Impaired EphA4 signaling leads to congenital hydronephrosis, renal injury, and hypertension

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Sällström J, Peuckert C, Gao X, Larsson E, Nilsson A, Jensen BL, Onozato ML, Persson AE, Kullander K, Carlström M. Impaired EphA4 signaling leads to congenital hydronephrosis, renal injury, and hypertension. Am J Physiol Renal Physiol 305: F71–F79, 2013. First published May 1, 2013; doi:10.1152/ajprenal.00694.2012.—Experimental hydronephrosis induced by partial ureteral obstruction at 3 wk of age causes hypertension and renal impairment in adult rats and mice. Signaling by Ephrin receptors (Eph) and their ligands (ephrins) importantly regulates embryonic development. Genetically modified mice, where the cytoplasmic domain of the EphA4 receptor has been substituted by enhanced green fluorescent protein (EphA4GF/0), develop spontaneous hydronephrosis and provide a model for further studies of the disorder. The present study aimed to determine if animals with congenital hydronephrosis develop hypertension and renal injuries, similar to that of experimental hydronephrosis. Ultrasound and Doppler techniques were used to visualize renal impairment in the adult mice. Telemetric blood pressure measurements were performed in EphA4GF/0 mice and littermate controls (EphA4+/+ ) during normal (0.7% NaCl)- and high (4% NaCl)-sodium conditions. Renal excretion, renal plasma flow, and glomerular filtration were studied, and histology and morphology of the kidneys and ureters were performed. EphA4GF/0 mice developed variable degrees of hydronephrosis that correlated with their blood pressure level. In contrast to EphA4+/+ , the EphA4GF/0 mice displayed salt-sensitive hypertension, reduced urine concentrating ability, reduced renal plasma flow, and lower glomerular filtration rate. Kidneys from EphA4GF/0 mice showed increased renal injuries, as evidenced by fibrosis, inflammation, and glomerular and tubular changes. In conclusion, congenital hydronephrosis causes hypertension and renal damage, similar to that observed in experimentally induced hydronephrosis. This study further reinforces the supposed causal link between hydronephrosis and later development of hypertension in humans.

ephrin; gene modified mice; human disorder; receptor; ureteral obstruction

CONGENITAL ANOMALIES OF THE kidney and urinary tract include a wide range of renal and urinary tract malformations and are detected in ~0.2–2% of all newborns (35). One of the most prevalent urological anomalies that can lead to renal insufficiency is congenital obstructive nephropathy (12). Histopathologically, the obstruction frequently becomes manifest as hydronephrosis (19, 24). Several underlying mechanisms for obstructive nephropathy have been identified, such as developmental abnormalities of the urinary collecting system as seen in duplicate collecting systems, posterior urethral valves or parenchyme malformations as renal dysplasia, renal agenesis, renal tubular dysgenesis, and polycystic renal diseases (2). The familiar accumulation of congenital kidney and urinary tract anomalies indicates that a genetic component may be involved in its formation. (11, 16, 32).

Although congenital urinary tract obstruction is a common disorder in infants, its pathophysiology remains poorly understood and its clinical management has been debated among urologists for many years. Studies have demonstrated that kidney function, in terms of renal perfusion and glomerular filtration remains, rather well preserved during infancy in hydronephrosis (3, 15, 34). These observations have led to a worldwide trend towards nonoperative management in children with nonsymptomatic unilateral hydronephrosis, but the long-term physiological consequences of this new policy are unclear (4). We have previously demonstrated that experimentally induced hydronephrosis, after completed nephrogenesis, is associated with renal injuries and is causally related to hypertension in both rats (5, 9) and mice (6, 8). However, the potential link between congenital hydronephrosis and later development of hypertension and renal dysfunction has not been investigated.

Eph receptors constitute the largest known family of receptor tyrosine kinases and are involved in the modulation of the cytoskeleton and in regulation of cell adhesion and migration (17). During development, Eph receptors and their ligands (i.e., ephrins subclass A and B) have central functions in patterning the embryo in somitogenesis, in neuronal circuit formation, and in angiogenesis (10, 13, 17, 26–28). Genetically modified mice, where the cytoplasmic domain of the EphA4 receptor has been substituted by enhanced green fluorescent protein (EphA4GF/0), develop spontaneous hydronephrosis due to the importance of intact Ephrin-Eph receptor signaling in embryonic development. Therefore, EphA4GF/0 mice are of interest for studying long-term consequences of congenital hydronephrosis on renal and cardiovascular function.

