Angiotensin type 2 receptor actions contribute to angiotensin type 1 receptor blocker effects on kidney fibrosis

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Naito T, Ma L-J, Yang H, Zuo Y, Tang Y, Han JY, Kon V, Fogo AB. Angiotensin type 2 receptor actions contribute to angiotensin type 1 receptor blocker effects on kidney fibrosis. Am J Physiol Renal Physiol 298: F683–F691, 2010. First published December 30, 2009; doi:10.1152/ajprenal.00503.2009.—Angiotensin type 1 (AT1) receptor blocker (ARB) ameliorates progression of chronic kidney disease. Whether this protection is due solely to blockade of AT1, or whether diversion of angiotensin II from the AT1 to the available AT2 receptor, thus potentially enhancing AT2 receptor effects, is not known. We therefore investigated the role of AT2 receptor in ARB-induced treatment effects in chronic kidney disease. Adult rats underwent 5/6 nephrectomy. Glomerulosclerosis was assessed by renal biopsy 8 wk later, and rats were divided into four groups with equivalent glomerulosclerosis: no further treatment, ARB, AT2 receptor antagonist, or combination. By week 12 after nephrectomy, systolic blood pressure was decreased in all treatment groups, but proteinuria was decreased only with ARB. Glomerulosclerosis increased significantly in AT2 receptor antagonist vs. ARB. Kidney cortical collagen content was decreased in ARB, but increased in untreated 5/6 nephrectomy, AT2 receptor antagonist, and combined groups. Glomerular cell proliferation increased in both untreated 5/6 nephrectomy and AT2 receptor antagonist vs. ARB, and phosphorylated Erk2 was increased by AT2 receptor antagonist. Plasminogen activator inhibitor-1 mRNA and protein were increased at 12 wk by AT2 receptor antagonist in contrast to decrease with ARB. Podocyte injury is a key component of glomerulosclerosis. We therefore assessed effects of AT1 vs. AT2 blockade on podocytes and interaction with plasminogen activator inhibitor-1. Cultured wild-type podocytes, but not plasminogen activator inhibitor-1 knockout, responded to angiotensin II with increased collagen, an effect that was completely blocked by ARB with lesser effect of AT2 receptor antagonist. We conclude that the beneficial effects on glomerular injury achieved with ARB are contributed to not only by blockade of the AT1 receptor, but also by increasing angiotensin effects transduced through the AT2 receptor.

podocyte; plasminogen activator inhibitor-1

ANGIOTENSIN II (ANG II) acts at two major receptors, type 1 (AT1) and type 2 (AT2). Classic ANG II effects are mediated through AT1, including vasoconstriction, sodium reabsorption, sympathetic nerve stimulation, aldosterone secretion, and inhibition of renin synthesis (46). ANG II also induces cell growth, oxidative stress, inflammation, and matrix accumulation through upregulation of molecules such as transforming growth factor-β (TGF-β) and plasminogen activator inhibitor-1 (PAI-1) (20). Human and experimental chronic kidney diseases (CKD) are characterized by progressive renal fibrosis. Inhibition of angiotensin actions, whether by angiotensin-converting enzyme inhibitor (ACEI) or AT1 receptor blocker (ARB), has been effective in ameliorating progression of CKD beyond effects on blood pressure (36). These beneficial effects have largely been linked to AT1 action blockade. However, beneficial effects of ARB could also be contributed to by increased levels of unbound ANG II after AT1 blockade being diverted to and activating the AT2 receptor. We sought to determine whether AT2 actions could play a role in these beneficial effects of ARB in a model of CKD widely studied and with relevance to human CKD, namely the renal ablation model (8). Pivotal studies from this model paved the way for understanding of the beneficial effects of ACEI in lowering glomerular hemodynamic stresses, and ameliorating or even reversing pathogenic events beyond hemodynamics, including direct effects to decrease matrix accumulation, and dampen mesangial, endothelial, and podocyte injuries (20, 21). Of note, these actions of the ARB and ACEI to ameliorate progressive kidney disease have been observed in human CKD even in patients classically regarded as having a low renin state, such as African-American patients with arterionephrosclerosis (51). These findings highlight the importance of local activation and interactions of components of the renin-angiotensin system, rather than systolic blood pressure (SBP) or peripheral renal levels, in mediating effects on sclerosis (51).

