Proinsulin C-peptide reduces diabetes-induced glomerular hyperfiltration via efferent arteriole dilation and inhibition of tubular sodium reabsorption

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Nordquist L, Brown R, Fasching A, Persson P, Palm F. Proinsulin C-peptide reduces diabetes-induced glomerular hyperfiltration via efferent arteriole dilation and inhibition of tubular sodium reabsorption. Am J Physiol Renal Physiol 297: F1265–F1272, 2009. First published September 9, 2009; doi:10.1152/ajprenal.00228.2009.—C-peptide reduces diabetes-induced glomerular hyperfiltration in diabetic patients and experimental animal models. However, the mechanisms mediating the beneficial effect of C-peptide remain unclear. We investigated whether altered renal afferent-effferent arteriole tonus or alterations in tubular Na+ transport (TNa) in response to C-peptide administration mediate the reduction of diabetes-induced glomerular hyperfiltration. Glomerular filtration rate, filtration fraction, total and cortical renal blood flow, total kidney O2 consumption (QO2), TNa, fractional Na+ and Li+ excretions, and tubular free-flow and stop-flow pressures were measured in anesthetized adult male normoglycemic and streptozotocin-diabetic Sprague-Dawley rats. The specific effect of C-peptide on transport-dependent QO2 was investigated in vitro in freshly isolated proximal tubular cells. C-peptide reduced glomerular filtration rate (~24%), stop-flow pressure (~8%), and filtration fraction (~17%) exclusively in diabetic rats without altering renal blood flow. Diabetic rats had higher baseline TNa (40%), which was reduced by C-peptide. Similarly, C-peptide increased fractional Na+ (80%) and Li+ (47%) excretions only in the diabetic rats. None of these parameters was affected by vehicle treatments in either group. Baseline QO2 was 37% higher in proximal tubular cells from diabetic rats than controls and was normalized by C-peptide. C-peptide had no effect on ouabain-pretreated diabetic cells from diabetic rats. C-peptide reduced diabetes-induced hyperfiltration via a net dilation of the efferent arteriole and inhibition of tubular Na+ reabsorption, both potent regulators of the glomerular net filtration pressure. These findings provide new mechanistic insight into the beneficial effects of C-peptide on diabetic kidney function.


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sured as fractional urinary Na\(^+\) and Li\(^+\) excretions and inhibition of cellular QO\(2\) related to electrolyte transport), or a combination thereof.

**MATERIALS AND METHODS**

Animals and induction of diabetes. Age-matched male Sprague-Dawley rats (250–330 g body wt) were purchased from M & B (Ry, Denmark). Animals had free access to water and standard rat chow (0.3% Na, 0.8% K, 21% protein; R3, Ewos, Södertälje, Sweden) throughout the study. All experiments were performed in accordance with the National Institutes of Health guidelines for use and care of laboratory animals and approved by the Animal Care and Use Committee of Uppsala University. Diabetes was induced by an injection of STZ (50 mg/kg body wt; Sigma-Aldrich, St. Louis, MO) into the tail vein. Animals were considered diabetic if blood glucose concentrations increased to >18 mM within 24 h after STZ injection and remained elevated. Test reagent strips (MediSence, Bedford, MA) were used to determine blood glucose concentrations from blood samples obtained from the cut tip of the tail in all animals.

**Surgical procedures.** At 2 wk after allocation to the study, the animals were anesthetized with thiobutabarbital (120 mg/kg body wt ip; Inactin, Sigma-Aldrich), placed on a thermocontrolled operating table at 37°C, and tracheotomized. Polyethylene catheters were placed in the right femoral vein for infusion of Ringer solution (5 and 10 ml·kg body wt \(^{-1}\)·h\(^{-1}\) for normoglycemic controls and diabetic animals, respectively), the right femoral artery for blood pressure measurements (model P23Db, Statham Laboratories, Los Angeles, CA), and the left renal vein for blood sampling. The left ureter was catheterized for collection of urine for subsequent analysis, and the urinary bladder was catheterized to allow urinary drainage. The left kidney was exposed by a left subcostal flank incision, immobilized in a plastic cup, and embedded in pieces of saline-soaked cotton wool, and the surface was covered with paraffin oil (Apoteksbolaget, Gothenburg, Sweden).

