Major role for ACE-independent intrarenal ANG II formation in type II diabetes

Sungmi Park,1 Benjamin J. Bivona,1 Hiroyuki Kobori,2 Dale M. Seth,2 Mark C. Chappell,3 Eric Lazartigues,4 and Lisa M. Harrison-Bernard1

Departments of 1Physiology and 4Pharmacology and Experimental Therapeutics, Louisiana State University Health Sciences Center and 2Department of Physiology, Tulane University Health Sciences Center, New Orleans, Louisiana; and 3Hypertension and Vascular Disease Center, Wake Forest University Health Sciences, Winston-Salem, North Carolina

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Recently, there has been growing interest in the role of ACE-independent ANG II production in various physiological and pathophysiologic states. ACE-independent enzymatic pathways include serine proteases, tonin, cathepsin G, trypsin, and kallikrein (38). Vascular chymase is a major serine protease (EC 3.4.21.39) implicated in the ACE-independent production of ANG II in human arteries (23, 31). Chymase, which cleaves ANG I at the same site as ACE, is completely inhibited by serine protease inhibitors; ACE inhibitors do not influence chymase activity (40). Markedly increased chymase expression in mesangial and vascular smooth muscle cells in human diabetic nephropathy (12), IgA nephropathy (33), and hypertensive nephropathy (44) has been reported. The involvement of renal mast cell chymase activity in ANG II generation has also been reported in patients with autosomal dominant polycystic kidney disease (24). Therefore, ACE-independent formation of ANG II may contribute significantly to progression of many forms of renal disease.

The db/db mouse (BKS.Cg-Dock7m +/+ Leprdb/J) is an animal model of type II diabetes which has a point mutation in the leptin receptor gene (22) that leads to hyperphagia, obesity, progressive hyperglycemia, hyperinsulinemia, and insulin resistance. Diabetic db/db mice exhibit progressive diabetic renal disease characterized by renal and glomerular hypertrophy, albuminuria, glomerulosclerosis, and mesangial matrix expansion, which are features of human diabetic nephropathy (3, 19, 47). Ye et al. (46) have demonstrated that renal cortical ACE protein expression is reduced, while ACE2 protein expression is elevated in diabetic db/db compared with control db/db mice. Elevated ACE2 protein expression is thought to provide a renoprotective effect on diabetic renal injury due to the ability of ACE2 to degrade ANG II and generate ANG1-7. ANG1-7 is a peptide with vasodilator and antiproliferative properties (21). The impact of altered ACE and ACE2 protein expression on intrarenal ANG II formation has not been determined in this model. We recently reported that the renal afferent arteriole vasoconstrictor responses to ANG II remain intact in db/db mice (28). However, the functional consequence of reductions in ACE enzyme activity on the intrarenal formation of ANG II blocker lowers blood pressure (2, 25) and provides greater protection in diabetic nephropathy (13, 27) than ACE inhibitor alone suggests that suppression of the RAS is incomplete. It has been suggested that dual blockade of RAS with inhibition of ACE and AT1 receptor blockade results in an additional reduction in proteinuria in patients with chronic kidney disease (5). Thus ACE inhibitor monotherapy may allow for the continued generation of ANG II via ACE-independent pathways.
from ANG I on the renal microvasculature of type II diabetes has not been previously investigated.

In the current study, we tested the hypothesis that there is a switch from renal ACE-dependent to ACE-independent ANG II formation in the progression of diabetic vascular disease. The leptin receptor deficient db/db animal model was utilized to study the renal complications of diabetes with an emphasis on afferent arteriole vascular function. Plasma ANG II and kidney and heart ANG I and ANG II levels were determined in control and diabetic mice. Also, kidney and heart ACE and ACE2 activity and protein expression, and urinary excretion of angiotensinogen, were measured. Studies on renal resistance vessel reactivity were carried out using the mouse in vitro blood-perfused juxtamedullary technique, which allows for direct videomicroscopic visualization of afferent arteriolar diameters studied in situ in kidneys obtained from control and diabetic mice. Renal afferent arteriolar vascular responses to the intrarenal conversion of ANG I to ANG II in the absence or presence of enzymatic inhibitors provided a mechanism for investigating the physiological responses to ACE-dependent and ACE-independent pathways for the formation of ANG II in the diabetic mouse kidney.

METHODS

Animals

The procedures used in this study were approved by the Institutional Animal Care and Use Committee of Louisiana State University Health Sciences and conducted according to the National Institutes of Health Guide for the Care and Use of Laboratory Animals. Experiments were performed in a total of 71 male (18 wk old) control db/db (Dock7m+/+ Lepr(db)) and 69 diabetic db/db (BKS.Cg-Dock7m+/+ Lepr(db/J)) mice littermates (stock no. 000642, The Jackson Laboratory, Bar Harbor, ME). Adult male Sprague-Dawley rats (50 ± 8 g body wt, n = 83; Charles River Laboratories, Raleigh, NC) were used as blood donors for the study of the mouse renal microvasculature. All animals were provided ad libitum access to food and water during the study except when food was removed during the time preceding fasted blood glucose measurements. Fasting blood glucose levels were measured in conscious diabetic (n = 27) and control (n = 24) mice housed in the home cage following 18 h of food removal (3 P.M.-9 A.M.). Onetouch Ultra 2 Blood Glucose Meter, Lifescan, Milpitas, CA). Measurements were taken ~1 wk before performance of the renal microcirculation experiments. Fed blood glucose measurements were also obtained in conscious diabetic (n = 20) and control (n = 20) mice on the morning of the preparation for renal microcirculation experiments. Mice were housed individually in metabolic cages specially designed for obese mice for a 3-day equilibration period. The 24-h urine samples were obtained for measurements of urinary angiotensinogen levels by ELISA (16) in diabetic (n = 8) and control (n = 10) mice.

