p66Shc mediates high-glucose and angiotensin II-induced oxidative stress renal tubular injury via mitochondrial-dependent apoptotic pathway

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Submitted 16 July 2010; accepted in final form 23 August 2010


Diabetic nephropathy (DN) is one of the common complications of both type 1 and type 2 diabetes mellitus. DN is characterized by glomerular and tubular hypertrophy initially and followed by thickening of both glomerular and tubular basement membranes (38, 39). Several studies have shown that damage to tubular cells plays a significant role in the progression of DN (38, 39), and the changes in the tubulo-interstitial compartment are well reflected in the deterioration of renal functional parameters, i.e., glomerular filtration rate (26, 27). The mechanisms relevant to its progression include increased activity of polyl and hexosamine pathways, formation of advanced glycation end products, activation of protein kinase C (PKC), generation of reactive oxygen species (ROS) and activation of intrarenal renin-angiotensin system (4, 17, 33, 37). Among the spectrum of biochemical changes induced by high-glucose (HG) ambience, the generation of ROS seems to be one of the main pathophysiological mechanism (7), as a consequence of which the downstream events lead to extracellular matrix accumulation, as well as apoptosis, with the latter being predominantly confined to tubular epithelia. The major source of ROS is believed to be the mitochondrial electron transport system (13). In addition, many factors, such as ischemic renal injury (6), advanced glycation end products (5), angiotensin II (ANG II) (25, 36), besides HG (14), may also induce excessive ROS production and mitochondrial dysfunction, which eventually leads to apoptosis and necrosis of the renal tubular cells (35). Our laboratory’s previous studies have shown that HG can reduce the mitochondrial membrane potential (ΔΨm) and enhance the release of mitochondrial cytochrome c (mCyt.C) into the cytosol with activation of caspases and ultimately apoptosis in tubular cells (32). However, the finer detailed mechanisms modulating HG or ANG II-induced mitochondrial ROS generation and subsequent downstream events in tubular cells need to be teased out further.

Adaptor protein p66Shc is a newly recognized mediator of mitochondrial dysfunction, and it is expressed in most of the mammalian tissues. It has been shown that p66Shc during the generation of hydrogen peroxide (H2O2) utilizes the mitochondrial electron transfer chain via the oxidation of Cyt.C (9, 21). During this process, the p66Shc is imported into the mitochondrial intermembrane space, which requires Ser36 phosphorylation (p-p66Shc) by PKC-β and Pin1 (29). The concept that p-p66Shc leads to the generation of ROS is supported by studies on p66Shc−/− cells, which exhibit reduced levels of intracellular ROS, decreased mitochondrial DNA (mtDNA) alterations, and resistance to apoptosis induced by a variety of stimuli, including H2O2, ultraviolet radiation, human immunodeficiency virus-1, and hypoxia/reoxygenation (12, 15, 24, 29). Similarly, p66Shc−/− mice exhibit increased resistance to oxidative stress and have a prolonged life span (24). They are also protected from the development of diabetic glomerulopathy, presumably due to a blockade of hyperglycemia-induced ROS production (23). Furthermore, it has been reported that the p66Shc mediates mitochondrial dysfunction in renal proximal tubule cells in the ischemia-reperfusion injury model (1), and ANG II induced apoptosis in myocardial cells (10). These literature reports certainly establish the role of p66Shc in states of oxidant stress. However, the role of p66Shc and the detailed mechanisms in the process of mitochondrial ROS generation...
and, subsequently, cell death induced by HG and ANG II in the pathobiology of the tubules remain to be investigated.

The aim of the present study was to determine the status of p66Shc expression and its phosphorylation status in diabetic kidneys, and to assess whether p66Shc contributes to mitochondrial ROS generation, their dysfunctions in renal proximal tubular cells subjected to HG ambience and ANG II exposure, and if PKC-β and Pin1 are involved in this signaling cascade.

MATERIALS AND METHODS

Animal models. Animal studies were conducted under a protocol approved by the Institutional Animal Care and Use Committee at Central South University of China. The animals were housed in a temperature-controlled room, and they were given free access to water and standard laboratory chow. A diabetic state was induced in 8-wk-old ICR mice (N = 20) by an intraperitoneal injection of freshly prepared streptozotocin (STZ; Sigma Chemical, St. Louis, MO). The STZ was prepared by dissolving in 10 mM citrate buffer, pH 4.2. The mice received five consecutive daily injections of STZ (50 mg/kg body wt), while saline buffer served as a control. Blood glucose levels were monitored by One-Touch Glucometer (Lifescan, Milpitas, CA) following vendor’s instructions. All mice were killed 18 wk after the start of STZ treatment. C57BL/KsJ-db/db (N = 20) and their age-matched nondiabetic db/db mice were also used in the study, they were killed at 18 wk of age, and their kidneys were utilized for various studies.