The aim of the present study was to further investigate the suggested link between congenital hydronephrosis and later development of hypertension and renal damage. Using EphA4GF/0 mice and littermate controls (EphA4+/+ ), we hypothesized that animals with congenital hydronephrosis would develop hypertension, renal dysfunction, and kidney injuries similar to those observed in experimentally induced hydronephrosis.
**MATERIALS AND METHODS**

**EphA4gf/gf** mice, generated as earlier described (17), and their corresponding wild-type littermates (**EphA4+/**+) were bred at the department and genotyped by PCR. The experiments were performed in 3- to 10-mo-old male and female mice from the breeding colony. Like in humans, hydronephrosis is occasionally found also in mice. In the present study, a small number of C57BL/6 mice with spontaneous hydronephrosis were included. These mice were discovered during routine induction of experimental hydronephrosis in 3-wk-old wild-type mice. In these mice, the abdomen was closed and blood pressure measurements were conducted at 3 mo of age. Corresponding sham-operated littermates (C57BL/6), with normal appearance of the kidneys, were used as control animals.

The animals were given a standardized normal-salt diet (0.7% NaCl, SD389-R36; Lactamin) followed by a high-salt diet (4% NaCl, SD312-R36; Lactamin, Kimstad, Sweden).

**Ultrasound investigation of renal anomalies.** Animals were anesthetized with isoflurane (Forene; Abbot Scandinavia, Kista, Sweden), the fur above the position of the kidneys was shaved, and water-based ultrasound gel was applied. The kidneys were examined at 15 MHz with a linear high-frequency ultrasound transducer (Sequoia; Siemens-Acuson, Mountainview, CA). The built in Doppler feature was used to visualize blood flow in the kidney.

**Telemetric measurements.** A telemetric device (PA-C10; DSI, St. Paul, MN) was implanted in adult mice in the same way as earlier described (8). In short, isoflurane gas anesthesia was used and the catheter of the transmitter was placed into the left carotid artery with a linear high-frequency ultrasound transducer (Sequoia; Siemens-Acuson, Mountainview, CA). The built in Doppler feature was used to visualize blood flow in the kidney.

**Renal plasma flow and glomerular filtration.** Renal plasma flow (RPF) was measured as the clearance of para-amino hippuric acid (PAH), and glomerular filtration rate (GFR) was measured byulin clearance. These parameters were measured in conscious animals on a normal-sodium diet as previously described (30). In short, animals were given a bolus injection of [3H]methoxy-inulin (ARC, St. Louis, MO) and of [14C]PAH (PerkinElmer, Waltham, MA) dissolved in 200 μl saline into the tail vein whereupon blood samples were taken from the cut tip of the tail at 1, 3, 7, 10, 15, 35, 55, and 75 min. PAH and inulin clearances were calculated using noncompartamental pharmacokinetic data analysis as earlier described. RPF was estimated from the PAH clearance using a renal extraction ratio of 0.7. The filtration fraction (FF) was calculated from the ratio of RPF and GFR (FF = GFR/RPF).

**Renal excretion measurements.** The mice were placed in metabolism cages for 24 h, with food and water given ad libitum to study renal excretion of fluid and electrolytes. Water consumption and urine production were measured gravimetrically. Sodium and potassium concentrations were determined by flame photometry (FLM3; Radiometer, Copenhagen, Denmark) and urine osmolality by depression of the freezing point (Fiske 210 Micro-Sample Osmometer; Fiske Associates, Norwood, MA).

**Renin measurements.** Mice were anesthetized as described above. Blood samples were taken from the carotid artery and immediately spun down using a cooled centrifuge, whereupon plasma was isolated and frozen in liquid nitrogen. PRC was measured by radioimmunoassay of angiotensin I, using the antibody-trapping technique described previously (22). Renin values were standardized by reference to renin standards (MRC; Holly Hill, London, UK) and expressed in Goldblatt units (GU).