ACE and ACE2 Activities

Kidney cortex, heart, and lung tissues were harvested from diabetic (n = 8) and control (n = 9) mice under pentobarbital sodium anesthesia (90 mg/kg ip). Tissues were homogenized briefly with a glass homogenizer in reaction buffer (30–50 mg of tissue in 300–400 μl of reaction buffer; 1 mM NaCl, 75 mM Tris, 0.5 mM ZnCl2, pH 7.5) and centrifuged at 14,000 g for 10 min. Protein concentrations were determined using a commercially available BCA kit (Pierce, Rockford, IL), and samples were stored at −20°C until assayed. Lung tissues from control mice were used for a positive control for ACE and negative control for ACE2 enzymatic activity. ACE and ACE2 activities were assessed using Fluorogenic Peptide Substrate VI [FPSVI; 7MCa-Y-V-A-D-A-PK (Dnp)-OH; R&D Systems, Minneapolis, MN] as we have previously described (7). Samples containing 100 μg of renal cortical, heart, or lung protein extract were incubated with 100 μM of FPSVI, 20 μM captopril (to inhibit ACE activity) or 1 μM DX600 (to inhibit ACE2 activity) in a final volume of 100 μl of reaction buffer to measure total ACE or ACE2 activity. Non-specific enzyme activity was measured by including the ACE inhibitor (10 μM captopril) and ACE2 inhibitor (1 μM DX600) in the reaction mixture. Fluorescence emission was monitored using a SpectraMax M2 Fluorescence Reader (Molecular Devices) at 10-min intervals for 2 h. Specific ACE activity was calculated by subtracting the activity in the presence of both 10 μM captopril and 1 μM DX600 from the total activity in the presence of 1 μM DX600. ACE2 activity was calculated by subtracting the activity in the presence of both 10 μM captopril and 1 μM DX600 from the total activity in the presence of 10 μM captopril. Data (arbitrary fluorescence units; AFU) were presented as amounts of substrate FPSVI converted to product per minute and were normalized for total protein (AFU·mg⁻¹·min⁻¹).

Blood was also collected from these anesthetized, open-chest mice via cardiac puncture using a 25-G needle and 1-ml syringe containing EDTA (200 mM) and centrifuged at 1,000 g for 10 min, and the plasma was stored at −20°C until assayed. Plasma leptin and insulin levels were measured using commercially available ELISA kits according to the manufacturer’s instructions (Mouse Leptin ELISA Kit EZML-82K, Linco Research; Merckodia Ultrasensitive Mouse Insulin ELISA 10-1150-01, respectively).

Renal ACE and ACE2 Immunohistochemistry

In vivo retrograde perfusion fixation (3% paraformaldehyde) was performed in anesthetized diabetic (n = 6) and control (n = 6) mice as we have previously described (11, 29). After fixation, the kidneys were routinely processed to paraffin and sectioned at 5 μm. Immunohistochemistry was performed on kidney sections incubated with the primary Ab or nonimmune serum, washed, and developed using the Vectastain ABC immunoperoxidase system as we have previously described (11, 29). Kidney sections from each animal were incubated with the following antibodies: 1) mouse ACE-specific goat IgG (1:1,000 and 1:2,000, AF1513; R&D Systems), and 2) mouse ACE2-specific rabbit IgG (9) (1:8,000 and 1:10,000). For densitometric analysis of the immunoreactivity for ACE (1:2,000) and ACE2 (1: 8,000) protein expression, 6–11 microscopic fields of renal cortex/tissue section/animal were obtained using ×40 objective and identical image exposure camera settings for all tissue sections with the microscopist blinded to the animal groups as we have previously described (29, 30). Combined tubular and endothelial ACE protein expression was quantified for intensity of immunostaining. Representative color micrographs were obtained from kidney sections immunostained for ACE (1:1,000) and ACE2 (1:10,000) using ×40 and ×100 objectives from control and diabetic kidneys.

Plasma ANG II and Tissue ANG I and ANG II Levels

Blood, kidneys, and hearts were harvested from conscious control (n = 11) and diabetic (n = 11) mice following decapitation. Trunk blood was collected into chilled tubes containing EDTA (5 mM), enalaprilat (20 μM), pepstatin A (10 μM), and 1,10-phenanthroline (1.25 mM) as we have previously described (14). Plasma was separated and stored at −20°C until assayed. Immediately after removal, kidneys and hearts were weighed and then homogenized in ice-cold methanol. ANG I and ANG II levels were quantitated by RIA as we previously reported (14).

Mouse In Vitro Blood-Perfused Juxtamedullary Nephron Technique

Experiments were conducted using the mouse in vitro blood-perfused juxtamedullary nephron technique as we have previously reported in detail (10, 28). Briefly, mice were anesthetized with...
pentobarbital sodium (90 mg/kg ip), the right renal artery was cannulated, the upper third of the cortex was removed, the distal arteries were tied, and the pelvic mucosa and connective tissues were removed, revealing the juxtamedullary nephrons. The kidney was placed on the stage of a microscope, and the afferent arteriole was visualized using a video camera, image enhancer, and recorded on a DVD. Kidneys were studied under euglycemic (5 mM = 110 mg/dl glucose) and hyperglycemic (30 mM = 540 mg/dl glucose) incubation conditions (5% BSA perfusion solution, 1% BSA superfusion solution, rat plasma) for control and diabetic mice, respectively (28). Donor blood was collected from anesthetized rats subjected to bilateral nephrectomy and exanguinated via a carotid arterial cannula. Rat plasma was spiked with glucose (4 mg of glucose/ml of plasma) to achieve a final concentration of 30 mM (526 ± 14 mg/dl, n = 18) for the perfusion of kidneys obtained from diabetic mice. The kidney was bathed with warmed Tyrode buffer containing 1% BSA and perfused with blood pressurized to 95 mmHg with 95% O2-5% CO2. Peptides and drugs were added to the 5% BSA perfusion solution and/or 1% BSA superfusion and/or rat plasma. A minimum of 5 min was allowed for equilibration of the renal vasculature upon initiation of the blood perfusion. Baseline afferent arteriolar diameters were measured during control conditions (1% BSA solution superfusion, 5 min). Each protocol was followed by a 15-min recovery period. Afferent arteriolar diameters were measured during the following protocols.