In vivo studies with kidney tissues in diabetic mice. First, apoptosis was assessed by terminal deoxynucleotidyl transferase dUTP-mediated nick-end labeling (TUNEL) method, using In Situ Cell Death Detection Kit, following vendor’s instructions (5). Renal tissue sections for immunostaining were deparaffinized and incubated with 3% H2O2 solution. The sections were then incubated with antibodies directed against p66Shc and phospho-p66Shc (Ser36) (Calbiochem), respectively. The tissue antigen retrieval was performed as previously described (18, 32). The sections were coverslipped mounted and examined with a Zeiss fluorescence microscope. Immunoreactive bands in cell or tissue lysates was adjusted to 1 mg/ml. This was followed by 10% FBS, 100 U/ml of penicillin, 100 μg/ml of streptomycin, and incubated at 37°C in a 5% CO2 environment. Cells were transfected with p66Shc wild type (WT) or the p66Shc536A mutant (Addgene, Cambridge, MA) using LIPOFECTAMINE 2000 reagent (Invitrogen, Carlsbad, CA), as previously discussed (32). After selection of stable transfectants, cells were maintained in a defined medium. After achieving 80% cell confluency, the medium was changed to FBS-free DMEM. Varying concentrations of D-glucose (5–45 mM) and ANG II (10-5–10-2 M) were added. HK-2 cells stably transfected with p66Shc or its mutant p66Shc536A were subjected to HG ambience. The mRNA and protein expression were determined by real-time PCR and Western blotting procedures, respectively.

Gene expression and mtDNA studies. Total RNAs of HK-2 cells were isolated by TRIzol method. First-strand cDNAs were generated by two-step RT-PCR (Fermentas Life Science). Real-time PCR was performed in an Applied Biosystems 7300 Real-time RCR System using a SYBRgreen PCR reagent kit (Invitrogen). The change in mRNA levels was determined by the formula 2-ΔΔCT, where ΔΔCT is the value from the threshold cycle (CT) of the treated sample subtracted from the CT value of untreated or zero time point control sample. The relative amount of mRNA in the sample was normalized to β-actin mRNA. The PCR primers were as follows: p66Shc-sense, 5’-CCACTCCGG AATGATGTTCT-3’, and antisense, 5’-GAAGGACACAGGTT TAGG-3’; β-actin-sense, 5’-ACTCTTCCA GCCTTCTTCC-3’, and antisense, 5’-GGAGGACAAATGCTT GAATCCT-3’. The mtDNA damage of high- and low-molecular-weight DNAs was evaluated by PCR using a REPLI-g mitochondrial DNA isolation kit (QIAGEN), following vendor’s instructions. The PCR was used to simultaneously coamplify both the high- and low-molecular-weight mtDNA. For the high-molecular-weight DNA, the PCR primers were as follows: sense, 5’-TACTAGTCCGGACGAC TTCAAAGC-3’, and antisense, 5’-GGGTGATCTTTGT TCAGGGGT-3’; for low-molecular-weight DNA the primers were as follows: sense, 5’-GCACACGT AAGACCCAAACTGGG-3’, and antisense, 5’-CCCATTTCTCCCATTCATGGC-3’. PCR products of 8,636 and 316 bp were visualized following agarose gel electrophoresis, and a ratio between the high- and low-molecular-weight DNA products was calculated.

Protein expression. The protein expression was evaluated by Western blotting procedures. The total protein of HK-2 cell was isolated with the use of RIPA lysis buffer containing a cocktail of protease inhibitors. Mitochondria and cytosolic extracts were prepared from HK-2 cells using a mitochondrial isolation kit (Pierce, Rockford, IL) and by following the vendor’s instructions. Expression of cytochrome c (Cyt.C) in isolated mitochondria, and total p66Shc (p66Shc), p-p66Shc, and caspase 9 in cytosolic extracts was assessed, using anti-Cyt.C, anti-p66Shc, anti-p-p66Shc (Ser36 site), and anti-caspase 9 antibodies (Santa Cruz, CA), respectively. The anti-mitochondrial heat shock protein 70 (mHSP70) antibody (Novus Biologicals, Littleton, CO) was used where mitochondrial fractions were needed as loading controls.

Detection of cellular apoptosis and Δψm. Apoptosis in HK-2 cells was assessed by using in situ cell death detection kit (Roche Applied Science) following vendor’s instructions. The Δψm was measured by staining the cells with tetramethylrhodamine ethylester perchorlate (TMRE; Molecular Probes). The samples were analyzed using a FACSCalibur BD Flow Cytometry, as previously described (31).

Detection of intracellular H2O2 and assessment of mitochondrial O2−. The H2O2-sensitive fluorescence probe dichloro fluorescein diacetate (DCFH-DA) was used to assess the generation of intracellular H2O2, as previously discussed (9, 31). To detect mitochondrial O2−, MitoSOX Red (Invitrogen) staining was carried out as described (31). The DCFH-DA fluorescence was visualized at an excitation at wavelength of 488 nm and emission at 520 nm. For detection of MitoSOX, 514-nm excitation and 585-nm emission wavelengths were used in a Zeiss LSM510 laser scanning confocal microscope equipped with band-pass filter (Zeiss LSM 510; Carl Zeiss, Thornwood, NY). Intracellular H2O2 and mitochondrial O2− were also detected by FACS analysis after staining with respective dyes (31).

