANG II at 1,000 ng·kg⁻¹·min⁻¹ plus olmesartan in mRNA or protein expression (n = 4).

The immunohistochemical analysis (Fig. 5) demonstrates that angiotensinogen in the mouse kidney is expressed mainly in proximal tubules and that this expression was enhanced by ANG II infusions only at low doses, as the semiquantitative analysis revealed a marked induction of tubular angiotensinogen protein expression in infused mice (Fig. 5B, 13.97 ± 4.64, n = 4, P < 0.001) over controls (Fig. 5A, normalized as 1.00 ± 0.40, n = 10); in contrast, the high-dose infusion did not elicit any significant increases (Fig. 5C) (3.00 ± 0.94, n = 5). A higher magnification (×1,000) also revealed the angiotensinogen staining tends to locate to the apical side of the proximal tubule cells (Fig. 5D).

DISCUSSION

The present data demonstrate that an infusion of ANG II at a dose of 400 ng·kg⁻¹·min⁻¹ in mice causes a progressive increase in BP accompanied by significant augmentation in intrarenal angiotensinogen mRNA and protein and that this response is dependent on the activation of AT₁R. In contrast, a high dose of ANG II (1,000 ng·kg⁻¹·min⁻¹) failed to produce significant increases in intrarenal angiotensinogen despite eliciting a greater increase in BP. The increase in angiotensinogen expression during chronic ANG II infusions at low doses was demonstrated at the mRNA level by qRT-PCR and at the protein level by Western blotting and supported by immuno-
histochemistry. The difference in magnitude of the observed changes in angiotensinogen protein expression by these two methods might be explained by their different nature as Western blotting is a quantitative method and immunohistochemistry is semiquantitative. Nevertheless, these observations indicate that intrarenal angiotensinogen augmentation is not directly dependent on the ANG II dose or the extent of the increase in BP. Thus other factors participate in proximal tubule angiotensinogen regulation during chronic ANG II infusions.

The results also show that different ANG II doses elicit different phenotypes in mice. The retarded growth shown by the group infused with the high dose is similar to what has been reported in the past by Song et al. (33). In their report, it was demonstrated that mice infused chronically with ANG II (500 ng·kg⁻¹·min⁻¹) develop muscle wasting that is mediated by ANG II-induced downregulation of IGF-1 in skeletal muscles. In turn, mice infused with the low dose of ANG II displayed a normal progression of weight gain. Mice infused with the high ANG II dose displayed a different BP profile characterized by a rapid and pronounced increase in BP compared with the slowly progressive increase observed with the low dose. These responses to the high dose may reflect a malignant form of hypertension and potential target organ damage and might help to explain why this dose failed to induce increases in angiotensinogen expression. However, no evidence specifically demonstrating renal injury in this model is presented. Wesseling et al. (37) reported that C57BL female mice infused with ANG II for 4 wk at different doses, including some higher than those used in the present report, were resistant to kidney damage despite the development of hypertension. It is possible that female mice may be more resistant to hypertension-induced injury. Using a transgene mouse model overexpressing human angiotensinogen in proximal tubule cells and human renin, Kobori et al. (15) reported the development of initial signs of renal injury, including afferent arteriolar wall thickening, increased interstitial macrophage/monocyte infiltration, and increased collagen deposition.

The rapid increase in BP observed during infusion with the high dose also indicates a direct vasoconstrictor effect of ANG II. Furthermore, this dose of ANG II infusion elicits substantial oxidative stress and other extrarenal ANG II effects that might influence renal angiotensinogen expression (12). It is also possible that high doses of ANG II facilitate interactions with other receptors, like AT₂R and megalin, that may counteract AT₁R actions. However, in the case of the AT₂R, findings by Cervenka et al. (4) and Wesseling et al. (37) point out that it is unlikely that AT₂R activation contributes significantly to the ANG II-mediated increases in arterial pressure during ANG II-dependent hypertension.

This study demonstrates that ANG II-induced hypertension in mice is characterized by elevated plasma and intrarenal ANG II levels similar to what has been observed in other animal models (8, 23, 39). However, when ANG II levels are compared with those in rats, the best characterized species in ANG II-dependent hypertension, mice display plasma and
kidney values that are higher even for the control animals (28). These findings indicate the existence of interspecies differences that need to be considered when the intrarenal RAS is studied and when such results are extrapolated between species, including humans. It has been shown that the development of hypertension elicited by chronic ANG II infusions is AT1R-dependent as it is prevented by treatment with angiotensin receptor blockers (16, 18). Also, ANG II infusions into mice lacking AT1Rs in the kidney fail to develop high BP (5). Results from this work further support this notion as olmesartan treatment prevents the development of hypertension, as well as the increases in intrarenal ANG II content or angiotensinogen expression.

The present study extends and confirms previous results regarding the effect of chronic infusions of ANG II at two different doses (400 and 1,000 ng·kg⁻¹·min⁻¹) on SBP (12) and MAP (36). Furthermore, it has been reported that BP and heart rate in mice show a circadian rhythm with higher blood pressure values with high levels of activity during the night phase (34). Although the circadian pattern was preserved during chronic ANG II infusion, higher BP levels were maintained throughout the 24-h period and did not return to normal levels even during the day phase.

Mice infused at a low dose of ANG II (400 ng·kg⁻¹·min⁻¹) display a BP profile that fits the description of the slow pressor response observed in other models for ANG II-dependent hypertension including rats (18, 39), dogs (2), and humans (1). Several observations support the fact that this response involves an activation of the intrarenal RAS as it is a salt-sensitive form of hypertension accompanied by ANG II-mediated reductions in kidney function and sodium excretion (22, 35), and oxidative stress (12, 18, 24, 36). The activation of the angiotensinogen gene observed in mice infused at the low dose likely provides the substrate for additional local generation of ANG II. Furthermore, circulating ANG II, while being effective at reducing plasma renin concentration, failed to suppress kidney renin content in mice infused at the low dose and caused only partial suppression in mice infused with the high dose. While juxtaglomerular renin is suppressed by ANG II infusions, it has been shown that renin in the distal nephron is actually upregulated (30), suggesting that this might be the source of maintained intrarenal renin content. Collectively, these findings suggest that chronic ANG II infusions at the low dose cause an augmentation of the intrarenal RAS. This augmentation is mediated by ANG II uptake by the AT1R and activation of angiotensinogen expression. Indeed, transgenic mice with induced overactivity of this intrarenal RAS develop hypertension and renal damage in a manner that is independent from the systemic RAS (15, 19, 32).

In summary, mice display plasma and intrarenal ANG II levels that are higher than in other species even under basal conditions. During ANG II-induced hypertension, in particular when low doses of ANG II infusions are used, these levels are further increased and they are associated with augmentation of intrarenal angiotensinogen mRNA and protein via AT1R activation as well as AT1R-mediated uptake. These observations provide the foundation for future studies designed to determine the quantitative contribution of the intrarenal RAS and, specifically, proximal tubule angiotensinogen-derived ANG II to the overall levels of ANG II in the kidney and its regulation.

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