Protocol 1: ANG I. Afferent arteriolar diameters were measured during superfusion with ANG I to determine the vascular effects of intrarenal conversion of ANG I to ANG II in kidneys of diabetic (n = 10) and control (n = 12) mice. Kidneys were exposed to 1, 10, 100, and 1,000 nM human ANG I (catalog no. 002-01, Phoenix Pharmaceuticals) applied in the superfusion solution for 5-min periods each. Only one ANG I dose-response was performed in each kidney. ANG I substrate availability was not limiting in this experimental method since the rate of delivery of ANG I via the superfusion is not limiting. The effects of ANG I were determined in the same vessels at the conclusion of the experiment.

Protocol 2: ANG I in the presence of AT1 receptor blockade. Kidneys were superfused with the AT1 receptor antagonist (100 μM candesartan; kindly provided by AstraZeneca) for 10 min followed by 1, 10, and 100 nM ANG I for 5-min periods each in diabetic (n = 7) and control (n = 6) mice. Since it was expected that the AT1 receptor antagonist would block the vasoconstriction resulting from conversion of ANG I to ANG II, the vasoconstrictor response to 1 μM norepinephrine (3 min) was examined in the same vessels at the conclusion of the experiment.

Protocol 3: ANG I in the presence of ACE inhibition. Captopril (catalog no. C4042, Sigma) was used for ACE inhibition in control and diabetic mice. Mice were pretreated using an injection of captopril (100 mg/kg ip) 30 min before kidney harvesting (4). Captopril (1 mM final concentration) was also added to the perfusion and superfusion solutions to ensure continuous ACE blockade throughout the entire experiment. The effects of ANG I (1, 10, 100 nM, 5-min period each) in the presence of ACE inhibition on afferent arteriolar diameter were measured in diabetic (n = 6) and control (n = 6) mice. The response to 100 nM ANG II was examined in the same vessels at the conclusion of the experiment.

Protocol 4: ANG I in the presence of serine protease inhibition. PMSF (catalog no. P7626, Sigma) was used for inhibition of serine proteases in kidneys of diabetic and control mice. Mice were pretreated with an injection of PMSF (3 mg ip) 30 min before kidney harvesting, and PMSF (1 mM final concentration) was added to the perfusion and superfusion solutions to ensure continuous enzymatic blockade (40). The effects of ANG I (1, 10, 100 nM, 5-min period each) in the presence of serine protease inhibition on afferent arteriolar diameter were measured in diabetic (n = 6) and control (n = 6) mice. The response to 100 nM ANG II was determined in the same vessels at the conclusion of the experiment. The stock solution of PMSF was diluted in 100% anhydrous ethanol (final concentration in perfusion and superfusion solutions 0.33% ethanol). Diluent control experiments were performed to determine the effect of 0.33% ethanol on renal vascular reactivity to ANG I in diabetic (n = 8) and control mice (n = 8).

Protocol 5: ANG I in the presence of both ACE and serine protease inhibition. The effects of ANG I (1, 10, 100 nM, 5-min period each) in the presence of the combination of ACE and serine protease inhibition on afferent arteriolar diameter were measured in diabetic (n = 6) and control (n = 7) mice. The response to 100 nM ANG II was determined in the same vessels at the conclusion of the experiment.

Data Analyses and Statistics

Renal arterial perfusion pressure and vessel diameters were sampled at 1 Hz and converted to a digital form using analog-to-digital data-acquisition and -analysis software as we have previously described (28, 42). Afferent arteriolar luminal diameters were measured manually and continuously throughout the protocol at a single site along the length of the afferent arteriole using a digital image-shearing monitor. Afferent arterioles were studied at a site from the glomerulus of 25–50% along the length of the vessel. The average diameter (μm) during superfusion with ANG I to determine the vascular effects of intrarenal conversion of ANG I to ANG II in kidneys of diabetic (n = 10) and control (n = 12) mice.

RESULTS

Metabolic Parameters

Experiments were performed in 18-wk-old adult male control (Dock7m+/+Leprdb) and diabetic (Dock7m−/+ Leprdb/J) mouse littermates. Body weight and food and fasting blood glucose levels were significantly elevated in diabetic compared with control mice at 18 wk of age (Table 1). Fed blood glucose levels were significantly higher than fasting levels.

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<th>Table 1. Metabolic parameters of control and diabetic mice</th>
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Values are means ± SE. BW, body wt. *P < 0.05 vs. control. †P < 0.05 vs. fed. ‡P < 0.05 vs. left kidney.
blood glucose levels in both groups. As there was no negative feedback on plasma leptin levels due to loss of leptin receptor signaling, plasma leptin levels were significantly higher in diabetic compared with control mice. Plasma insulin levels were significantly higher in diabetic compared with control mice. There were no differences in kidney weight or tibia length between control and diabetic mice. Right kidney weight was significantly greater than left kidney weight in each group. Heart weight was significantly reduced in diabetic compared with control mice. These diabetic mice exhibited significant albuminuria (822 ± 365 vs. 28 ± 8 µg/day, n = 11–12) as we have published previously (28).