**Simultaneous measurements of GFR, total and cortical RBF, and total QO\(2\).** GFR was estimated by measurements of \(^{3}\)Hinulin clearance. \(^{3}\)H (American Radiolabeled, St. Louis, MO) was initially administered as a 185-kBq bolus and then continuously infused in Ringer solution (185 kBq·kg body wt \(^{-1}\)·h\(^{-1}\)). \(^{3}\)H activities in urine and plasma were measured using a standard liquid scintillation technique, and GFR was calculated as follows: GFR = U\(\text{v} \cdot \text{W}/\text{P}, where U and P denote the activity of \(^{3}\)Hinulin in the urine and plasma, respectively, and V denotes the urine flow rate (in ml/min). After a 45-min recovery period, baseline measurements were obtained for 30 min. A single bolus dose of rat C-peptide (5 nmol/kg body wt; Sigma-Genosys, Cambridge, UK; >95% purity as measured by HPLC) was followed by a 40-min continuous infusion of C-peptide (50 pmol·kg body wt \(^{-1}\)·min\(^{-1}\)), which should result in plasma C-peptide concentrations within the physiological range (51). Thereafter, all parameters were measured during a second 30-min experimental period. Additional control and diabetic animals were given only saline vehicle and served as time controls, since the scrambled C-peptide has previously been shown to have no physiological effects (2, 35, 51).

Total RBF was measured using an ultrasound probe (Transonic Systems, Ithaca, NY) placed around the left renal artery, and cortical RBF was measured with laser-Doppler flowmetry (model PF 4001-2, Perimed, Stockholm, Sweden) (10). All measured parameters were continuously recorded with a MacLab instrument (AD Instruments, Hastings, UK) connected to a Macintosh Power-PC 6100. Blood gas parameters were analyzed on samples drawn from the left renal vein and femoral artery during the first and second measurement periods. Kidney weights were determined at the end of the experiment. Urine volumes were measured gravimetrically, and urinary Na\(^+\) concentrations were determined by flame spectrophotometry (model IL543, Instrumentation Lab, Milan, Italy).

Estimations of proximal tubular reabsorption using Li\(^+\) clearance. In a separate set of control and diabetic rats (n = 7–8/group), Li\(^+\) clearance was measured according to a previously published procedure (32). Briefly, the animals were subjected to the surgical protocol described above (see **Surgical procedures**), except only the bladder was catheterized for urine collection, and the subcostal flank incision and the immobilization of the kidney with the plastic cup were not performed. Li\(^+\) was administered as a 4-mg ip bolus of LiCl at the onset of anesthesia followed by a continuous intravenous infusion (2.1 mg·h\(^{-1}\)·rat\(^{-1}\)), resulting in plasma concentration of 0.2–0.4 mM. After surgery, the experimental protocol described above was performed for measurements of Li\(^+\) and inulin clearance in these animals. Since C-peptide reduces GFR and, therefore, tubular Li\(^+\) load via the same mechanism, estimations of proximal tubular reabsorption were calculated as fractional urinary Li\(^+\) excretion.

**Calculations.** The filtration fraction (FF) was estimated as follows:

\[
\text{FF} = \frac{\text{GFR}/\text{RBF} \times (1 - \text{Hct})}{\text{RBF}},
\]

Renal vascular resistance (RVR) was calculated as mean arterial pressure (MAP) divided by RBF. In vivo renal oxygen consumption (QO\(2\)) (µmol/min) was estimated from the arteriovenous difference in O\(2\) content (\(Q_{\text{O2}}\text{ct} = ([\text{Hb}]\cdot\text{oxygen saturation}) + P_{O2}\cdot0.22\)) × total RBF.