Confocal microscopy. Confocal microscopy was also used to localize the distribution of Cyt.C. HK-2 cells were incubated with 25 nM of Mitotracker red (Molecular Probes, Eugene, OR) at 37°C for 10 min. Slides were washed with PBS and fixed in 4% formaldehyde. They were then incubated with primary anti-Cyt.C antibody (Santa Cruz, CA), followed by another incubation with FITC-labeled secondary antibody. The cells were also subjected to 4,6-diamidino-2-phenylindole staining to visualize nuclei by confocal microscopy, using different excitation filters, followed by merging of various images into a single frame (31).

Lactate dehydrogenase and malondialdehyde measurements in cellular oxidant injury. The levels of lactate dehydrogenase (LDH) and malondialdehyde (MDA) released from damaged HK-2 cells into the culture media were measured using LDH and MDA assay kits both from Promega (Madison, WI).
**Immunoprecipitation and Western blotting studies with p66Shc, p-p66Shc, Cyt.C, and Pin1.** First, the protein concentration in each sample was adjusted to 1 mg/ml, and it was divided into two equal aliquots. Each aliquot containing 100 µg of protein was used for immunoprecipitation (IP) with two different antibodies. Samples were individually incubated with various antibodies [anti-phosphorylated (Ser36) p66Shc antibody, anti-Pin1 antibody, or anti-mCyt.C antibody; Sigma-Aldrich] in an IP buffer for 12 h at 4°C with gentle orbital rotation. Then 50 µl of protein A-Sepharose beads were added, and the incubation was extended for another 12 h. The beads were washed with the IP buffer and resuspended in SDS loading buffer, and then the entire sample was subjected to 10% SDS-PAGE followed by Western blotting procedures (32).

**RESULTS**

p66Shc and p-p66Shc expression, apoptosis, and ROS production in the kidneys of mice with type 1 and type 2 diabetes. p66Shc and p-p66Shc expression in diabetic mice kidneys was evaluated by immunohistochemical procedures (Fig. 1A, a–h). Apoptosis was assessed by TUNEL assay (A, i–l), and ROS production measured using ROS-sensitive vital dye dihydroethidium (A, m–p). In control ICR mouse and db/m mice, expression of p66Shc (Fig. 1A, a and c) and of phosphorylated (p)-p66Shc (Ser36 residue) (Fig. 1A, e and g) was quite low, and it localized mainly in the renal proximal tubules and glomerular mesangium, while the signal was notably increased in the renal proximal tubules of the kidneys of mice with STZ-induced diabetes (Fig. 1A, b and f) and db/db mice at 14 wk of age (Fig. 1A, d and h). In comparison to control, apoptotic cells were increased, and they were mainly confined to the cortical tubules of STZ mice (Fig. 1Aj) and db/db mice at 18 wk (Fig. 1Af). In addition, ROS production was also increased in the renal proximal tubules of STZ mice (Fig. 1An) and db/db mice at 18 wk (Fig. 1Ap). The p66Shc and p-p66Shc protein expression was also evaluated by Western blotting.

**Fig. 1.** A: expression of p66Shc and phosphorylated p66Shc (at Ser36 residue) (p-p66Shc; a–h), status of apoptosis (i–l), and oxidant stress (m–p) in kidneys of control CD1 (ICR) mice (column 1: a, e, i, and m) and those with streptozotocin (STZ)-induced diabetes mice (column 2: b, f, j, and n), db/m mice (column 3: c, g, k, and o), and db/db mice (column 4: d, h, l, and p) mice. In control mice (ICR and db/m), p66Shc and p-p66Shc expression mainly localized in the proximal tubules, and it was increased in mice with STZ-induced diabetes and db/db mice. The terminal deoxynucleotidyl transferase dUTP-mediated nick-end labeling (TUNEL) staining to mark the apoptotic nuclei also notably increased in kidney sections of diabetic mice, compared with the controls (i–l). Similarly, dihydroethidine (DHE) staining to assess the ROS generation showed increased staining in the cortical tubular cells of STZ-induced and db/db mice (m–p). Magnifications: ×200. B and C: Western blot (WB) analyses showed that both p66Shc and p-p66Shc expression were increased in the cortical kidney homogenates of diabetic mice. D and E: densitometry of the autoradiographic bands were depicted in B and C, respectively. Each bar graph represents a relative density ratio between p66Shc or p-p66Shc to β-actin. Values are means ± SE; N = 20 in each group. *P < 0.01 vs. control (Con).
procedures. A dramatically increased p66Shc and p-p66Shc expression was observed in diabetic mice kidneys (Fig. 1, B and C). Densitometric analyses revealed two- to threefold increase of p66Shc/p-p66Shc expression in the kidney tissues of STZ mice and db/db mice compared with the control and db/m mice, respectively (Fig. 1, D and E).