**ACE and ACE2 Activities in Renal Cortex, Heart, and Lung**

Renal cortical ACE activity was significantly decreased (Fig. 1A, 2.1 ± 0.8 vs. 9.2 ± 2.1 AFU·mg⁻¹·min⁻¹, n = 8, 8), while ACE2 activity was significantly increased (Fig. 1B, 16.7 ± 3.2 vs. 7.2 ± 2.4 AFU·mg⁻¹·min⁻¹, n = 8, 8) in diabetic compared with control mice. The ACE/ACE2 ratio in kidneys was significantly lower in diabetic compared with control mice (Fig. 1C, 0.13 ± 0.04 vs. 1.88 ± 0.92). However, ACE and ACE2 activities, and ACE/ACE2 ratios were similar in the hearts of diabetic and control mice (Fig. 1D, 6.1 ± 3.8 vs. 14.1 ± 4.5 AFU·mg⁻¹·min⁻¹, P = 0.20, n = 5; Fig. 1E, 11.4 ± 0.9 vs. 12.4 ± 5.0 AFU·mg⁻¹·min⁻¹; n = 4–5; Fig. 1F, 0.47 ± 0.37 vs. 1.36 ± 0.38). Lung ACE (29.9 ± 5.6 AFU·mg⁻¹·min⁻¹; n = 4) and ACE2 (4.1 ± 1.0 AFU·mg⁻¹·min⁻¹; n = 4) activities were measured in control mice. In tissues of control mice, the lung ACE/ACE2 ratio (9.8 ± 4.3) was significantly greater than the renal cortical ACE/ACE2 ratio.

**ACE and ACE2 Protein Immunohistochemical Localization and Quantitative Analysis**

ACE protein immunohistochemical localization was determined in kidney sections of control (Fig. 2, A–C, n = 6) and diabetic (Fig. 2, D–F, n = 5) mice. ACE protein expression is localized to the proximal tubule brush border and endothelial cells of both renal vasculature and glomerular capillaries of mouse kidneys (Fig. 2, A–F). Quantitative analysis of the density of ACE protein immunostaining (Fig. 2M) indicated that ACE protein expression (arbitrary units relative to control) is significantly reduced in the diabetic (0.4 ± 0.2) compared with control (1.0 ± 0.2) kidneys. ACE protein immunohistochemical localization was determined in kidney sections of control (Fig. 2, G–I, n = 6) and diabetic (Fig. 2, J–L, n = 6) mice. ACE2 protein expression is localized to the proximal tubule brush border (Fig. 2, G–L). Endothelial cells do not stain for ACE2 protein (Fig. 2, H and I). The number and intensity of ACE2 positively stained proximal tubules were qualitatively greater in diabetic compared with control kidneys. High-power magnification of the inset in Fig. 2J is illustrated in Fig. 2L. Note the intense ACE2 immunostaining in the proximal tubule brush border of the diabetic kidney. ACE2 immunostaining density (Fig. 2N) is significantly greater in diabetic compared with control kidneys (2.0 ± 0.3 vs. 1.0 ± 0.3).
Fig. 2. ACE (A–F) and ACE2 (G–L) immunohistochemical localization and quantitative analysis (M; n = 6, 5; N; n = 6, 6) of renal cortex of control (A–C, G–I, hatched bars) and diabetic (D–F, J–L, filled bars) mice. ACE and ACE2 proteins are localized to the proximal tubule brush border of control and diabetic kidneys. ACE is also expressed in endothelial cells of the renal vasculature and glomerular capillaries. The density (M) of renal cortical ACE protein immunostaining is significantly reduced in diabetic compared with control kidneys. However, ACE2 immunostaining density (N) is significantly greater in diabetic compared with control kidneys. Antibody dilutions for ACE and ACE2 antibodies were 1:1,000 and 1:10,000, respectively. Images were obtained from the midcortical region of the kidney using a ×40 (A, D, G, J) or ×100 oil-immersion lens (B and C, E and F, H and I, K and L). g, Glomerulus; arrows, endothelial cells; inset in J indicates higher magnification in L. Bar = 50 μm. *P < 0.05 control vs. diabetic.
Urinary Angiotensinogen Excretion, ANG I, and ANG II Peptide Levels

Urinary angiotensinogen excretion was significantly higher in diabetic compared with control mice (66 ± 16 vs. 12 ± 2 ng/day, \( n = 8 - 10 \)) (Fig. 3A) and was significantly correlated with urinary albumin excretion (\( r = 0.412, P < 0.01 \)). Kidney ANG I and ANG II levels were not different between control and diabetic mice (110 ± 45 and 788 ± 99 vs. 113 ± 24 and 1,017 ± 165 fmol/g, \( n = 7 - 11 \)) (Fig. 3, B and C). Plasma ANG II levels were similar in control and diabetic mice (391 ± 105 vs. 251 ± 26 fmol/ml, \( n = 10; P = 0.20 \)) (Fig. 3C). Cardiac ANG I and ANG II levels were not different between control and diabetic mice (112 ± 25 and 836 ± 83; 129 ± 43 and 965 ± 147 fmol/g, \( n = 4 - 10 \), respectively) (Fig. 3, B and C). Despite reduced ACE activity, ANG II levels in diabetic kidneys were not different compared with control kidneys.

Baseline Afferent Arteriole Diameters

To determine the influence of the type II diabetic condition on afferent arteriole tone, baseline diameters from all arterioles from each group were pooled for statistical analysis. Baseline afferent arteriole diameters of kidneys from diabetic mice averaged 14.3 ± 0.5 μm (\( n = 44 \)) and was significantly larger than in kidneys from control mice, which averaged 12.2 ± 0.3 μm (\( n = 45 \)).