Tubular Na\(^+\) clearance (\(\text{TNa}\text{cl}\)) was calculated as follows:

\[
\text{TNa}_{\text{cl}} = [\text{PNa}] - \text{GFR}, where [\text{PNa}] is plasma Na\(^+\) concentration. \text{TNa}_{\text{cl}} \text{per} \text{QO}_2 \text{was calculated as} \text{TNa}_{\text{cl}}/\text{QO}_2. \text{Fractional Na}\(^+\) \text{excretion was estimated from} [[\text{Una}]\cdot[\text{Pnula}] / [\text{Pna}]\cdot[\text{Uinulin}], and fractional Li\(^+\) \text{excretion was calculated according to} [[\text{Uli}]\cdot[\text{Pnulia}] / [\text{Pli}]\cdot[\text{Uinulin}], where [\text{Una}] is urinary Na\(^+\) concentration, [\text{Pnulia}] and [\text{Uinulin}] represent plasma and urinary inulin concentration, and [\text{Pli}] is plasma Li\(^+\) concentration.

**Measurements of tubular pressures.** The net glomerular filtration pressure (\(P_{\text{net}}\)) was estimated by stop-flow technique at baseline and after 40 min of infusion of rat C-peptide (5 nmol/kg body wt bolus + continuous infusion of 50 pmol·kg body wt \(^{-1}\)·min\(^{-1}\) in 5 control and 6 diabetic animals). Early proximal tubular segments (≥5 rat) were randomly chosen on the kidney surface before and after C-peptide administration. A pipette, filled with 1 M NaCl and Lissamine green and connected to a servo-null pressure system (World Precision Instruments, New Haven, CT), was used to determine proximal tubular free-flow pressure (\(P_{\text{f}}\)) and stop-flow pressure (\(P_{\text{s}}\)), the latter after tubular flow was interrupted with a wax blockade distal to the pressure pipette. \(P_{\text{s}}\) was calculated as \(P_{\text{f}} - P_{\text{g}}\).

**Measurements of \(QO_2\) in vitro.** Proximal tubular cells were isolated from normoglycemic controls (n = 9) and STZ-diabetic rats (n = 12) as previously described (27, 39, 40). Briefly, the renal cortical tissue

![Fig. 1. Glomerular filtration rate in normoglycemic control and hyperglycemic diabetic rats before and after infusion of vehicle or C-peptide. Values are means ± SE (n = 8–10/group). NS, not significant.](https://ajprenal.physiology.org/doi/10.1152/ajprenal.00334.2017)
Mm CaCl₂, 10.0 mM HEPES, 0.5 mM Ca lactate, 2.0 mM glutamine, injected into the chamber, and the rate of O₂ disappearance was 23.2 mM for cells from diabetic animals. O₂ content was measured by the medium was 5.8 mM for cells from normoglycemic controls and stirred with an air-driven magnetic stirrer. Glucose concentration in and 50 U/ml streptomycin, 298°C controlled (37°C) gas-tight Plexiglas chamber with a total volume of 1.10 ml described procedure (19). Briefly, a custom-made thermostatically con-
rolled (37°C) gas-tight Plexiglas chamber with a total volume of 1.10 ml was used. The chamber was filled with buffer solution (113.0 mM NaCl, 4.0 mM KCl, 27.2 mM NaHCO₃, 1.0 mM KH₂PO₄, 1.2 mM MgCl₂, 1.0 mM CaCl₂, 10.0 mM HEPES, 0.5 mM Ca lactate, 2.0 mM glutamine, and 50 U/ml streptomyacin, 298 ± 2 mosM, pH 7.40) and continuously stirred with an air-driven magnetic stirrer. Glucose concentration in the medium was 5.8 mM for cells from normoglycemic controls and 23.2 mM for cells from diabetic animals. O₂ content was measured by a modified O₂ sensor (model 500, Unisense, Aarhus, Denmark) calibrated with the air-equilibrated buffer solution as 228°C.

