ANP-induced signaling cascade and its implications in renal pathophysiology

Franziska Theilig¹ and Qingyu Wu²

¹Institute of Anatomy, Department of Medicine, University of Fribourg, Fribourg, Switzerland; and ²Molecular Cardiology, Lerner Research Institute, Cleveland Clinic, Ohio

Submitted 25 March 2014; accepted in final form 22 January 2015

Theilig F, Wu Q. ANP-induced signaling cascade and its implications in renal pathophysiology. Am J Physiol Renal Physiol 308: F1047–F1055, 2015. First published January 28, 2015; doi:10.1152/ajprenal.00164.2014.—The balance between vasoconstrictor/sodium-retaining and vasodilator/natriuretic systems is essential for maintaining body fluid and electrolyte homeostasis. Natriuretic peptides, such as atrial natriuretic peptide (ANP), belong to the vasodilator/natriuretic system. ANP is produced by the conversion of pro-ANP into ANP, which is achieved by a proteolytical cleavage executed by corin. In the kidney, ANP binds to the natriuretic peptide receptor-A (NPR-A) and enhances its guanylyl cyclase activity, thereby increasing intracellular cyclic guanosine monophosphate production to promote natriuretic and renoprotective responses. In the glomerulus, ANP increases glomerular permeability and filtration rate and antagonizes the deleterious effects of the renin-angiotensin-aldosterone system activation. Along the nephron, natriuretic and diuretic actions of ANP are mediated by inhibiting the basolaterally expressed Na⁺-K⁺-ATPase, reducing apical sodium, potassium, and protein organic cation transporter in the proximal tubule, and decreasing Na⁺-K⁺-2Cl⁻ cotransporter activity and renal concentration efficiency in the thick ascending limb. In the medullary collecting duct, ANP reduces sodium reabsorption by inhibiting the cyclic nucleotide-gated cation channels, the epithelial sodium channel, and the heteromeric channel transient receptor potential-vanilloid 4 and -polycystin 2 and diminishes vasopressin-gated cation transporter potential. ANP-induced signaling cascade and its implications in renal pathophysiology, corin; edema; volume retention; ANP

MAINTAINING BODY FLUID AND electrolyte homeostasis is a fundamental requirement in virtually all animals. The regulation is controlled by a variety of endocrine, autocrine, and neuronal factors, which can be divided into two antagonizing systems: the renin-angiotensin-aldosterone system (RAAS), the sympathetic nervous system, endothelin-1, and antiuretic hormones; and the vasodilator/natriuretic systems, such as the natriuretic peptides and nitric oxide (NO) (94).

The natriuretic peptides were evolved in the earliest vertebrates to serve as osmoregulatory hormones (105, 106). In mammals, the natriuretic peptide system is well preserved. Three similar peptides, i.e., atrial, brain, or B-type and C-type natriuretic peptides (ANP, BNP, and CNP, respectively) have been identified. Among them, ANP and BNP are more closely related, as indicated by high sequence similarities and a shared receptor, natriuretic peptide receptor-A (NPR-A), also called guanylyl cyclase A. ANP is produced mainly in atrial and ventricular myocytes and secreted in response to cardiac wall stretching and various stimuli such as endothelins and α-adrenergic factors (29). BNP is generated mostly in cardiac ventricles and released in response to volume or pressure overload (48). The function of CNP differs from that of ANP and BNP. CNP is widely expressed in the central nervous system, the vasculature, and the bones, where it acts in a paracrine fashion by binding to a separate receptor; natriuretic peptide receptor-B (NPR-B) regulates cell differentiation and organ function (30, 78, 84). All three natriuretic peptides indiscriminately bind to a third receptor, NPR-C, which serves as the clearance receptor for these peptides (37).

Like many peptide hormones, the natriuretic peptides are synthesized in precursors, i.e., pro-forms. Proteolytical cleavage to remove the pro-peptide is an essential step to activate the natriuretic peptides. In cardiomyocytes, where most ANP and BNP are produced, the membrane-bound serine protease corin has been identified as a critical enzyme for activating the natriuretic peptides (118). Corin, however, is not involved in pro-CNP processing. It has been shown that pro-CNP processing is mostly mediated by furin, an intracellular pro-protein convertase (114).