The role of the AT2 receptor in renal injury has been less defined than that of the AT1 receptor. The AT2 receptor counteracts the vasoconstrictor effect of the AT1 receptor (11, 26, 47), inhibits cellular proliferation, and promotes apoptosis (10, 27, 25, 38). However, whether these actions are also active in vivo in CKD is controversial. AT2 receptor transgenic mice had less sclerosis than wild-type (WT) mice after 5/6 nephrectomy (Nx), a model of CKD. Furthermore, specific overexpression of a novel molecule that specifically mediates some AT2 receptor actions, namely the ANG II type 2 receptor-interacting protein 1 (ATIP1), resulted in protection of mice against vascular injury (22). Correspondingly, sclerosis was worse in AT2−/− mice (5, 25), and interstitial fibrosis was also worsened with AT2 receptor antagonism (40). On the other hand, AT2 receptor blockade was renoprotective when given in the early phase of 5/6 Nx in rats (9). The balance of AT1 vs. AT2 activation also modulates NF-κB and thus may modify net injury (17). In this study, we therefore investigated the role of the AT2 in the beneficial effects induced by ANG type 1 receptor blocker on remodeling of existing CKD in the widely studied renal ablation model, which importantly has relevance for progression of human CKD (3, 4, 7).

MATERIALS AND METHODS

Adult male Sprague-Dawley rats (200–250 g; Charles River, Nashville, TN) were studied. All protocols were approved by the Institutional Animal Care and Use Committee according to the National...
Detection Kit; Chemicon, Temecula, CA). Positive control slides were according to manufacturer's directions (ApopTag Peroxidase In Situ Apoptosis labeling (TUNEL), which may also detect preapoptotic cells, followed the positive nuclei per glomerulus were counted and averaged for all

antisera instead of primary antibody showed no staining. PCNA-mouse antibody (Dako), or rabbit anti-rat PAI-1 antibody (American Diagnostica), Rochester, NY). Creatinine clearance was calculated from 24-h 12 wk by Vitros CREA slides (Johnson & Johnson Clinical Diagnostics, Rochester, NY). In situ hybridization. 35S-labeled sense and antisense riboprobes for PAI-1 were prepared as previously described (41). Negative control in situ hybridization was done with sense probes and showed no signal. PAI-1 mRNA was assessed by image analysis using SCION Image Beta 4.03 (Scion, Frederick, MD). Briefly, at least eight nonoverlapping random ×200 dark fields in the cortex were assessed for positive signals calculated as percent area of field occupied by PAI-1 mRNA signal, and average was calculated for each section. All sections were examined without knowledge of the treatment protocol.

Gene expression studies. Total RNA was isolated from kidney cortex using the RNeasy mini kit according to the manufacturer’s instructions (Qiagen). RNA was reverse transcribed using the TaqMan reverse transcription kit (Applied Biosystems). Real-time PCR amplifications were performed on an ABI Prism 7000 Sequence Detection System (Applied Biosystems) using TaqMan Universal PCR Master Mix (Applied Biosystems). Following inventoried TaqMan assays, IDs were used: Rn01435427_m1 for rat AT1a, Rn02132799_s1 for Rat AT1b, and Hs99999901_s1 for 18s ribosome. 18s rRNA was used as internal control. The relative levels of mRNA expression for these genes were obtained as previously described (32).

Western blot analysis. Tissue protein extracts were analyzed with rabbit anti-rat PAI-1 antibody (American Diagnostica) and rabbit anti-phospho-p44/42 MAP kinase (Erk1 and Erk2) antibody (Cell Signaling Technology, Beverly, MA) as primary antibodies with appropriate secondary antibody and visualized using an ECL Western Blotting Detection Reagents (Amerham Biosciences). Membranes were reprobed with mouse anti-β-actin (Sigma) and mouse anti-MAP kinase 2 (Erk2; Upstate, Lake Placid, NY) as loading controls and expression was semiquantitatively assessed by SCION Image Beta 4.03 (Scion) as a densitometric ratio relative to each loading control.