Table 1. MAP, total RBF, RVR, cortical RBF, in vivo renal O₂ consumption, and Tₙa per O₂ consumed in normoglycemic control and hyperglycemic diabetic rats acutely treated with vehicle or C-peptide.

<table>
<thead>
<tr>
<th>n</th>
<th>MAP, mmHg</th>
<th>RBF, ml/min</th>
<th>RVR, mmHg·min⁻¹·ml⁻¹</th>
<th>Cortical RBF, LU</th>
<th>O₂ Consumption, µmol/min</th>
<th>Tₙa/O₂ Consumed</th>
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</thead>
<tbody>
<tr>
<td>10</td>
<td>Baseline</td>
<td>119 ± 4</td>
<td>9 ± 1</td>
<td>15.0 ± 1.3</td>
<td>425 ± 26</td>
<td>12 ± 1</td>
</tr>
<tr>
<td></td>
<td>Vehicle</td>
<td>106 ± 4*</td>
<td>9 ± 1</td>
<td>13.5 ± 1.3*</td>
<td>409 ± 25</td>
<td>14 ± 2</td>
</tr>
<tr>
<td>8</td>
<td>Baseline</td>
<td>116 ± 2</td>
<td>8 ± 1</td>
<td>14.9 ± 1.5</td>
<td>402 ± 21</td>
<td>12 ± 2</td>
</tr>
<tr>
<td></td>
<td>C-peptide</td>
<td>108 ± 2*</td>
<td>9 ± 1</td>
<td>12.4 ± 0.9*</td>
<td>430 ± 33</td>
<td>16 ± 3</td>
</tr>
<tr>
<td>9</td>
<td>Baseline</td>
<td>122 ± 4</td>
<td>9 ± 1</td>
<td>14.3 ± 1.3</td>
<td>370 ± 19</td>
<td>17 ± 2†</td>
</tr>
<tr>
<td></td>
<td>Vehicle</td>
<td>119 ± 8</td>
<td>10 ± 1</td>
<td>12.8 ± 1.0*</td>
<td>375 ± 18</td>
<td>19 ± 4</td>
</tr>
<tr>
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<td>122 ± 3</td>
<td>10 ± 1</td>
<td>13.5 ± 1.8</td>
<td>382 ± 34</td>
<td>18 ± 2†</td>
</tr>
<tr>
<td></td>
<td>C-peptide</td>
<td>113 ± 3*</td>
<td>10 ± 1</td>
<td>12.2 ± 1.5*</td>
<td>403 ± 33</td>
<td>20 ± 3</td>
</tr>
</tbody>
</table>

Values are means ± SE. LU, laser units; MAP, mean arterial blood pressure; RBF, renal blood flow; RVR, renal vascular resistance; Tₙa, Na⁺ transport. *P < 0.05 vs. baseline in the same group. †P < 0.05 vs. normoglycemic at baseline.

RESULTS

All diabetic animals were hyperglycemic compared with normoglycemic controls (22.3 ± 1.4 vs. 5.8 ± 0.2 mM). Diabetic animals gained less weight than the age-matched normoglycemic controls (281 ± 5 vs. 330 ± 6 g). Left kidney weight increased in diabetic animals compared with normoglycemic controls (1.71 ± 0.03 vs. 1.24 ± 0.03 g).

Diabetic rats displayed increased GFR (Fig. 1), which was reduced by C-peptide. FF was elevated in diabetic animals (Table 1) and decreased in vehicle- and C-peptide-treated controls. However, MAP only decreased after C-peptide administration in the diabetic animals. Baseline Tₙa was 70% higher in diabetic animals than in controls (Fig. 3) and was only reduced by C-peptide in the diabetic group. Baseline fractional Na⁺ excretion was similar in all groups (Fig. 4), but C-peptide increased fractional Na⁺ excretion by 77% selectively in the diabetic group. Furthermore, fractional Li⁺ excretion increased only in the diabetic animals treated with C-peptide (Fig. 5).