Afferent Arteriole Vasoconstrictor Responses to ANG I

Figure 4A demonstrates the time course of the afferent arteriole vasoconstriction to 1, 10, 100, and 1,000 nM ANG I in kidneys of control and diabetic mice. ANG I produced a significant vasoconstriction of afferent arterioles of both groups. There was no significant difference in the waning effects (average response over the final 30 s of each dose) of ANG I (1, 10, 100, 1,000 nM) in control (−6 ± 2, −11 ± 2, −20 ± 3, −20 ± 3% of baseline) and diabetic (−2 ± 5, −4 ± 5, −25 ± 5, −15 ± 5% of baseline) kidneys, respectively. Figure 4B illustrates the average afferent arteriole responses plotted in micrometers, and Fig. 4C illustrates the average afferent arteriole responses plotted as the delta % of baseline to ANG I (1–1,000 nM) in kidneys from control and diabetic mice. Significant afferent arteriole vasoconstriction to 10, 100, 1,000 nM ANG I was observed in kidneys of control (−10 ± 2, −24 ± 4, −26 ± 3%, \( n = 12 \)) and diabetic mice (−10 ± 2, −29 ± 4, −28 ± 3%, \( n = 11 \)) (Fig. 4C). ANG I produced similar responses in afferent arterioles from control and diabetic mice (\( P = 0.3 \)). Significant peak afferent arteriole vasoconstriction to 10, 100, and 1,000 nM ANG I was observed in kidneys of control (−15 ± 3, −39 ± 7, −42 ± 6%) and diabetic (−28 ± 6, −47 ± 6, −54 ± 6%) kidneys (Fig. 4D). ANG I produced a significantly greater peak vasoconstriction in afferent arterioles from diabetic compared with control mice (\( P = 0.01 \)). The time from application of the peptide to the peak vasoconstrictor responses to 1, 10, 100, and 1,000 nM ANG I in control (158 ± 34, 148 ± 28, 110 ± 22, 40 ± 8 s) and diabetic (151 ± 29, 116 ± 15, 133 ± 22, 34 ± 5 s) kidneys was similar (\( P = 0.7 \)). The time to reach the peak vasoconstriction to 1,000 nM ANG I was significantly shorter than all of the lower doses of ANG I in kidneys of diabetic mice. The time to reach the peak vasoconstriction to 1,000 nM ANG I was significantly shorter than for 10 and 1 nM ANG I in kidneys of control mice. A maximal vasoconstriction was observed with 100 nM ANG I; 1,000 nM ANG I did not produce a further vasoconstriction. Afferent arteriole diameters of control and diabetic kidneys had not completely returned to the baseline values following the 15-min recovery period (93 ± 3 and 93 ± 2% of baseline, respectively). The afferent arteriole responses to ANG I are compared with the ANG II responses that we have previously published (28) in control (Fig. 5A) and diabetic (Fig. 5B) kidneys. Since the afferent arteriole responses to ANG II and ANG I were not significantly different between kidneys of control and diabetic mice, the data were collapsed. The EC50 values for ANG II and ANG I were 2.5 ± 0.5 (\( n = 21 \)) and 48.2 ± 11.1 nM (\( n = 20 \)), respectively. The EC50 values for ANG II and ANG I were significantly different (\( P = 0.0001 \)). There was a 20-fold shift to the right in the vasoconstrictor response to ANG I compared with ANG II. The ANG I dose-response could not be repeated in the same kidney due to a significant 40% reduction in the afferent arteriole response to the second application of ANG I in diabetic kidneys (−29 ± 6 vs. −17 ± 5%, first vs. second, at 100 nM ANG I, \( n = 6 \)). Two consecutive norepinephrine dose responses (100, 300, 1,000 nM) were performed with a 20-min recovery between the first and second applications in kidneys of C57BL6 mice (\( n = 6 \)). There was no significant difference in the magnitude of the afferent arteriole vasoconstriction between the
first and second applications of norepinephrine (−11 ± 5, −24 ± 6, −36 ± 6% and −6 ± 3, −26 ± 7, −41 ± 7%, first vs. second). Afferent arterioles exhibited full recovery to baseline diameters upon removal of norepinephrine from the bath.

Afferent Arteriole Responses to ANG I in the Presence of AT1 Receptor Blockade

The afferent arteriole vasoconstrictor response to ANG I was significantly attenuated by AT1 receptor blockade in both groups (Fig. 6A). Pretreatment with an AT1 receptor blocker (100 μM candesartan) did not produce a change from the baseline diameter in either group. In the presence of AT1 receptor blockade, afferent arteriole diameters were 1 ± 2 and −1 ± 3% of the baseline in control (n = 6) and 1 ± 2 and −2 ± 2% of baseline in diabetic (n = 7) mice in response to 10 and 100 nM ANG I, respectively. During the recovery periods, afferent arteriole diameters of control and diabetic kidneys were not significantly different than baseline diameters. Renal microvascular responses to 1 μM norepinephrine were determined in the same vessels that demonstrated attenuated vasoconstrictor responsiveness to ANG I in the presence of AT1 receptor blockade. Norepinephrine produced a rapid and significant vasoconstriction in afferent arterioles of control and diabetic kidneys (−52 ± 2, −52 ± 5%; n = 6–7, respectively) (Fig. 7A), and there was no significant difference between the groups.

Afferent Arteriole Vascular Responses to ANG I in the Presence of ACE Inhibition

The vasoconstriction produced by conversion of ANG I to ANG II was significantly attenuated by ACE inhibition in control mice (−1 ± 3, −3 ± 5% at 10 and 100 nM ANG I, respectively, n = 6) (Fig. 6B). However, in the kidneys of diabetic mice the average afferent arteriole vasoconstriction produced by cleavage of ANG I to ANG II was not altered by ACE inhibition (−15 ± 4, −20 ± 4% at 10 and 100 nM ANG I, respectively, n = 6) (Fig. 6B). There was a significant difference in the magnitude of the peak afferent arteriole vasoconstriction to ANG I in the presence of ACE inhibition between control (−10 ± 2, −10 ± 5%) and diabetic (−22 ± 4, −32 ± 4%) at 10 and 100 nM ANG I, respectively. There was a significant difference in the enzymatic conversion of ANG I to ANG II during ACE inhibition between kidneys of diabetic and control mice. Afferent arteriole diameters were not different from baseline at the end of the recovery periods in the two groups.
Afferent Arteriole Vascular Responses to ANG I in the Presence of Serine Protease Inhibition