Collagen content. Renal cortex was isolated and frozen, and collagen content was calculated from the concentration of hydroxyproline, measured as their phenylisothiocyanate derivatives by reverse-phase high-performance liquid chromatography (RP-HPLC) (35). Briefly, tissue homogenates were hydrolyzed in vacuo in constant boiling HCl (110°C, 16 h) and were dried and taken up in 1 ml of redrying solution [ethanol/water/(C2H5)2N; 2:2:1], and 100 µl were redried under nitrogen. Twenty microliters of derivatization reagent [ethanol/water/(C2H5)2N/phenyliothiazoy-cyanate, 7:1:1:1] were added, and the reaction was allowed to proceed at room temperature for 20 min. The sample was then lyophilized during centrifugation to remove the reagents. Samples were either frozen at this stage or diluted to 1 ml with buffer (5% acetonitrile in 5 mmol/l disodium phosphate, pH 7.4). A solution containing amino acid standards was dried and coupled in the same manner. The coupling efficiency was >99% under these conditions. Collagen content was expressed as percent of total protein.

Podocyte culture. Podocyte injury is crucial for progression of glomerulosclerosis. We therefore studied the effects of AT1 vs. AT2 blockade on injury induced by Ang II in cultured podocytes. Our in vivo studies pointed to a key role of PAI-1 in glomerulosclerosis. We therefore also investigated the role of PAI-1 and interaction with AT1 vs. AT2 in podocyte injury. To achieve these goals, we cultured podocytes from WT and PAI-1/−− mouse kidneys as previously described (34). Podocyte identity was confirmed by WT-1 and synaptopodin staining (34). WT and PAI-1/−− podocytes were preincubated for 24 h with ARB (10−5 M) or AT1-RA (10−5 M) or both or left untreated and then exposed to Ang II (10−7 M) for 48 h in the presence of ARB, AT1-RA, or both, and compared with control cells without Ang II or additional treatment. Doses were based on a previous study in cultured cells (23, 31). Collagen α1(IV) accumulation in the supernatant was chosen as a marker of profibrotic injury to the podocyte and was assessed by ELISA. Activation of the Erk pathway was assessed in WT podocytes. WT podocytes were serum-
starved (control) or serum-starved with additional treatment for 15 min with ANG II with added ARB, AT2RA, or combination in doses as above as indicated.

Statistical analysis. Data are expressed as means ± SE. Differences between groups were examined for statistical significance using one-way ANOVA or Mann-Whitney U-test. P values <0.05 were considered significant.

RESULTS

SBP and renal function. Body weight was not different among the groups at any time (Table 1). SBP tail-cuff plethysmograph, 24-h proteinuria, and serum creatinine were by study design equivalent among groups before treatments began 8 wk after 5/6 Nx, a time point of established early sclerosis in this model. SBP at 12 wk was decreased only numerically in AT2RA alone and significantly in ARB and combined groups vs. the untreated 5/6 Nx group (untreated 5/6 Nx 206.6 ± 3.5, ARB 157.1 ± 12.8, AT2RA 180.3 ± 12.9 mg/dL, combined 164.3 ± 4.8, untreated 5/6 Nx vs. ARB P < 0.05, untreated 5/6 Nx vs. combined P < 0.05; Table 1). Proteinuria at 12 wk was decreased vs. the untreated 5/6 Nx group in ARB and combined groups but not in AT2RA-treated rats. Proteinuria was decreased by ARB, and in contrast remained high in untreated 5/6 Nx rats and in AT2RA-treated rats (untreated 5/6 Nx 463.2 ± 39.1, ARB 199.3 ± 48.9, AT2RA 386.5 ± 34.8, combined 242.1 ± 79.5 mg/day, untreated 5/6 Nx vs. ARB P < 0.01, untreated 5/6 Nx vs. combined P < 0.05, ARB vs. AT2RA, P < 0.05; Table 1). Serum creatinine and creatinine clearance after 4 wk of treatment were not significantly different between groups, mirroring our previous experience in this model and that of others (see Fig. 3). CCR untreated 5/6 Nx 0.8 ± 0.2, ARB 0.8 ± 0.3, AT2RA 0.6 ± 0.1, combined 0.6 ± 0.2 ml/min) (1). These relatively insensitive measures of glomerular filtration rate (GFR) did not reflect the improved proteinuria or the structural improvement with ARB, likely reflecting both the marked decrease in GFR resulting from the removal of a large portion of nephron mass before initiating treatment and the relatively short period of treatment.