There was no difference in total or cortical RBF in the diabetic animals compared with the normoglycemic controls (Table 1). C-peptide did not alter RBF in any of the groups. Total in vivo QO₂ was ~50% higher in the diabetic animals (Table 1), although QO₂ did not differ between diabetic and...
normoglycemic control rats during baseline conditions (Table 1). C-peptide did not affect total QO2 or TNa/QO2 (Table 1).

Infusion of vehicle did not alter any of the parameters in controls or diabetic animals, except for RVR, which decreased in all groups as a function of time, and TNa/QO2, which decreased in the normoglycemic controls (Table 1).

C-peptide reduced Pff by 5.2% (11.6 ± 0.2 vs. 11.0 ± 0.2 mmHg), Psf by 8.4% (41.9 ± 0.3 vs. 38.4 ± 0.3 mmHg), and calculated Pnet by 9.6% (30.3 ± 0.4 vs. 27.4 ± 0.3 mmHg) in the diabetic animals, whereas it had no effect in controls (Fig. 6).

Baseline QO2 was increased in isolated proximal tubular cells from diabetic animals compared with controls (39 ± 3 vs. 28 ± 2 nmol·mg protein⁻¹·min⁻¹; Fig. 7). C-peptide reduced QO2 in diabetic cells (28 ± 3 nmol·mg protein⁻¹·min⁻¹) but had no effect on cells from normoglycemic controls (26 ± 5 nmol·mg protein⁻¹·min⁻¹; Fig. 7). Pretreatment with L-NAME had no effect in any of the groups and did not alter the effect of C-peptide in the diabetic cells (Fig. 7).

**DISCUSSION**

The main new finding from the present study was that acute administration of C-peptide reduced GFR in diabetic rats via two seemingly different mechanisms. 1) C-peptide caused a net dilation of the efferent arteriole in the diabetic rats, as evident from decreased FF and Psf in the absence of altered RBF. 2) C-peptide inhibited tubular Na⁺ reabsorption in diabetic rats, as evident from increased fractional Na⁺ and Li⁺ excretions and reduced transport-dependent QO2 in isolated proximal tubule cells from diabetic rats.

Acute alterations in GFR are determined by the glomerular capillary hydrostatic pressure (Pcap), the hydrostatic pressure in Bowman’s space, and the oncotic pressure (ΔΠonc). Changes in Pcap are a result of alterations in the interplay of afferent and efferent arteriolar diameter, which alter the pressure transmitted to the glomerular capillaries (41, 43). Constriction of the afferent arteriole or dilation of the efferent arteriole will reduce Pcap and lower Pnet, which reduces GFR. Intratubular hydrostatic pressure is mainly determined by early proximal tubular reabsorption and, under certain conditions, by the hydraulic resistance in the distal nephron segments, e.g., obstruction of the tubules (22). ΔΠonc is determined by differences in the protein concentration between the blood in the glomerular capillaries and the fluid in Bowman’s space. The main intrarenal factor influencing acute changes in ΔΠonc is the FF (3, 38), which reflects the balance between the afferent and efferent arterioles during normal conditions. Reduced FF will decrease the concentration of proteins in the blood that leaves the glomerulus through the afferent arteriole and, thus, reduce the driving force for reabsorption along the proximal part of the nephron into the peritubular capillaries, commonly referred to as the glomerulotubular balance.