**Determination of hydropnephrotic ratio.** Once the renal and cardiovascular studies had been conducted, animals were anesthetized as described above whereupon the abdomen was opened using a midline incision. A macroscopic examination of both kidneys was performed, and the hydropnephrotic ratios (HNR) were calculated in the same way as earlier described (i.e., HNR = residual urine weight/renal parenchymal weight; Ref. 8). The kidneys were cut latitudinally and the middle part was used for histology.

**Histology and morphology.** Embryos at day 15 (E15.5) and mice at postnatal day 4 (P4) as well as kidneys from adult mice were fixed in formalin (4% in phosphate-buffered saline) for 12 h (embryos and pups) or 48 h (adult tissue) and embedded in paraffin. The tissue blocks were cut into 5-μm sections, stained with hematoxylin and eosin, periodic acid-Schiff, and picro-sirius, and a blinded histopathological evaluation was performed. Based on their HNRs, the kidneys of EphA4gf/gf mice were grouped for the histopathological evaluation as the least or most affected kidney.

The renal cortex, medulla, and the papilla were investigated for fibrosis, inflammation (i.e., infiltration of mononuclear cells), tubular changes (i.e., atrophy, dilatation with hyaline cast deposition) and glomerular changes (i.e., sclerosis, epithelial cell proliferation, Bowman’s space dilatation with periodic acid-Schiff-positive material deposition). The presence of glomerular change (G) was assessed and scored in 100 glomeruli in each section as G0 for a normal glomerulus; G1, mild sclerosis (<25%); G2, moderate segmental sclerosis (25–50%); and G3, severe segmental sclerosis (>50%). Tubular changes (T) were categorized semiquantitatively using a grading system as T0, normal; T1, focal tubular epithelial simplification with periarterial and peritubular fibrosis; and T2, multiple focal areas of tubular epithelial atrophy, dilatation with hyaline casts, and/or mononuclear cell infiltration in a large area. The damage score was calculated as (0 × number of S0 + 1 × number of S1 + 2 × number of S2 + 3 × number of S3)/(number of S0 + S1 + S2 + S3), where S represents glomerulosclerosis or tubulointerstitial injury.

**Investigation of ureter morphology ex vivo.** The bladder and the ureters from adult mice were dissected during immersion with PBS, and a ligature was placed around the ureters, close to the kidneys. Bromphenol blue solution (2 mg/ml in PBS) was injected into the bladder and allowed to diffuse at 4°C, and the morphology of the ureter lumen was studied and photographed.

**Calculations and statistics.** Values are presented as means ± SE. Statistics were calculated using GraphPad Prism 6 (GraphPad Software, La Jolla, CA). Single comparisons between normally distributed parameters were tested for significance with Student’s paired or unpaired t-test. For multiple comparisons, two-way ANOVA (genotype and diet) followed by the Fisher’s post hoc test was used. Scored data for the histological evaluation were analyzed by the Kruskal-Wallis test followed by the Mann-Whitney U-test. Statistical significance was defined as P < 0.05.

**Ethics.** The experiments were approved by the regional animal ethics committee in Uppsala and conducted in accordance with the National Institutes of Health Guide for Care and Use of Laboratory Animals.

**RESULTS**

At the time of the experiments there was no difference in body weight, age, or gender distribution between the EphA4gf/gf and EphA4+/**+** groups. EphA4gf/gf mice displayed variable degrees of pelvic dilatation that was not seen in their littermate controls (Fig. 1). Bilateral hydronephrosis was evident in 17% of all mice displayed pelvic dilatation and reduced renal Doppler blood flow (Fig. 2).

**Telemetric measurements.** The hydropnephrotic Epa4gf/gf mice developed hypertension of different degrees. The blood pressure was significantly higher in the hydropnephrotic animals than in the EphA4+/**+** mice both during normal- and high-sodium conditions (Fig. 3). Furthermore, EphA4gf/gf mice dis-
played salt-sensitive blood pressure, which was not found in the EphA4<sup>+/+</sup> group. The heart rate was not different between genotypes given the same diet. The high-salt diet did not significantly change heart rate in EphA4<sup>gf/gf</sup> (P = 0.13) or in EphA4<sup>+/+</sup> mice (P = 0.09). In EphA4<sup>gf/gf</sup> mice, the degree of hydronephrosis correlated with the blood pressure level and their salt sensitivity (i.e., increase in blood pressure in response to high-salt intake). No such correlation was observed in EphA4<sup>+/+</sup> mice (Fig. 4).