Kidney scarring. By study design, glomerulosclerosis was similar in 8-wk biopsies in all groups (sclerosis index at 8 wk untreated 5/6 Nx 0.92 ± 0.16, ARB 0.86 ± 0.17, AT2RA 0.79 ± 0.10, combined 0.78 ± 0.10). At 12 wk after 5/6 Nx, glomerulosclerosis, tubular atrophy, dilatation, cast formation, and interstitial fibrosis were far more advanced compared with biopsy in untreated 5/6 Nx and also in rats treated with ARB (Fig. 1A). Normal baseline kidneys removed at time of 5/6 Nx showed no glomerulosclerosis, interstitial fibrosis, or tubular atrophy. Average sclerosis increase was significantly less in ARB-treated rats, with regression in some rats (percent change in sclerosis from biopsy to autopsy; untreated 5/6 Nx 93.4 ± 17.8, ARB 50.6 ± 18.3, AT2RA 121.8 ± 25.8, combined 57.3 ± 26.6%; ARB vs. AT2RA P < 0.05; Fig. 1B).

Kidney cortex collagen content, assessed by HPLC (37), was dramatically increased compared with normal kidney tissue removed at time of 5/6 Nx in untreated 5/6 Nx, AT2RA, and combined groups (Fig. 2), and nearly normalized in rats treated with ARB (normal kidney 1.7 ± 0.3, untreated 5/6 Nx 18.7 ± 3.8, ARB 2.5 ± 0.7, AT2RA 14.9 ± 6.6, combined 16.3 ± 2.0%, normal vs. untreated 5/6 Nx and combined P < 0.01, normal vs. AT2RA P < 0.05, ARB vs. AT2RA P < 0.05, ARB vs. combined and untreated 5/6 Nx P < 0.01).

We next studied whether AT1a vs. AT1b receptor expression shifted with interventions. Compared with untreated 5/6 Nx at 12 wk, expression of AT1a mRNA in kidney cortex increased 9.6 ± 5.1-fold in ARB-treated rats, 5.0 ± 2.7-fold in AT2RA-treated rats, and 6.8 ± 2.3-fold in combined group (P = 0.06 ARB vs. untreated 5/6 Nx). AT1b mRNA expression in kidney cortex also increased 3.7 ± 0.5-, 2.5 ± 0.4-, and 17.2 ± 11.0-fold, respectively, in ARB, AT2RA, and combined group, compared with untreated 5/6 Nx.

Cellular proliferation and apoptosis. Immunostaining for PCNA was negative in the normal rat glomerulus in kidneys untreated 5/6 Nx at time of 5/6 Nx in untreated 5/6 Nx, AT2RA, and combined groups (Fig. 2), and nearly normalized in rats treated with ARB (normal kidney 1.7 ± 0.3, untreated 5/6 Nx 18.7 ± 3.8, ARB 2.5 ± 0.7, AT2RA 14.9 ± 6.6, combined 16.3 ± 2.0%, normal vs. untreated 5/6 Nx and combined P < 0.01, normal vs. AT2RA P < 0.05, ARB vs. AT2RA P < 0.05, ARB vs. combined and untreated 5/6 Nx P < 0.01).