Corin Structure and Function

Corin is a trypsin-like serine protease (116). The protein contains a single-span transmembrane domain at the N terminus, which anchors corin on the cell surface. Among normal
tissues, corin is most abundantly expressed in cardiomyocytes (117). Functional studies have shown that corin is critical for activating the natriuretic peptides and regulating salt-water balance and blood pressure (6, 116). In mice, for example, lack of corin leads to salt-sensitive hypertension (20, 111), a phenotype similar to that in ANP knockout mice (59). To date, corin gene variations and mutations have been reported in patients with hypertension and heart disease, supporting the physiological importance of corin in maintaining normal blood pressure and cardiac function (35, 38, 93, 124). Most recent studies also indicate a local function of corin and ANP in the pregnant uterus to promote spiral artery remodeling and to prevent pregnancy-induced hypertension (26). Interestingly, corin mutations that impair the natriuretic peptide-processing activity have been identified in patients with preeclampsia, suggesting a role for corin in preeclampsia-induced symptoms (26).

**ANP Receptors and Downstream Signaling**

NPR-A/NPR-B and NPR-C are two major types of receptors for the natriuretic peptides. NPR-A and NPR-B are membrane guanylyl cyclase receptors, whereas NPR-C lacks guanylyl cyclase activity (70). The binding of ANP to the receptor NPR-A leads to the conversion of guanosine triphosphate (GTP) to the intracellular second messenger cGMP (55, 90). The active NPR-A is a homodimer. Each NPR-A monomer contains an extracellular ANP-binding domain at its amino terminus and an intracellular guanylyl cyclase domain at its carboxyl terminus (47). Synthesized cGMPs bind to target proteins, including cGMP-dependent protein kinases (PKG) I and II, cyclic nucleotide-gated ion channels (CNG), and the cyclic nucleotide phosphodiesterase (11).

Two different subtypes of NPR-C with molecular masses of 67 and 77 kDa, respectively, have been identified. These receptors may bind to a broad range of ligands, including ANP, BNP, and CNP. 

4-23C-ANP is a cleaved form of ANP, which also binds to NPR-C, but not NPR-A or NPR-B. The 77-kDa NPR-C primarily serves as a clearance receptor for ligand internalization, whereas the 67-kDa NPR-C also participates in adenyl cyclase activity inhibition through inhibitory guanine nucleotide-regulatory protein Gi and activation of phospholipase C to exert antihypertensive effects (70).

Recently, both NPR-A and NPR-C were shown to mediate the effects of ANP by enhancing Ca2+/calmodulin-dependent NO synthase (NOS) activity, another factor of the vasodilator/natriuretic system with similar renal effects as ANP. It appears that these receptors may act through different mechanisms; NPR-A-induced NO production was shown to be cGMP dependent, whereas NPR-C-dependent NO release was partially mediated by Gi protein (41, 42).

**Renal Expression of ANP Signaling Cascade Components and Its Functional Implications**

Within the kidney, components of the ANP signaling cascade are widely expressed and antagonize the vasoconstrictor/sodium-retaining system by affecting the activity and expression of various transporters, channels, and other signaling components (Fig. 1).

**Glomerulus.** Within the glomerulus, corin expression was undetectable (43), suggesting that glomerular ANP production and activation are negligible, if at all present. ANP from the circulation, however, may act on glomerular epithelial and mesangial cells (21, 66). The receptor NPR-A has been localized to the surface of podocytes and mesangial cells (82, 103). Attributed renal actions of ANP include the regulation of theglomerular filtration rate (GFR) and glomerular permeability. ANP-induced increases in GFR and filtration fractions have been reported in numerous studies (for a review, see Ref. 52). This function has been suggested to occur by raising the capillary glomerular pressure through relaxation of the afferent arteriole and contraction of the efferent arteriole (40). ANP was also shown to cause a rapid (within 5 min) increase in glomerular permeability (7). Analysis of the glomerular filtration barrier revealed that ANP directly and in a reversible manner increases the radius and the number of large glomerular pores without affecting the charge selectivity. As a result of the augmented glomerular permeability, ANP may induce microalbuminuria, which is associated with many diseases such as diabetes and congestive heart failure (CHF) (7, 125).
ANP-induced cGMP production and its functional consequences are not fully understood. It may involve reorganization of F-actin filaments, thereby causing cellular relaxation (101). The possible function of podocytes in this process remains to be elucidated. Mesangial cells express NPR-A and respond to ANP, with strong elevations in intracellular cGMP (10). ANP may also relax mesangial cells by increased hyperpolarization, leading to an increase in the ultrafiltration coefficient associated with augmented glomerular hydrostatic pressure, GFR, and filtration fraction (52).