To identify an ACE-independent pathway that might be responsible for the conversion of ANG I to ANG II in kidneys of diabetic mice, experiments were conducted using serine protease inhibition. The average afferent arteriole vasoconstriction produced by ANG I to ANG II was significantly attenuated by serine protease inhibition in diabetic mice (\( \frac{n_1}{n_2} \), \( \frac{n_3}{n_4} \% \) at 10 and 100 nM ANG I, respectively, \( n = 6 \)). There was a significant difference in the magnitude of the peak afferent arteriole vasoconstriction to ANG I in the presence of serine protease inhibition between control and diabetic mice. (Fig. 6C). There was a significant difference in the enzymatic conversion of ANG I under serine protease inhibition between the kidneys of diabetic and control mice. The afferent arteriole vasoconstrictor response due to the conversion of ANG I to ANG II was not affected by the solvent used for dissolving PMSF (0.33% EtOH; \( n = 8 \)). There was no significant difference in the effect of 0.33% EtOH on the

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conversion of ANG I to ANG II compared with vasoconstrictor response to ANG I alone in kidneys of diabetic (n = 8) and control (n = 8) mice.

**ANG I in the Presence of both ACE and Serine Protease Inhibition**

Afferent arteriole vasoconstriction produced by conversion of ANG I to ANG II was significantly attenuated by the combination of ACE and serine protease inhibition compared with the responses to ANG I alone in kidneys of diabetic and control mice (Fig. 6D). There was no significant difference in the responses to ANG I in the presence of combined blockade of ACE and serine protease between diabetic and control kidneys. Afferent arteriole diameters following the recovery period were not different compared with baseline diameters in both groups.

In summary, in control mice, average afferent arteriole vasoconstriction produced by conversion of ANG I to ANG II was significantly attenuated by AT1 receptor blockade, ACE inhibition, and combined ACE and serine protease inhibition (−1 ± 3, −3 ± 5, and −5 ± 5% of baseline at 100 nM ANG I), but not by serine protease inhibition alone (−22 ± 4%). Importantly, the average afferent arteriole vasoconstriction produced by conversion of ANG I to ANG II was not altered by ACE inhibition (−20 ± 3% of baseline at 100 nM ANG I) in diabetic mice. Afferent arteriole vasoconstriction produced by cleavage of ANG I to ANG II was prevented by AT1 receptor blockade, serine protease inhibition, and combined ACE and serine protease inhibition (−2 ± 2, −6 ± 6, and −6 ± 3% of baseline at 100 nM ANG I) in kidneys of diabetic mice.

A significant afferent arteriole vasocstriction to a single dose of ANG II (100 nM) was observed in control (−15 ± 2%, n = 11) and diabetic (−13 ± 2%, n = 12) kidneys in the absence of enzymatic inhibitors (Fig. 7B). At the conclusion of the enzyme inhibitor experiments, afferent arteriole contractile responsiveness to ANG II remained intact in control (−15 ± 3%, n = 18) and diabetic (−14 ± 3%, n = 16) kidneys (Fig. 7B). The magnitude of the ANG II-induced vasoconstriction in the absence or presence of pharmacological inhibitors of ACE and/or serine protease was not significantly different between kidneys of control and diabetic mice.

**DISCUSSION**

The most significant finding of the present study is the identification of serine proteases as the major pathway for the formation of ANG II in the type II diabetic leptin receptor-deficient mouse kidney. Intrarenally formed ANG II from the substrate ANG I produces a potent afferent arteriole vasoconstriction in the presence of ACE inhibition in the diabetic kidney. However, in the normal kidney, afferent arteriolar vasoconstriction to intrarenally formed ANG II is blocked by ACE inhibition. Inhibition of serine proteases abolished the afferent arteriole response to intrarenal conversion of ANG I to ANG II in the diabetic kidney, but not the control kidney. These studies provide a potential explanation for the mechanism involved in the superior effects of combining an ACE inhibitor with an AT1 receptor antagonist relative to ACE inhibitor therapy alone to provide additional protection from diabetic nephropathy in humans (1, 13). In addition, the presence of this ACE-independent pathway for ANG II formation may explain the continued proteinuria in some patients on maximal ACE inhibitor therapy (5). We suggest that ACE inhibitor monotherapy may allow for the continued generation of ANG II via these ACE-independent pathways.

The experimental model selected to perform these studies involves the adult (18 wk) male type II diabetic mouse which exhibits obesity, hyperglycemia, hyperleptinemia, hyperinsulinemia, and albuminuria. Renal hypertrophy was not observed, although heart weight was reduced in our study. Previous studies have indicated that renal hypertrophy is only observed in younger (8 wk) diabetic db/db mice (34). The changes in body weight and organ weight were not related to a change in tibial length, suggesting that the growth of diabetic mice is not different from control mice.

Recently, Ye et al. (46) reported that renal cortical ACE activity and protein expression are reduced, while ACE2 activity is elevated in young, female diabetic db/db mice which do not display signs of diabetic renal disease. These alterations in ACE and ACE2 activity were not observed in the diabetic heart (46). Additionally, proximal tubule immunohistochemical protein expression of ACE was reduced, while ACE2 protein expression was elevated in diabetic compared with control mice (46). We performed the present study to determine whether an imbalance of intrarenal ACE and ACE2 also exists in the older male diabetic mouse with renal disease. In agreement with Ye et al., we also found that renal cortical ACE activity was reduced in kidneys of diabetic compared with control mice, while renal cortical ACE2 activity was elevated. There was no difference in ACE or ACE2 activity in the heart between the groups, indicating that this effect may be kidney specific. Wysocki et al. (45) have demonstrated that renal cortical ACE protein expression is reduced, while ACE2 protein expression is elevated in streptozocin-induced type I diabetic mice compared with control mice (37). Therefore, the pattern of change in intrarenal ACE and ACE2 is similar in mouse models of types I and II diabetes and suggests that hyperglycemia may be the common regulator. In addition, Ferrario et al. (8) have shown that inhibition of ACE results in augmented renal cortical ACE2 activity. Since ACE is involved in the formation of ANG II from ANG I while ACE2 degrades ANG II to ANG1-7, one might predict that a decreased ACE/ACE2 ratio might result in lower intrarenal ANG II peptide levels in diabetes. However, kidney ANG peptide levels were not measured in these earlier studies.