Table 1. Systemic and renal parameters

<table>
<thead>
<tr>
<th>Week</th>
<th>0</th>
<th>4</th>
<th>8</th>
<th>12</th>
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<tbody>
<tr>
<td>Body wt, g</td>
<td>CONT 5/6 Nx</td>
<td>252.5 ± 6.2</td>
<td>312.8 ± 10.0</td>
<td>347.8 ± 19.0</td>
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<tr>
<td>ARB</td>
<td>250.7 ± 10.7</td>
<td>301.5 ± 14.8</td>
<td>355.7 ± 20.1</td>
<td>353.8 ± 32.5</td>
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<tr>
<td>AT2RA</td>
<td>234.0 ± 13.9</td>
<td>278.7 ± 8.2</td>
<td>340.8 ± 8.4</td>
<td>364.7 ± 24.7</td>
</tr>
<tr>
<td>Combined</td>
<td>226.2 ± 13.5</td>
<td>283.7 ± 13.6</td>
<td>330.2 ± 3.7</td>
<td>368.3 ± 16.1</td>
</tr>
<tr>
<td>Systolic blood pressure, mmHg</td>
<td>CONT 5/6 Nx</td>
<td>130.0 ± 6.4</td>
<td>186.6 ± 13.9</td>
<td>208.2 ± 8.8</td>
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<tr>
<td>ARB</td>
<td>118.2 ± 4.6</td>
<td>186.5 ± 5.7</td>
<td>200.8 ± 6.7</td>
<td>157.1 ± 12.8*</td>
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<tr>
<td>AT2RA</td>
<td>114.0 ± 3.5</td>
<td>183.6 ± 14.5</td>
<td>186.3 ± 11.5</td>
<td>176.4 ± 10.7</td>
</tr>
<tr>
<td>Combined</td>
<td>120.1 ± 5.6</td>
<td>181.5 ± 7.8</td>
<td>192.7 ± 10.0</td>
<td>163.5 ± 4.0*</td>
</tr>
<tr>
<td>Proteinuria, mg/day</td>
<td>CONT 5/6 Nx</td>
<td>8.7 ± 1.9</td>
<td>200.8 ± 37.4</td>
<td>280.3 ± 53.0</td>
</tr>
<tr>
<td>ARB</td>
<td>7.0 ± 1.8</td>
<td>206.0 ± 27.1</td>
<td>293.3 ± 32.8</td>
<td>270.1 ± 55.6</td>
</tr>
<tr>
<td>AT2RA</td>
<td>10.3 ± 1.8</td>
<td>142.0 ± 29.1</td>
<td>283.0 ± 58.2</td>
<td>397.2 ± 30.4</td>
</tr>
<tr>
<td>Combined</td>
<td>7.1 ± 0.9</td>
<td>148.4 ± 27.1</td>
<td>256.8 ± 76.5</td>
<td>250.5 ± 65.5*</td>
</tr>
<tr>
<td>Serum creatinine, mg/dL</td>
<td>CONT 5/6 Nx</td>
<td>0.33 ± 0.03</td>
<td>0.93 ± 0.13</td>
<td>0.90 ± 0.07</td>
</tr>
<tr>
<td>ARB</td>
<td>0.30 ± 0.00</td>
<td>0.85 ± 0.03</td>
<td>1.00 ± 0.11</td>
<td>2.28 ± 1.41</td>
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<tr>
<td>AT2RA</td>
<td>0.38 ± 0.02</td>
<td>1.14 ± 0.43</td>
<td>1.06 ± 0.07</td>
<td>1.82 ± 0.62</td>
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<tr>
<td>Combined</td>
<td>0.32 ± 0.02</td>
<td>0.88 ± 0.04</td>
<td>0.96 ± 0.05</td>
<td>2.00 ± 0.70</td>
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Values are means ± SE. ARB, angiotensin type 1 receptor blocker; CONT 5/6 Nx, control 5/6 nephrectomy. *P < 0.05 vs. CONT.
Glomerular PCNA-positive cells included mesangial area and endothelial cells, and rare cells consistent with podocytes. Tubular cells were frequently positive for PCNA in untreated 5/6 Nx and AT2RA, with rare tubular positivity in ARB and combined groups. PCNA-positive interstitial cells were occasionally present in untreated 5/6 Nx and AT2RA groups, and were very rare in ARB and combined groups. Apoptosis as assessed by TUNEL-positive cells was present mostly in tubular epithelial cells, with few positive cells in glomeruli with no difference among groups (glomerulus: untreated 5/6 Nx 0.4 ± 0.2, ARB 0.3 ± 0.1, AT2RA 0.5 ± 0.1, combined 0.3 ± 0.2/glomerulus; cortex: untreated 5/6 Nx 2.1 ± 0.2, ARB 2.0 ± 0.1, AT2RA 2.2 ± 0.2, combined 2.2 ± 0.2). Thus, the balance of proliferation vs. apoptosis was shifted in untreated and AT2RA-treated glomeruli.