Diabetes-induced glomerular hyperfiltration is reduced by C-peptide in patients with type 1 diabetes (19, 20) and in animal models of insulinopenic diabetes (16, 36, 48, 49, 51), with concomitant reduction of diabetes-induced renal damage (14, 16, 48). The glomerular arterioles exert critical actions to...
control \(P_{\text{cap}}\), thereby influencing the GFR. We previously reported that C-peptide constricts isolated afferent glomerular arterioles from diabetic mice in vitro (35). A similarly pronounced constriction of the afferent arterioles in vivo with no effect on efferent arterioles would undoubtedly reduce GFR and RBF and increase RVR. However, C-peptide reduced the elevated GFR and FF in the diabetic rats in the present study, but did not affect RBF, which is consistent with previous reports (20, 49). None of these parameters were altered in the animals treated with vehicle only. Furthermore, RVR was not increased in the diabetic animals treated with C-peptide but, rather, decreased as a function of time in all groups. Taken together with our finding of reduced \(P_{\text{sf}}\) after C-peptide administration, this can only be attributed to a balancing dilation of the efferent arteriole. Similar dual actions have been reported for nitric oxide (NO) synthase inhibitors, \(Ca^{2+}\) channel blockers, atrial natriuretic peptide, and adenosine (1, 11, 15, 31). However, given the lowering of \(P_{\text{net}}\), this mechanism could have a significant effect on the outcome for diabetic patients, since it lowers GFR and, thus, the tubular \(Na^+\) load while maintaining RBF.

C-peptide may also decrease diabetic hyperfiltration through inhibition of \(Na^+\)-\(K^+\)-ATPase activity, as evident from the increase in fractional urinary \(Na^+\) and \(Li^+\) excretions and decreased transport-independent \(Q_{O_2}\) in isolated proximal tubular cells from diabetic rats. It has previously been postulated that diabetic hyperfiltration occurs as a result of primary hyperreabsorption upstream of the macula densa and the normal physiology of tubuloglomerular feedback (53, 54). However, it has been suggested that diabetes-induced glomerular hyperfiltration is caused by TGF-independent mechanisms (10, 46), although conflicting results have been reported (55).
studies utilize adenosine A1 receptor-deficient mice, which have been shown to lack the TGF mechanism, but additional TGF-independent effects of adenosine A1 receptors on afferent arteriolar tone complicate the interpretation of these findings. Also, the degree of hyperglycemia must be considered in a discussion of the potential role of TGF in diabetes-induced glomerular hyperfiltration (55). The diabetic rats displayed reduced proximal tubular \( \text{Na}^+/-\text{H}^+ \) reabsorption after C-peptide administration, and it is possible that the reduced GFR in these animals is caused by a TGF activation. Furthermore, earlier work demonstrated that tubular reabsorption directly influences hydraulic resistance in the proximal tubule and, therefore, affects GFR (30). It has been shown that sustained hyperglycemia in patients (56), as well as experimental animal models (27, 54), results in increased proximal tubular \( \text{Na}^+ \) reabsorption secondary to increased \( \text{Na}^+/-\text{H}^+ \)-linked glucose reabsorption. It has also been shown that early distal tubular \( \text{Na}^+ \) concentrations are decreased during hyperglycemia, together with increased cortical \( \text{Na}^+/-\text{K}^+/-\text{ATPase} \) activity (27, 50). Conversely, decreased \( \text{Na}^+ \) reabsorption in the early proximal tubule will increase hydraulic resistance, which in turn lowers \( P_{\text{net}} \) and, therefore, reduces GFR. This hypothesis is consistent with reports of a linear correlation between GFR and \( \text{Na}^+/-\text{K}^+/-\text{ATPase} \) activity in the kidney cortex during experimental diabetes (27, 50). Therefore, a reduced or normalized \( \text{Na}^+ \) reabsorption in the early proximal tubule is likely to reduce GFR in the diabetic kidney. The differences in the reduction of \( P_{\text{sf}} \) (−8.4%) and \( P_{\text{ff}} \) (−5.2%) in this study are not definite proof but indicate that proximal tubular reabsorption is involved in the decrease in GFR. No reduction was seen in total \( \text{Na}^+ \) excretion after C-peptide, a finding that is consistent with previous studies (42). However, the reduction in tubular \( \text{Na}^+ \) load, due to reduced GFR after C-peptide, will conceal the inhibitory effect on total urinary \( \text{Na}^+ \) excretion. The calculated fractional \( \text{Na}^+ \) and \( \text{Li}^+ \) excretions increased in the diabetic animals after C-peptide administration, which shows that C-peptide directly inhibits \( \text{Na}^+ \) reabsorption exclusively in the diabetic rats.