p66Shc and p-p66Shc expression increased in HK-2 cells subjected to HG or ANG II treatment. HK-2 cells (an immortalized proximal tubule epithelial cell line derived from normal adult human kidney) were exposed to different concentrations of D-glucose or ANG II, as indicated, and p66Shc mRNA and protein expression were assessed. As shown in Fig. 2, HG significantly increased the levels of p66Shc mRNA in a time- and dose-dependent manner (Fig. 2, A4 and B4). Similarly, protein expression of total p66Shc, as well as phosphorylated p66Shc (at Ser36 residue), was significantly increased in a time- and dose-dependent manner in HK-2 cell treated with HG (Fig. 2, A and B). The expression of p66Shc in cells treated with L-glucose was unchanged (data not included). Treatment of HK-2 cells with different concentrations of ANG II (10^{-9}, 10^{-7}) (Fig. 2, C and D) also caused an increased in the expression of mRNA, as well as of the protein of both the total p66Shc and its phosphorylated form, and these changes paralleled that seen with the exposure of HG.

Effect of p66ShcS36A mutant, p66Shc-short interfering RNA, and PKC-β-short interfering RNA on the p66Shc and p-p66Shc protein expression in HK-2 cells subjected to HG or ANG II. HK-2 cells were transfected with mutant p66ShcS36A, p66Shc-short interfering RNA (siRNA), or PKC-β-siRNA and then exposed to HG or ANG II. The p66Shc and p-p66Shc protein expression was assessed by Western blotting procedures with anti-p66Shc antibody and anti-phosphorylated (Ser36) p66Shc antibody, respectively. Treatment with HG (30 mM), ANG II (10^{-7}), or overexpression of WT p66Shc increased expression of both p66Shc and p-p66Shc compared with that of the control (P < 0.01) (Fig. 3). The increase was

Fig. 2. Expression of p66Shc and p-p66Shc in HK-2 cells following exposure to high glucose (HG; 5–45 mM) and angiotensin II (ANG II; 10^{-9}–10^{-5} M), as assessed by WB analysis (A1–A3, B1–B3, C1–C3, D1–D3) and real-time PCR (A4, B4, C4, D4). Expression of both p66Shc and p-p66Shc increased in a time- and dose-dependent manner in HK-2 cells treated with D-glucose (A and B) and ANG II (C and D). Expression of β-actin was unchanged. The bar graphs (columns 2, 3, and 4) represent the expression of p66Shc and p-p66Shc relative to β-actin. Values are means ± SE; N = 6. *P < 0.05, #P < 0.01 vs. control.
not observed in cells transfected with a dominant-negative Ser36 mutant of p66Shc (p66ShcS36A), vector alone, or p66Shc-siRNA. A minor increase in the expression of total protein was observed in HK-2 cells transfected with p66ShcS36A mutant from the baseline (Fig. 3). The HG or ANG II-induced expression of p66Shc and p-p66Shc in HK-2 cells was abolished following transfection with p66ShcS36A or pretreatment with p66Shc-siRNA or PKC-β-siRNA (Fig. 3B). This suggested that HG and ANG II induced p66Shc expression, as well as its phosphorylation at Ser36 residue, and it is conceivably mediated by PKC-β.

Overexpression of p66Shc S36A and PKC-β siRNA attenuates the generation of intracellular ROS (H₂O₂) and mitochondrial superoxide in HK-2 cells subjected to HG and ANG II treatments or individually transfected with wild-type (WT) p66Shc, p66ShcS36A (mutant), p66Shc-siRNA, or PKC-β-siRNA. The HG and ANG II treatment or overexpression of p66Shc significantly increased the expression of both p66Shc and p-p66Shc. The overexpression of p66ShcS36A mutant increased the expression of p66Shc, but not of its phosphorylated form, p-p66Shc. Transfection of empty vector or p66Shc-siRNA or PKC-β-siRNA had no significant effect on both p66Shc and p-p66Shc expression. B1–B3: effect of p66Shc, p66ShcS36A, p66Shc siRNA, or of PKC-β-siRNA on the expression of p66Shc and p-p66Shc on the expression of p66Shc and p-p66Shc in HK-2 cell that were treated with HG or ANG II. The p66Shc and p-p66Shc protein expression was increased in HK-2 cell treated with HG and ANG II, and this effect was abolished following transfection with p66Shc-siRNA or PKC-β-siRNA. The transfection of p66ShcS36A had no effect on the expression of p66Shc compared with HG or ANG II-treated groups, but it reduced the expression of p-p66Shc. Overexpression of p66Shc significantly increased the p66Shc and p-p66Shc protein expression, which was reduced following PKC-β-siRNA transfection. The bar graphs represent the densitometric measurements of the protein expression. Values are means ± SE; N = 6. *P < 0.01 vs. 5 mM d-glucose [low glucose (LG)]. @ P < 0.05 vs. 30 mM d-glucose. #P < 0.01 vs. 30 mM d-glucose (HG). & P < 0.01 vs. ANG II (10⁻⁷ M). $ P < 0.01 vs. p66Shc WT.