**Erk 1/2 expression.** We next evaluated the activation of Erk1/2, a downstream signaling molecule for the angiotensin system. In normal kidney cortex, phosphorylated Erk was present at low levels. Phospho-Erk2 expression was upregulated by AT2RA vs. ARB or combined (normal kidney 0.39 ± 0.01, untreated 5/6 Nx 0.78 ± 0.10, ARB 0.56 ± 0.04, AT2RA 0.86 ± 0.07, combined 0.61 ± 0.10; normal vs. untreated 5/6 Nx and AT2RA P < 0.01, ARB vs. AT2RA P < 0.05, AT2RA vs. combined P < 0.05; Fig. 3).

**Fig. 3.** Representative Western blots of phospho-Erk 2 protein in the renal cortex of rats at 12 wk after 5/6 Nx (A). The expression of phospho-Erk 2 protein is presented as phospho-Erk 2/total Erk 2 ratio (B). ***P < 0.01, normal vs. control untreated 5/6 Nx, normal vs. AT2RA, AT2RA vs. combined. *P < 0.05, normal vs. AT2RA, ARB vs. AT2RA; n = 4 for normal; n = 5–6 for all other groups.
PAI-1 and plasmin expression. In normal kidney, PAI-1 mRNA by in situ hybridization showed diffuse low-level tubular and no glomerular expression (data not shown). PAI-1 mRNA expression was augmented in untreated 5/6 Nx and AT2RA at 12 wk and was localized mainly in sclerotic glomeruli, mesangial areas, tubules, interstitial cells, and occasionally in glomerular endothelial cells, podocytes as identified by anatomical location, and small artery endothelial and vascular smooth muscle cells. Image analysis showed that PAI-1 mRNA was significantly increased in untreated 5/6 Nx, and even further increased in AT2RA, compared with significant decrease in ARB and combined group (untreated 5/6 Nx 9.79 ± 1.00, ARB 4.36 ± 0.54, AT2RA 17.24 ± 1.62, combined 7.00 ± 1.69%; untreated 5/6 Nx vs. ARB P < 0.05, AT2RA vs. untreated 5/6 Nx, ARB or combined P < 0.01; Fig. 4).

PAI-1 protein expression paralleled PAI-1 mRNA. PAI-1 immunostaining was mainly increased in areas of sclerosis and fibrosis with less staining in ARB and combined vs. untreated 5/6 Nx or AT2RA (Fig. 5). Western blotting verified that cortical PAI-1 protein expression was increased in untreated 5/6 Nx and AT2RA compared with ARB, with intermediate level in combined (untreated 5/6 Nx 0.70 ± 0.03, ARB 0.56 ± 0.04, AT2RA 0.73 ± 0.03, combined 0.64 ± 0.03; untreated 5/6 Nx vs. ARB P < 0.01, ARB vs. AT2RA P < 0.01; Fig. 6).