Electrolyte transport via \( \text{Na}^+/-\text{K}^+/-\text{ATPase} \) activity accounts for ∼80% of total QO2 in the kidney (52). Therefore, diabetes-induced increase in \( \text{Na}^+/-\text{K}^+/-\text{ATPase} \) activity is likely to influence total QO2 and, therefore, possibly limit renal O2 availability. The finding of increased cellular QO2 in diabetic rats in the present study is consistent with previous reports (27, 39). However, the present study shows that C-peptide directly inhibits QO2 in isolated proximal tubular cells but that the in vivo total kidney QO2 was not altered by C-peptide administration, even though \( T_{\text{Na}} \) decreased by ∼17%. A possible explanation can be a shift of the \( \text{Na}^+ \) reabsorption site along the nephron after inhibition of proximal tubular \( \text{Na}^+/-\text{K}^+/-\text{ATPase} \) activity. QO2/TNa is significantly less in the proximal tubule than in more distal parts of the nephron, since a notable part of the Na reabsorption in the proximal tubule is achieved via paracellular transport (29). This explanation would account for lack of reduced QO2 in the diabetic rats in vivo after C-peptide administration.
Na\textsuperscript{+}-K\textsuperscript{-}-ATPase is inhibited by ouabain, although complete inhibition cannot be achieved in rats (12, 34). Ouabain treatment reduced cellular QO\textsubscript{2} in both groups. C-peptide had no further effect on ouabain-treated cells, which indicates that C-peptide reduces Na\textsuperscript{+}-K\textsuperscript{-}-ATPase activity.

C-peptide reduced blood pressure in the present study, similar to previously reported findings (36), although most studies show no such effect. However, it is predicted that increased NaCl load to the macula densa will decrease renin release and, therefore, angiotensin II production (25), and it is possible that reduced tubular Na\textsuperscript{+}-K\textsuperscript{-}-ATPase activity may reduce long-term arterial blood pressure, which should further add to the renoprotective effects of C-peptide. However, the effect of long-term C-peptide administration on arterial blood pressure control in diabetic patients is beyond the scope of this study.

The beneficial effects of C-peptide on diabetes mellitus have been observed in various tissues (14, 17). However, on the molecular level, the data may seem contradictory. In the present study, C-peptide decreased the ouabain-sensitive QO\textsubscript{2} of cells from diabetic rats but did not elicit an effect in cells isolated from normoglycemic rats. The inhibitory effect of C-peptide on isolated diabetic proximal tubular cells persisted also when the cells where pretreated with a nonselective NO synthase blocker, indicating that the mechanism does not involve a stimulated NO release with concomitant inhibition of mitochondrial respiration. Several previous studies report stimulatory effects of C-peptide on Na\textsuperscript{+}-K\textsuperscript{-}-ATPase, but these studies were performed under normoglycemic conditions (37, 60). Furthermore, Na\textsuperscript{+}-K\textsuperscript{-}-ATPase activities in neurons and erythrocytes are reduced during diabetes, and C-peptide normalizes these impairments (8, 13, 17). However, Na\textsuperscript{+}-K\textsuperscript{-}-ATPase activity in the kidney is elevated during the early onset of diabetes, probably due to the increased tubular electrolyte load resulting from the elevated GFR (6, 27, 57, 58). We report evidence for a causal role in governing fluid reabsorption and glomerulotubular balance by the renal proximal tubule. J Clin Invest 50: 336–349, 1971.


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