ROS, as assessed by cells stained with DCFH-DA (a fluorochrome that detects ROS, H₂O₂) and MitoSOX (a fluorescent indicator of mitochondrial superoxide, O₂⁻), and then examined by confocal microscopy (Fig. 4A). The effect on H₂O₂ production was abolished in cells transfected with p66ShcS36A or PKC-β siRNA. In addition, HG or ANG II also increased the mitochondrial “superoxide” (O₂⁻) levels, which were not significantly reduced by transfection with p66ShcS36A, but certainly by the treatment of PKC-β-siRNA (Fig. 4, A and C). The results were also confirmed by FACS analyses (Fig. 4, B and C). By TUNEL staining, a notable increase of apoptosis in cells treated with HG or ANG II was observed, and it was dramatically reduced following treatment with p66ShcS36A or PKC-β-siRNA (Fig. 4, A, bottom row, and D). As assessed by TMRE staining, a notable increase of apoptosis in cells treated with HG or ANG II was observed, and it was dramatically reduced following treatment with p66ShcS36A or PKC-β-siRNA (Fig. 4, A, bottom row, and D). As assessed by TMRE staining, a notable increase of apoptosis in cells treated with HG or ANG II was observed, and it was dramatically reduced following treatment with p66ShcS36A or PKC-β-siRNA (Fig. 4, A, bottom row, and D).
bation with HG or ANG II, which was almost restored to baseline with the overexpression of p66ShcS36A or PKC-β-siRNA (Fig. 4E). Transfection of HK-2 cells with p66Shc WT plasmid induced H2O2 generation, increased apoptosis, and decreased Δψm, and these changes were blocked with the PKC-β-siRNA treatment.

Overexpression of p66ShcS36A and PKC-β-siRNA normalized HG or ANG II-induced Cyt.C release and caspase-9 activation. Compared with low glucose (5 mM), HG (30 mM) and ANG II (10^{-7} M) induced the translocation of mCyt.C from mitochondria into the cytosolic compartment in HK-2 cells (Fig. 5A, rows 2 and 3 vs. 1), as assessed by confocal laser scanning microscopy. Following transfection with p66ShcS36A or PKC-β-siRNA, an inhibition of translocation was observed (Fig. 5A, rows 4 and 6 vs. 2, and rows 5 and 7 vs. 3). These results were confirmed by Western blot analyses. As shown in Fig. 5B, the HK-2 cells treated with HG or ANG II exhibited dramatic decreased expression of mCyt.C in the mitochondrial fractions. The mHSP70 served as a loading control (Fig. 5B, top panel). The expression of mCyt.C and caspase-9 increased significantly in the cytosolic extracts (Fig. 5B, bottom panel). These effects were abolished with the transfection of p66ShcS36A and or PKC-β-siRNA (Fig. 5B, column 4 vs. 2, column 5 vs. 3, column 6 vs. 2, column 7 vs. 3). Using an antibody that specifically recognizes p-p66Shc at Ser36, the Western blotting procedures indicated that HG and ANG II treatments increased the degree of apoptosis, which was reduced with the transfection of p66ShcS36A or treatment of PKC-β-siRNA (row 3). B and C: the bar graphs represent a summary of flow cytometry analyses of cells stained with DCFH-DA and MitoSOX. D: the bar graph represents the quantification of apoptosis in HK-2 cells by TUNEL assay. E: Δψm as measured by tetramethylrhodamine ethylester perchlorate (TMRE) staining revealed a decrease of Δψm in HK-2 cells following HG and ANG II treatments, which was restored with the overexpression of p66ShcS36A or treatment of PKC-β-siRNA. Values are means ± SE. N = 6. *P < 0.01 vs. 5 mM d-glucose. #P < 0.01 vs. 30 mM d-glucose. &P < 0.01 vs. ANG II (10^{-7} M). $ P < 0.01 vs. p66Shc WT.