We performed the next series of experiments to determine the impact of reduced ACE activity and elevated ACE2 activity on intrarenal ANG II peptide levels in the diabetic kidney. To our surprise, ANG I and ANG II levels in the kidney were not different between control and diabetic mice. Plasma ANG II levels and cardiac ANG I and ANG II levels were also similar in control and diabetic mice. Wei et al. (43) have shown that although plasma ANG II is undetectable and kidney ACE activity is significantly reduced in ACE null mice, the elevation in ACE-independent (likely chymase) ANG II formation produces kidney ANG II levels that are not different from wild-type mice. However, Campbell et al. (4) demonstrated a significant suppression of ANG II in the kidney of mice with reduced somatic ACE gene expression. The data of Wei et al.
glomerulus, and/or interstitium may act in an autocrine/para-
dent pathways within the renal endothelium, microvasculature,
kidneys. ANG II synthesized via ACE and/or ACE-indepen-
dent pathways for ANG II formation may predominate
in the diabetic kidney.

Additionally, the urine concentration of angiotensinogen
was determined in our study. We found a significant increase in
the urinary excretion of angiotensinogen in diabetic compared
with control mice, suggesting that substrate availability is
increased in the diabetic kidney and may support ACE-indepen-
dent formation of ANG II. Singh et al. (35) have shown that
increased formation of ANG II in glomerular mesangial cells
cultured in high glucose is mediated by increased angiotensino-
gen and ANG I levels. Miyata et al. (26) have shown increased
intrarenal angiotensinogen protein levels in type II diabetic
rats. Also, in the present study, urinary excretion of angio-
tensinogen was significantly correlated with urinary excretion
of albumin, suggesting that the rise in intrarenal production of
angiotensinogen correlates with the severity of the renal dis-
ease. Although urinary protein excretion in deoxycorticoste-
rone acetate salt-induced volume-dependent hypertensive rats
was increased to the same or greater extent than in ANG
II-dependent hypertensive rats, urinary angiotensinogen excre-
tion was significantly lower in volume-dependent than in ANG
II-dependent hypertensive rats and was not greater than in
control rats (17). Kobori et al. (18) have demonstrated that
urinary angiotensinogen levels were enhanced in chronic kid-
ney disease patients compared with control subjects. The
source of urinary angiotensinogen is thought to be from an-
giotensinogen that is formed and secreted by the proximal
tubules and not from plasma (15). Therefore, our data are
consistent with recent studies indicating that urinary angio-
tensinogen levels could be a new therapeutic diagnostic test for
the progression of diabetic renal disease.

Renal hemodynamic dysfunction plays a central role in the
pathophysiological consequences of diabetic renal disease. We
have previously reported that db/db diabetic mice display
normal blood pressure and reductions in circadian blood pres-
sure amplitude, heart rate, and locomotor activity (28). In
addition, renal microvascular vasoconstrictor responses to
bath-applied ANG II were not altered in diabetic compared
with control mice (28). The afferent arteriole responses to
ANG II were not significantly different in kidneys of control
and diabetic kidneys incubated under acute euglycemic or
hyperglycemic conditions (28). However, the vasoconstrictor
response to bath-applied ANG I has not been previously
reported in juxtamedullary afferent arterioles of rodent kid-
nies. In our studies, the baseline afferent arteriole diameter
was significantly larger in diabetic compared with control
kidneys as we have previously reported (28). Intrarenal
conversion of ANG I to ANG II by ACE and/or ACE-independent
pathways produced a rapid and potent dose-dependent afferent
arteriole vasoconstriction in diabetic and control mice. The
average afferent arteriole responses to conversion of ANG I to
ANG II were not different between diabetic and control kid-
nies, although the magnitude of the peak response to ANG I
was significantly greater in the diabetic compared with control
kidneys. ANG II synthesized via ACE and/or ACE-indepen-
dent pathways within the renal endothelium, microvasculature,
glomerulus, and/or interstitium may act in an autocrine/para-
crine manner via binding to plasma membrane AT1 receptors
located on afferent arteriolar vascular smooth muscle cells. The
afferent arteriole responses to intrarenal conversion of ANG I
to ANG II are mediated by AT1 receptors, as evidenced by
complete blockade of the responses in the presence of candes-
sartan in diabetic and control mice. The time to reach peak
afferent arteriole vasoconstriction to ANG I was not different
between diabetic and control kidneys. This may indicate that
the AT1 receptor intracellular signaling response time is the
same and/or enzymatic rates of conversion of ANG I to ANG
II via ACE-dependent and ACE-independent pathways are
similar. We found rapid and significant arteriolar vascular
contractile response to norepinephrine in the presence of AT1
receptor blockade. Therefore, we document a maintained vas-
cular contractile responsiveness of the arterioles which lacked
a response to ANG I in the presence of AT1 receptor blockade.
Future studies will determine the major enzymatic pathways
for ANG II formation in the efferent arterioles of diabetic and
control mice.

It is generally thought that ACE is the primary route of ANG
II formation in the kidney. Campbell et al. (4) have demon-
strated a 97 and 92% reduction in plasma and kidney ANG II
levels, respectively, following acute ACE inhibitor treatment
in mice, indicating that ACE is the predominant pathway of ANG
II formation in blood and tissues of normal mice. In agreement
with these reports, we found that afferent arteriole vasocon-
striction produced by intrarenal conversion of ANG I to ANG
II was blocked by ACE inhibition in control kidneys. Interest-
ingly, ACE inhibition did not affect afferent arteriole vasocon-
striction produced by intrarenal conversion of ANG I to ANG
II in diabetic kidneys. Inhibition of conversion of ANG I to
ANG II by captopril indicates that ACE is the predominant
pathway for ANG II formation in the normal mouse kidney.
Our findings support a novel role of ACE-independent forma-
tion of ANG II in the diabetic kidney.