C57BL/6J mice (n = 3) with congenital hydronephrosis also displayed salt-sensitive hypertension (113 ± 1 to 122 ± 3 mmHg; HNR = 1.56 ± 0.49), which was not found in normal C57BL/6J (n = 8) controls (103 ± 3 to 105 ± 1; HNR = 0.04 ± 0.00).

Renal excretion, plasma flow, and glomerular filtration. The hydronephrotic mice had lower RPF and GFR compared with that of the EphA4<sup>+/+</sup> group (Fig. 5). No difference in FF was found between EphA4<sup>gf/gf</sup> (0.28 ± 0.02) and EphA4<sup>+/+</sup> mice.
The kidneys from EphA4gf/gf was the most predominant feature observed in this mice model deposition, and glomerular changes (i.e., sclerosis, mesangial cells, tubular changes (i.e., atrophy, dilatation with hyaline cast, epithelial fibrosis with infiltration of inflammatory mononuclear cells, tubular changes (i.e., atrophy, dilatation with hyaline cast), and glomerular changes (i.e., sclerosis, mesangial matrix increase, and shrunken glomeruli). Glomerulosclerosis was the most predominant feature observed in this mice model with glomerular extracapillary proliferation, and cystic changes were rarely observed in this model. The histopathological changes in the least affected kidney were somewhat reduced compared with those of the most affected kidney. In EphA4gf/gf mice with unilateral hydronephrosis, increased histopathological changes were also found in the contralateral kidney. All EphA4+/+ mice had normal HNR values (<0.05) and displayed normal histoarchitecture with no significant histopathological changes. Representative sections from EphA4gf/gf and EphA4+/+ are shown in Fig. 8.

Ureter morphology. Injection of bromophenol blue into the bladder of the urogenital system dissected out from adult animals revealed morphological changes in ureters from hydronephrotic EphA4gf/gf mice (Fig. 7). The ureters were dilated and had a tortuous appearance compared with the straight shape of the controls, indicating impaired peristalsis.

DISCUSSION

In the present study, a genetic mouse model reveals a link between congenital hydronephrosis and later development of hypertension. Furthermore, the hydronephrotic animals used in this study displayed impaired renal function and kidney anomalies similar to that described in experimentally induced hydronephrosis (4).

Clinically, the incidence of postnatally confirmed urinary tract dilatation is reported to be 1–1.4% (21, 29, 33). The phenotype of the EphA4gf/gf mice is associated with a duplica-

(0.27 ± 0.02) mice. Urine osmolality was reduced and urine production tended to be higher in EphA4gf/gf compared with the EphA4+/+ mice (Fig. 5). HNR was significantly different in kidneys from EphA4gf/gf mice compared with EphA4+/+ (0.38 ± 0.08 vs. 0.05 ± 0.01).

Plasma renin levels. There was no significant difference in PRC between EphA4gf/gf (2.481 ± 393; n = 4) and EphA4+/+ (3.307 ± 710; n = 5) mice. HNR was significantly different in kidneys from EphA4gf/gf mice compared with EphA4+/+ (0.20 ± 0.07 vs. 0.02 ± 0.01).

Histology and morphology. At the embryonic stage, EphA4gf/gf mice displayed duplax formations at different levels, bifid ureters, duplicated collecting system and duplex kidneys, but not hydronephrosis (Fig. 6). Postnatally those malformations were associated with hydronephrosis and hydroureter (observed penetrance 50%, n = 35; Fig. 7). The malformations were predominantly unilateral, but bilateral changes were also observed in 14% of the embryos and newborns.