Fig. 4. A, rows 1–3: confocal microscopy images of cells subjected to dichlorofluorescein-diacetate (DCFH-DA), MitoSOX, and TUNEL staining, and quantification of apoptosis and mitochondrial membrane potential (Δψm). By DCFH-DA staining, overexpression of p66ShcS36A, or treatment of PKC-β-siRNA attenuated intracellular reactive oxygen species (ROS) generation in HK-2 induced by HG (30 mM) or ANG II (10^{-7} M), while treatment of PKC-β-siRNA reduced ROS production in HK-2 induced by p66Shc serving as positive control (row 1). Similar results were seen upon staining with MitoSOX, however, there was no change observed in cells transfected with p66ShcS36A or p66Shc siRNA (row 2). TUNEL staining showed that HG or ANG II increased the degree of apoptosis, which was reduced with the transfection of p66ShcS36A or treatment of PKC-β-siRNA (row 3). B and C: the bar graphs represent a summary of flow cytometry analyses of cells stained with DCFH-DA and MitoSOX. D: the bar graph represents the quantification of apoptosis in HK-2 cells by TUNEL assay. E: Δψm as measured by tetramethylrhodamine ethylester perchlorate (TMRE) staining revealed a decrease of Δψm in HK-2 cells following HG and ANG II treatments, which was restored with the overexpression of p66ShcS36A or treatment of PKC-β-siRNA. Values are means ± SE. N = 6. *P < 0.01 vs. 5 mM d-glucose. #P < 0.01 vs. 30 mM d-glucose. &P < 0.01 vs. ANG II (10^{-7} M). $ P < 0.01 vs. p66Shc WT.
fected with p66Shc WT plasmid compared with HG treatment or p66Shc transfection alone (lane 10 vs. 6 and 2). However, PKC-β-siRNA partially reversed this effect (lane 11 vs. 10). The MDA and LDH levels increased in HK-2 cells exposed to HG, ANG II, or p66Shc compared with the controls. Conversely, their levels were reduced in cells transfected with p66Shc-siRNA (Fig. 6, C and D). The PKC-β-siRNA notably decreased LDH and MDA levels induced by HG, ANG II or by overexpression of p66Shc-S36A mutant or treatment of PKC-β-siRNA reversed these changes. Transfection of p66Shc WT had similar effects, as induced by HG or ANG II treatments on the expression of Cyt.C, p-p66Shc, and caspase-9, and they were inhibited by PKC-β-siRNA. DAPI, 4,6-diamidino-2-phenylindole. Values are means ± SE; N = 6. *P < 0.01 vs. control of 5 mM D-glucose. #P < 0.01 vs. 30 mM D-glucose. &P < 0.01 vs. ANG II (10⁻⁷ M). $P < 0.01 vs. p66Shc WT.

PKC-β and Pin1 pathway modulates the mitochondrial events related to altered expression of p66Shc induced by HG or ANG II. As shown in Fig. 7, A and B, compared with control, HG or ANG II significantly increased p66Shc-S(36) phosphorylation in cytosolic fraction, and it was blocked in a dose-dependent manner in HK-2 cells treated with PKC-β inhibitor, LY333531 (10–100 nM). The expression of β-actin was unchanged (Fig. 7, A and B, top panels). The mCyt.C expression decreased in mitochondrial extracts of HK-2 treated with HG or ANG II, and PKC-β inhibitor reversed the effect in a dose-dependent manner (Fig. 7, A and B, bottom panels). Expression of mHSP70, serving as a loading control, was unchanged. An increased expression of p-p66Shc in the cytoplasm, while a decreased expression of mCyt.C in the mitochondrial fraction of HK-2 treated with 30 mM D-glucose or 10⁻⁷ M of ANG II, was seen compared with that of low glucose. After treatment with Pin1-siRNA (50–250 nM), the level of p-p66Shc expression in the cytoplasm was increased (Fig. 7, C and D, top panels), and also the Cyt.c expression in the mitochondrial fraction was restored almost to baseline in a dose-dependent manner (Fig. 7, C and D, bottom panels, lanes 3–5 vs. 2). The expression of both β-actin and mHSP70 was unchanged. These data indicate that the PKC-β inhibitor mediates p66Shc phosphorylation and subsequent events related to altered expression of p66Shc in HK-2 cells.
quen Cyt.C release from mitochondria into the cytosol in HK-2 cell treated with HG and ANG II, while the Pin1-siRNA reduces the p66Shc translocation into the mitochondria and decreases the mCyt.C release.

HG and ANG II modulate the protein-protein interactions between phosphorylated form of p66Shc (p-p66Shc) and Pin1 in the cytosol and with Cyt.C in the mitochondrial. The HK-2 cells were treated with HG or ANG II, following which the cytosolic and mitochondrial proteins were isolated and subjected to immunoprecipitation/Western blotting procedures. A notable increase in the binding was seen in HK-2 cells treated with HG or ANG II, which represented an interaction between p-p66Shc and Pin1 in the cytosolic extracts (Fig. 8, A1, A2, B1, and B2, lane 2 vs. 1), while the PKC-β-selective inhibitor LY333531 ameliorated this effect (lanes 3–5 vs. 2) in a dose-dependent manner. An increased association of p-p66Shc with mCyt.C in the mitochondrial fraction of HK-2 cells treated with HG or ANG II was observed compared with control (Fig. 8, C and D, lane 2 vs. 1), whereas this effect was abolished with the treatment of Pin1 siRNA (Fig. 8, C1, C2, and D1, D2, lanes 3–5 vs. 2). Similar results were seen with the treatment of PKC-β inhibitor, LY333531 (Fig. 8, E1, E2, F1, and F2), indicating that HG or ANG II enhances the interactions between p-p66Shc and Pin1 in the cytosol, and between p-p66Shc and Cyt.C in mitochondria, and such associations are conceivably mediated by PKC-β and Pin1.