Plasmin activity of kidney cortex at 12 wk after 5/6 Nx showed a trend for decrease in AT2RA and combined groups even below the slightly decreased level seen in untreated 5/6 Nx, whereas ARB-treated rats had normalized plasmin levels (normal kidney 0.16 ± 0.05, untreated 5/6 Nx 0.12 ± 0.02, ARB 0.16 ± 0.08, AT2RA 0.08 ± 0.02, combined 0.06 ± 0.01) (28, 36).

Effects of AT1 vs. AT2 receptor blocker on podocytes. The podocyte is a key responder and modulator of development of glomerulosclerosis. We therefore examined podocyte-specific responses to ANG II, evaluating whether this response was influenced by AT1 or AT2 receptor blocker. We further assessed whether PAI-1 contributed to any differential response by studying WT and PAI-1−/− podocytes. We used mouse primary podocyte cultures as rat knockout for PAI-1 is not available. In WT, but not PAI-1−/− podocytes, ANG II significantly increased collagen mRNA in glomeruli and the tubulointerstitial area. PAI-1 immunostaining was especially increased in sclerotic glomeruli and was markedly increased in control untreated 5/6 Nx and AT2RA-treated rats (A, B, E, F), with less staining in ARB and combined rats (C, D, G, H). A, C, E, G: ×400. B, D, F, H: ×200, anti-PAI-1 staining.

Fig. 5. PAI-1 immunostaining at 12 wk after 5/6 Nx (control 5/6 Nx: A, B. ARB: C, D. AT2RA: E, F; combined: G, H). PAI-1 protein paralleled that of PAI-1 mRNA in glomeruli and the tubulointerstitial area. PAI-1 immunostaining was especially increased in sclerotic glomeruli and was markedly increased in control untreated 5/6 Nx and AT2RA-treated rats (A, B, E, F), with less staining in ARB and combined rats (C, D, G, H). A, C, E, G: ×400. B, D, F, H: ×200, anti-PAI-1 staining.

Fig. 4. Expression of plasminogen activator inhibitor-1 (PAI-1) mRNA in situ hybridization in kidney cortex was quantitatively analyzed by SCION Image Software and expressed as % area occupied by PAI-1 mRNA signal. PAI-1 mRNA expression was higher in control untreated 5/6 Nx, and further increased in AT2RA, compared with significant decrease in ARB and combined group. **P < 0.01, control untreated 5/6 Nx vs. AT2RA, ARB vs. AT2RA, AT2RA vs. combined. *P < 0.05, control untreated 5/6 Nx vs. ARB.
346.7 ± 186.3 ng/ml, ANG II vs. ARB *P < 0.05, ANG II vs. AT2RA or combination *P = NS; Fig. 7). Although phosphorylation of Erk was increased by ANG II, ARB and AT2RA were equally effective in blunting this response in WT podocytes (p-Erk/β-actin ratio: control 0.70 ± 0.13, ANG II 1.60 ± 0.09, ANG II + ARB 0.89 ± 0.01, AT2RA 0.97 ± 0.18, combination 0.94 ± 0.09; Fig. 8). These studies support that the podocyte’s profibrotic response to ANG II is largely AT1 mediated and is dependent on the presence of PAI-1.

**DISCUSSION**

The pathophysiological role of the AT2 receptor in CKD has not been defined. Renoprotection linked to the AT2 receptor has been variably reported even in the same model, 5/6 Nx (9, 25, 50). These results could, in part, reflect different time points of...
intervention and observation. Thus, the AT$_2$ receptor was linked to injurious effects at 4 wk after 5/6 Nrx (9). In contrast, later or earlier time points suggested beneficial effects of the AT$_2$ receptor on sclerosis (19). Species differences in mice vs. rats could also underlie some of the conflicting reports. We chose to study the rat model of 5/6 Nrx with ligation of renal artery branches, as results from this model previously showed relevance to insights and treatment of human CKD. Effects of AT$_2$R may differ in other models with less hypertension and other pathogenic mechanisms. Next, we decided to study the role of the AT$_2$ receptor in existing glomerulosclerosis, as this delayed timing of intervention is most relevant to human CKD where treatment is initiated after the disease is clinically evident. Our studies show that in the setting of such delayed intervention, not only is blockade of the AT$_2$ receptor of no evident. Our studies show that in the setting of such delayed timing of intervention is most relevant to human CKD.