The results from the renal histopathological evaluation of kidneys from adult mice are summarized in Table 1. Left and/or right kidneys of EphA4gf/gf mice displayed variable degrees of pelvic dilatation, with flattening of the renal medulla. The hydronephrotic kidneys exhibited areas with subepithelial fibrosis with infiltration of inflammatory mononuclear cells, tubular changes (i.e., atrophy, dilatation with hyaline cast deposition), and glomerular changes (i.e., sclerosis, mesangial matrix increase, and shrunken glomeruli). Glomerulosclerosis was the most predominant feature observed in this mice model with glomerular extracapillary proliferation, and cystic changes were rarely observed in this model. The histopathological changes in the least affected kidney were somewhat reduced compared with those of the most affected kidney. In EphA4gf/gf mice with unilateral hydronephrosis, increased histopathological changes were also found in the contralateral kidney. All EphA4+/+ mice had normal HNR values (<0.05) and displayed normal histoarchitecture with no significant histopathological changes. Representative sections from EphA4gf/gf and EphA4+/+ are shown in Fig. 8.

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tion of the collecting system and the ureter. These changes are evident in the embryonic stage; however, hydronephrosis was only found in postnatal mice. In humans, duplications of the renal collecting system are considered to be common congenital anomalies and a combination of clinical reports and autopsies has yielded an incidence of 0.8% (31). In the clinic, duplex systems have been described to cause hydronephrotic changes as a consequence of ureteropelvic junction obstruction (1, 20, 25) and with an ectopic ureterocoele (24). Furthermore, hydronephrosis in EphA4gf/gf is associated with dilated and tortuous ureters, which indicates impaired peristaltis. Consequently, the occurrence of hydronephrosis in EphA4gf/gf mice might be caused by ureteropelvic junction obstruction due to the observed duplex systems but could also be caused by impaired ureter peristaltis.

In humans, the hydronephrosis is usually found during routine prenatal ultrasound. Previously, surgical relief of the obstruction was routinely performed; however, during the last decade, nonoperative management has been applied if the patient displays normal renal function. At present, no systematic prospective study has been conducted to investigate the long-term consequences of the conservative treatment strategy in children with congenital hydronephrosis. However, there are several case reports on adult patients with hypertension that is obviously caused by hydronephrosis, as they became normotensive following nephrectomy or pyeloplasty (4). Recently, a
bromophenol blue. Both kidneys display hydrourter. Right kidney displays a massively dilated collecting system whereas the dilatation is less pronounced in the left kidney. B–D: ureters from adult mice injected with bromophenol blue. Ureters from hydroureneric EphA4gf/gf mice were dilated and displayed a tortuous appearance (D) suggesting a defective peristalsis. In contrast, ureters from wild types (B) and EphA4gf/gf mice without hydronephrosis (C) displayed a straight shape. Scale bar = 1 mm.

Table 1. Histology of the kidneys from EphA4+/+ and EphA4gf/gf

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<tr>
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<th>EphA4+/+</th>
<th>EphA4gf/gf</th>
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<tr>
<td></td>
<td>Left or right</td>
<td>Most affected</td>
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<tr>
<td>KW/mg</td>
<td>127 ± 8</td>
<td>141 ± 11</td>
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<tr>
<td>KW/BW, ×10³</td>
<td>5.2 ± 0.2</td>
<td>4.9 ± 0.2</td>
</tr>
<tr>
<td>HNR</td>
<td>0.03 ± 0.00</td>
<td>0.40 ± 0.06*</td>
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<tr>
<td>Fibrosis</td>
<td>0.4 ± 0.2</td>
<td>1.5 ± 0.2*</td>
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<tr>
<td>Inflammation</td>
<td>0.3 ± 0.3</td>
<td>1.5 ± 0.3*</td>
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<td>Tubular changes</td>
<td>0.0 ± 0.0</td>
<td>0.5 ± 0.2*</td>
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<td>Glomerular changes</td>
<td>0.0 ± 0.0</td>
<td>1.0 ± 0.2*</td>
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<td>n</td>
<td>8</td>
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Values are presented as means ± SE; n = number of mice. BW, body weight; KW, kidney weight; HNR, hydronephrotic ratio. *P < 0.05, compared with EphA4+/+. †P < 0.05, compared with most affected kidney of EphA4gf/gf.
short period of time. This type of obstruction is very rare in clinical practice, even though it is often referred to in clinical discussions. The renal outcome following chronic partial ureteral obstruction, which is a far more common disorder, is more relevant from a clinical point of view. In the present study, EphA4gf/gf mice with congenital hydronephrosis displayed many of the characteristic features associated with long-term ureteral obstruction and could therefore provide useful information in the future.