**DISCUSSION**

A number of pathways and molecules have been incriminated in the progression of DN. These mechanisms have been mainly elucidated in the context of pathobiology of the glomerulus, and conceivably they may be also applicable to the proximal tubular compartment that is enriched with mitochondria having metabolic energy demands. The renal proximal tubular cell damage and apoptosis following the generation of ROS has been reported to be responsible for the early stages of DN (3), and their sustained assault is believed to be critical in the progression of DN (34). The HG and ANG II seemed to be the key factors in the induction of intracellular ROS generation in the mitochondrial compartment, and that eventually leads to oxidative damage and apoptosis in renal cells (8, 11, 16, 28). The p66Shc is a newly recognized mediator of mitochondrial dysfunction, which, by oxidizing Cyt.C in the mitochondria, leads to the generation of H$_2$O$_2$, and these sequence of events have been reported to be responsible for the pathogenesis of acute kidney injury in ischemia-reperfusion model (1). However, very limited literature information is available that documents the status of p66Shc phosphorylation in the diabetic state and the effects of p66Shc in HG and ANG II-induced intracellular ROS generation in the mitochondria of tubular cells in hyperglycemia-induced injury models.

To pursue the present study, first we established that both p-p66Shc and total p66Shc protein are mainly localized in the renal proximal tubules, and their expression increases in mice with STZ-induced diabetes (type 1) and db/db mice (type 2), which apparently induce an increased oxidative damage to the tissues, leading to apoptosis (Fig. 1). Similarly, an increased expression of both total and p-p66Shc protein expression was seen in the HK-2 cells treated with HG or ANG II. The HG has been shown to increase the...
synthesis of ANG II (8) and expression of p53, a transcription factor. Interestingly, the consensus sequences for the binding of this transcription factor have been localized in the promoter region of p66Shc (19). Thus the observed increase of p66Shc mRNA expression in DN may be via the ANG II-induced modulation of p53. Other factors modulating the biology of p66Shc may include PKC-β/H9252, which is known to be activated by hyperglycemia and oxidative stress (20). Conceivably, the PKC-β can phosphorylate p66Shc at Ser36 residue (9), and, consequently, the p66Shc would further increase the generation of ROS (9, 29).

Among the Shc protein family, the p66Shc has an additional NH2-terminal calponin homology (CH) domain, known as...
CH2, which includes serine36 amino acid residue phosphorylation site. This domain is believed to function as an oxido-reductase redox enzyme and has been implicated in mitochondrial H₂O₂ generation and translation of oxidative stress signals into apoptosis (2). The next question addressed in this investigation was whether the p-p66Shc is involved in the intracellular ROS production in HK-2 cells following the treatment of HG or ANG II. This was assessed by the use of DCFH-DA, a dye probe that measures ROS (H₂O₂) in the cytosol, as well as in cellular organelles, such as mitochondria. As elucidated in Fig. 4, HG and ANG II, or overexpression of p66Shc (a positive control) increased intracellular ROS production, and this effect was blocked by overexpression of either p66ShcS36A or PKC-β siRNA. To assess if the superoxide, O₂⁻, is generated in the mitochondria of HK-2 via p66Shc mediation, MitoSOX, a mitochondrial O₂⁻ indicator, was employed. The p66Shc S36A mutant failed to reduce significantly the mitochondrial O₂⁻ generation induced by HG and ANG II, while reduced by PKC-β siRNA. These observations support the results of previous studies that ANG II and

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**Fig. 8.** Protein-protein interactions between p-p66Shc and Pin1 in the cytosol (A and B), and between Cyt.C and p-p66Shc (C–F) in the mitochondria following HG, ANG II, PKC-β inhibitor (LY333531), and Pin1-siRNA treatments. The interactions were assessed by immunoprecipitation (IP)/WB procedures. A and B: the LY333531 inhibits the protein-protein interactions between p-p66Shc and Pin1 in the cytosolic fraction of HK-2 cells treated with HG or ANG II in a dose-dependent manner. A1 and B1 are autoradiograms, while A2 and B2 represent the densitometric analysis of density of the respective bands. C–F: both LY333531 and Pin1-siRNA inhibit the protein-protein interactions between p-p66Shc and mCyt.C in the mitochondrial fraction of HK-2 cells treated with HG or ANG II in a dose-dependent manner. C1, D1, E1, and F1 are autoradiograms, while C2, D2, E2, and F2 represent the densitometric analysis of density of the respective bands. The data included in A2, B2, C2, D2, E2, and F2 are representative of 3 experiments.
HG induce mitochondrial H$_2$O$_2$ production (4, 14). Additional conclusions that can be drawn from our studies include that HG or ANG II leads to the activation of PKC-β, which then induces Ser36-phosphorylation of p66Shc and its translocation into the mitochondria to stimulate H$_2$O$_2$ generation in HK-2 cell, while p66Shc seems to not be responsible for directly mediating mitochondrial O$_2^-$ production, since its dominant-negative mutant (p66ShcS36A) had a marginal effect. However, p66Shc is capable of inducing cellular apoptosis and perturbing Δψm under the influence of HG and ANG II, and these effects are certainly consistent with the fact that there was an increase in the levels of intracellular ROS. Collectively, these data demonstrate that PKC-β/p66Shc pathway activated by HG or ANG II induced H$_2$O$_2$ generation within mitochondria, followed by mitochondrial dysfunction and apoptosis.