In the present study, afferent arteriole vasoconstriction pro-
duced by ANG I to ANG II was significantly attenuated by
serine protease inhibition in kidneys of diabetic mice, but not
in control mice. Afferent arteriole vasoconstriction produced by
intrarenal conversion of ANG I to ANG II was attenuated
by the combination of ACE and serine protease inhibition
in both diabetic and control kidneys. These data support our
hypothesis that serine proteases are the major enzymatic path-
way for ANG II formation in diabetes.

A review by Urata et al. (41) carefully outlines the involve-
ment of ACE inhibitor-insensitive ANG II formation in the
cardiovascular system and highlights the importance of these
pathways for the mechanisms that contribute to pathophysi-
ological changes in cardiovascular diseases. Less is known
about ACE-independent formation of ANG II in renal disease.
Urata et al. have demonstrated that in normal human serum,
ANG II formation is completely inhibited by the ACE inhibitor
captopril, but not by SBTI or PMSF, indicating that ACE is the
main enzyme involved in plasma ANG II formation. Interest-
ingly, the majority of the ANG II-forming activity in the
human heart is not inhibited by ACE inhibitors or aprotinin,
but is inhibited by the serine protease inhibitors SBTI or
PMSF, suggesting that chymase is the main enzyme involved
in ANG II formation in the human heart in vitro. It has been
shown that the formation of ANG II from ANG I in human
coronary arteries is dependent on both ACE (captopril inhib-
itable) and ACE-independent (chymostatin inhibitable) path-
ways (23). Seventy percent of the vasoconstrictor effect of ANG I is due to ACE, while serine proteases account for the remaining 30% in rat afferent and muscular efferent arterioles (39). Therefore, formation of ANG II by serine proteases contributes a minor effect on pre- and postglomerular responses in the rat kidney. Our findings of a major influence of ACE-independent formation of ANG II in the diabetic kidney are consistent with the predominant ACE-independent ANG II-forming pathways in the human heart.

Alternative pathways for converting ANG I to ANG II that do not require ACE have been recognized for a long time. There is accumulating evidence in humans and experimental animals of a major contribution of ACE-independent ANG II formation in the heart, vasculature, and kidney. Studies carried out in cultured myocytes, vascular smooth muscle cells, mesangial cells, and podocytes suggest that high glucose is a regulator of ACE-dependent and ACE-independent ANG II formation. ANG II peptide formation was increased in concert with increased protein expression of angiotensinogen, renin, and chymase in myocytes exposed to high glucose, although ACE protein expression was not altered (36). High glucose decreased expression of ACE, downregulated ACE-dependent ANG II generation, and upregulated chymase-dependent ANG II generation in rat vascular smooth muscle cells (20). Singh et al. (35) have shown increased formation of ANG I and ANG II in glomerular mesangial cells cultured in high glucose. Durvasula and Shankland (6) recently reported that hyperglycemia-induced activation of chymase-dependent ANG II formation in podocytes may promote progressive podocyte injury and loss in diabetic nephropathy. Activation of ANG II formation by ACE-independent mechanisms by high-glucose conditions indicates the need for new therapeutic strategies for treatment of vascular complications in diabetes.

In the kidney, markedly increased chymase expression and upregulated ACE expression in mesangial and vascular smooth muscle cells in human diabetes have been reported (12). The involvement of renal mast cell chymase activity in ANG II generation has also been reported in patients with autosomal dominant polycystic kidney disease (24). Upregulation of ACE-independent ANG II production is observed in the clipped kidney, but not the nonclipped kidney of renal vascular hypertensive rats, contributing to the maintenance of similar ANG II levels in clipped and nonclipped kidneys (32). These data suggest that ACE-independent pathways may be activated in the absence of hyperglycemia and may be driven by ischemia. Most importantly, our data represent that serine protease inhibition produced a complete blockade of the ANG I-to-ANG II-induced vasoconstriction in diabetic kidney. These data indicate a major pathway for serine protease-dependent ANG II formation in diabetic kidneys. Therefore, the concept of ACE-independent production of ANG II in pathogenic conditions involving the heart, vasculature, and kidney seem to solidify the concept that inhibition of these pathways may lead to new effective therapeutic approaches.

In summary, we demonstrated that ACE-independent mechanisms and increased intrarenal synthesis of angiotensinogen contribute to the intrarenal production of ANG II in diabetes. In kidneys of control mice, afferent arteriole vasoconstrictor responses are mediated by ACE-dependent conversion of ANG I to ANG II. In contrast, afferent arteriole vasoconstrictor responses to conversion of ANG I to ANG II in kidneys of diabetic mice were not altered by ACE inhibition. Interestingly, in the diabetic kidney, afferent arteriole vasoconstriction is mediated by serine protease-dependent conversion of ANG I to ANG II. Our future studies will determine whether chymase is responsible for serine protease-dependent ANG II formation in the diabetic kidney. Therefore, taken together, our findings in the diabetic kidney of reduced renal ACE activity and protein expression, maintained renal ANG I and ANG II peptide levels, elevated intrarenal formation of angiotensinogen, combined with functional afferent arteriolar vasoconstrictor responses to intrarenally produced ANG II from ANG I which are serine protease inhibitable and unresponsive to ACE inhibition, we conclude that serine protease-dependent formation of ANG II may be a new pharmacotherapeutic target for reducing diabetic renal vascular disease.

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DISCLOSURES

No conflicts of interest are declared by the authors.

REFERENCES