The hydronephrotic EphA4gf/gf mice displayed reduced renal plasma flow and lower GFR, which most likely is caused by impaired function of the diseased kidney due to altered renal autoregulation and damages to the cortical-medullary region. In earlier studies, hydronephrotic kidneys had increased oxidative stress (6, 18), reduced nitric oxide bioavailability, and a sensitized tubuloglomerular feedback response (5, 7). These changes may protect the hydronephrotic kidney from excessive pressure by reducing glomerular perfusion and filtration but might also play an important role in the development of hypertension by increasing preglomerular resistance and promote fluid and solute retention. In the present study, the GFR values reflect total filtration of both kidneys. Since the nonhydronephrotic kidney in unilateral hydronephrosis, or the least obstructed kidney in bilateral hydronephrosis, increases its function to compensate (23), the actual GFR in the hydronephrotic kidney is probably even lower. In the present study, the histopathological changes found in contralateral kidneys of EphA4gf/gf mice with unilateral hydronephrosis are most likely caused by long-term hyperfiltration. However, we cannot fully rule out the possibility that the EphA4gf/gf genotype has contributed to the changes.

In the present study, ultrasound was used to visualize the hydronephrotic kidneys. The dilated pelvic area and the reduced blood flow in the kidneys of EphA4gf/gf were clearly seen and supports the PAH clearance studies. Consequently, ultrasound appears to be a useful tool for visualization of mouse kidneys.

Fig. 8. Representative picro-sirius-stained sections from EphA4+/+ (A and B) and EphA4gf/gf (D and E) kidneys. Kidneys of EphA4+/+ mice have a normal appearance with only minor fibrotic changes, while the EphA4gf/gf mice shows pelvic dilatation and areas with pronounced interstitial fibrosis. C, F, G, H, and I: periodic acid Schiff-stained slides from EphA4gf/gf mice. C: interstitium showing infiltration of inflammatory cells, predominantly mononuclear cells, disrupting the normal tubular architecture. F: dilated tubule with cast (*). Note the shortened height of tubular epithelial cells. Glomerulus seen in this field is normal. G: Enlarged glomerulus showing mild pericapsular fibrosis at urinary pole (arrow head). H: 3 glomeruli, 1 of them showing collapse of capillary loops. Surrounding tubulointerstitial area is normal. I: micrograph showing 1 normal and 1 sclerotic glomeruli surrounded by normal tubules.
kidneys and can be used as a noninvasive method to characterize renal blood flow as well as different degrees of pelvic dilatation.

In earlier studies on rats with experimentally induced PUUO, plasma renin was elevated during normal-sodium conditions but not during low- or high-sodium diets. Furthermore, the plasma renin concentration was not related to the degree of hypertension (9). No differences in renin have been found in mice with experimentally induced PUUO (8). In the present study, plasma renin concentration was not significantly altered in the EphA4gf/gf mice compared with wild types. Given that an elevated blood pressure normally suppresses renin, the fact that renin was unaffected in the present study, as well as the earlier mouse model despite hypertension, could indicate a deficient regulation. Consequently, renin might be involved to some extent in the hypertensive development but could not be the sole cause. In further studies, intrarenal renin-angiotensin system components could be characterized since they can be substantially different compared with plasma.

In conclusion, there is a causal link between congenital hydronephrosis and later development of salt-sensitive hypertension and renal damage. Since heart rate was unaltered, increased NaCl intake may increase peripheral resistance and contribute to the observed hypertension in this model. Even though clinical prospective studies are warranted, the present findings together with earlier observations in experimentally induced hydronephrosis strengthen the apprehension that a conservative treatment strategy in hydropnephrotic children may lead to hypertension and renal disease later in life.

GRANTS

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

AUTHOR CONTRIBUTIONS


REFERENCES