The rodent has two subtypes of the AT$_1$ receptor, the major AT$_1$A receptor, and the less abundant AT$_1$B receptor. The latter has recently been suggested to contribute to increased inflammation and glomerular injury (13). ARB blocks both receptors. Our mRNA expression data suggest the possibility that AT$_2$RA could not only block the potential beneficial effects of AT$_2$ receptor, but also result in upregulated AT$_1$B receptor, which might also contribute to worsened injury with AT$_2$RA alone. Whether receptor protein and signal transduction of AT$_1$B are affected by AT$_2$ receptor modulation and possible interactions awaits further study.

ANG II has manifold profibrotic effects in addition to induction of hypertension: it induces extracellular matrix (ECM) accumulation directly and indirectly by inducing TGF-$eta$, and inhibitors of ECM turnover, such as PAI-1, induced by AT$_2$ effects (15, 21, 41). Effects of AT$_2$ on the ECM are less well-delineated. Our results show that PAI-1 mRNA expression was increased by AT$_2$ receptor antagonism even beyond untreated 5/6 Nrx levels, in contrast to the marked decrease in PAI-1 affected by ARB. Furthermore, addition of AT$_2$RA to ARB resulted in higher PAI-1 expression by in situ hybridization compared with ARB alone. Western blots confirmed high levels of PAI-1 in untreated 5/6 Nrx and AT$_2$RA-treated rats, and decreased PAI-1 with ARB, with numerically intermediate level with combined therapy. These results support that the AT$_2$ receptor might suppress progression of glomerulosclerosis and renal scarring through downregulation of PAI-1 in the remnant kidney model, and conversely, that inhibition of AT$_2$ action increases PAI-1, thus promoting fibrosis.

The Erk pathway could induce PAI-1 through transactivation of downstream transcriptional factors such as hypoxia-inducible factor-1, Elk-1, and AP-1 (12, 24, 48). We speculate that AT$_2$ receptor antagonism augments PAI-1 expression through upregulation of Erk phosphorylation in the 5/6 Nrx kidney. Our in vitro studies of podocytes showed that AT$_1$ vs. AT$_2$ effects on ANG II-induced collagen were not linked to altered Erk activation. Possible newly identified mediators of AT$_2$ signaling, such as ATIP1, might be involved in fibrosis responses (22). However, PAI-1 expression might also be modulated by AT$_2$-dependent mechanisms through, e.g., inhibition of bradykinin/NO/cGMP pathway (6). Future in vitro studies may elucidate in greater detail the relationship between the AT$_2$ receptor and PAI-1.

We also showed that renal collagen was significantly decreased by ARB, with even greater benefit of ARB on cortical collagen content, a measure largely reflecting tubulointerstitial fibrosis, than on glomerulosclerosis. Furthermore, added AT$_2$ receptor antagonism in particular dampened ARB benefits on collagen content, supporting key effects of AT$_1$/AT$_2$ balance on the tubulointerstitium in fibrotic states. Effects of the AT$_2$ receptor on fibrosis have been controversial both in vitro and in vivo (2, 9, 29, 30, 33, 35, 39, 40, 45). AT$_2$ receptor stimulation in vitro increased collagen synthesis in vascular smooth muscle and mesangial cells via a $G_{s1}$-mediated mech-
anism, while essentially opposite effects were observed in fibroblasts (39). In vivo studies of transgenic mice with up-regulation of ATIP1 showed protection against neointimal response in the injured femoral artery (22). These results suggest that AT2 receptor-mediated signals might have heterogeneous effects on different cell types.

In summary, our results indicate that the beneficial effects of ARB on renal injury are complex and involve both blockade of deleterious AT1 actions and actions of the AT2 receptor.

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DISCLOSURES

No conflicts of interest are declared by the authors.

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