With respect to the distal events in the mitochondrial that lead to apoptosis, Lee et al. demonstrated ANG II-induced Cyt.C release from the mitochondria concurrent with caspase-3 activation and DNA fragmentation, as well as apoptosis in primary lung endothelial cells (22). As our laboratory reported earlier, the activation of mitochondrial proapoptotic pathway can also be induced by HG (18, 32); however, the mechanisms by which ANG II modulates mitochondrial cell death in renal proximal tubular cells has not been previously reported. Our findings clearly implicate that HG and ANG II or overexpression of p66Shc increases p66Shc phosphorylation, enhances the release of Cyt.C from the mitochondria into the cytoplasm, activates caspase-9, and damages the mitochondrial DNA, and these effects are reversed by the treatment with p66ShcS36A, p66Shc-siRNA, and PKC-β-siRNA (Fig. 5). Other markers of cell death include LDH and lipid hydroperoxides (MDA), and they have been widely used as indicators of ROS production in various cells, including tubular cells (30). Both HG and ANG II induced a notable increase in the cellular levels of LDH and MDA in HK-2 cells, and they were also associated with increased fragmentation of high-molecular-weight mtDNA (Fig. 6). The low-molecular-weight mtDNA was unaffected. These findings basically confirm our contention that PKC-β/p66Shc are the essential modulators that lead to the HG or ANG II-induced mitochondrial dysfunction and subsequent cellular changes in renal proximal tubular cells.

Peptidyl prolyl cis-trans isomerase 1, Pin1 (21), belongs to the parvulin subfamily of peptidyl prolyl cis-trans isomerases. The PKC-β-mediated phosphorylation of p66Shc triggers its translocation into the mitochondrial compartment, where it gets associated with Pin1, and this association plays a key role in the generation of H$_2$O$_2$. Experiments were carried out if the PKC-β-mediated p66Shc phosphorylation leads to the association of p-p66Shc with Pin1 to induce cellular oxidant stress. The HK-2 cells exposed to HG and ANG II were treated with LY333531, a PKC-β selective inhibitor. As shown in Fig. 7, A and B, HG and ANG II induced p66Shc and Cyt.C expression in the cytosolic and mitochondrial compartments, respectively, while LY333531 treatment inhibited the expression of p-p66Shc. The treatment of HK-2 cells subjected to HG and ANG II with Pin1 siRNA increased p-p66Shc expression in cytosolic and Cyt.C in the mitochondrial fraction, indicating that PKC-β and Pin1 not only modulate p66Shc activation, but also are involved in its translocation into the mitochondria, which is followed by p66Shc-Pin1 protein-protein interactions. This contention is indirectly supported by the mutational analyses experiments, where transfection of p66ShcS36A, a mutant lacking serine phosphorylation site, failed to yield the above-described effects. Besides protein-protein interaction between p-p66Shc and Pin1, interaction between p-p66Shc and Cyt.C can conceivably occur to accomplish the cascade of apoptosis. This was assessed by the Western blotting combined with IP experiments, in which usage of the antibodies was exchanged between these two methods. These interactions are believed to occur via the phosphotyrosine binding domain in the NH$_2$-terminus, having conserved glutamic acid residues (9). As shown in Fig. 8, the HG or ANG II certainly accentuated the interactions between p-p66Shc and Pin1 in the cytosol while between p-p66Shc and Cyt.C in the mitochondrial compartment. These data indicated that HG and ANG II can heighten the interaction of p-p66Shc with Pin1 in cytoplasm, and p-p66Shc binding to Cyt.C in mitochondria via the PKC and Pin1 pathway.

In conclusion, these findings highlight the activation of a novel PKC-β/p66Shc-Pin1-Cyt.C pathway (Fig. 9) by HG and ANG II treatment in proximal tubular epithelial cells, and it is anticipated that this information would yield impetus for future studies to study the biology of the tubular compartment in the pathogenesis of DN.

GRANTS
This work was supported by grants from National Foundation Committee of Natural Science of China (C1404085, 30971379), Fuzhou Scholars Fund from Education Department of Hunan Province, Natural Science Foundation of Hunan Province (09JJ6031), and National Institute of Diabetes and Digestive and Kidney Diseases Grants DK28492 and DK60635, USA.

DISCLOSURES
No conflicts of interest, financial or otherwise, are declared by the author(s).
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