Additionally, the endogenous ANP/NPR-A/cGMP system may have pleiotropic and renoprotective properties (82) by antagonizing the RAAS. Ogawa et al. (82) demonstrated that aldosterone-induced glomerular injury led to more severe proteinuria and fibrotic changes in NPR-A knockout mice than control wild-type mice. The authors proposed that ANP-induced NPR-A activation may inhibit local extracellular signal-regulated kinase and p38 MAPK signaling pathways as well as the formation of reactive oxygen species. Another possible mechanism underlying the aldosterone-antagonizing effects of ANP was reported recently. In transfected human embryonic kidney cells, ANP attenuated the aldosterone-induced nuclear translocation of the mineralocorticoid receptor (MR) via NPR-A/cGMP-dependent PKGI, resulting in a association of NPR-A with the MR and thus preventing MR activation (75).

Furthermore, the importance of the balance between the RAAS and the ANP systems is demonstrated by an ANG-(1–9)-induced ANP secretion from cardiomyocyte in an AT₂ receptor-dependent manner involving phosphoinositide kinase 3, Akt, NO, and cGMP (19). Thus, the balance between the RAAS and the ANP-induced signaling cascade may be regulated through receptor interactions, hormone secretion and intracellular signaling pathways.

**Proximal tubule.** In the proximal tubule, corin was localized to the apical membrane of the endocytic apparatus and in the brush-border membrane (BBM) of rodent kidneys (89) as well as in human kidneys (43). The particular corin expression pattern suggests the cleavage of filtered and locally produced pro-ANP into active ANP. In this nephron segment, local production of ANP mRNA was detected by in situ hybridization (67, 89). Natriuresis elicited by ANP has been attributed to the inhibition of sodium reabsorption at both proximal (122) and distal nephron segments, including the collecting duct. Lithium clearance studies showed that ANP inhibited proximal tubular sodium reabsorption (17, 50), in part by counteracting angiotensin-stimulated sodium reabsorption (51). ANP also inhibits several Na⁺-dependent transport systems, such as the Na⁺/H⁺ exchanger (112) and type IIa Na⁺-Pi cotransporter (8). The Na⁺⁺-K⁺-ATPase plays a pivotal role in sodium reabsorption in all tubular segments, including the proximal tubule. It has been shown that the Na⁺⁺-K⁺-ATPase is an ANP target (4, 15) and that the inhibition of the Na⁺⁺-K⁺-ATPase by ANP is mediated, to a large extent, via renal dopamine 1-like receptors. Among other transporters that may be regulated by ANP include protein organic cation transporters and Cl⁻ transport in IMCD (40).

**Distal tubule.** Within the distal tubule, corin is highly expressed in vesicles near the apical plasma membrane of the medullary thick ascending limb (89), where it is believed to cleave locally produced pro-ANP (67, 89). Cytocchemical localization revealed a plasma membrane expression of NPR-A in the thick ascending limb upon ANP treatment (92). In this nephron segment, ANP was shown to inhibit Cl⁻ transport in a cGMP- and PKG-dependent manner (9, 80). Furthermore, increased intracellular cGMP levels were shown to directly inhibit Na⁺⁺-K⁺-2Cl⁻ cotransporter (NKCC2) activity (85, 86) by decreasing its apical surface expression through a mechanism mediated by cGMP-stimulated phosphodiesterase 2 (PDE2) (5). Presumably, this effect may affect NKCC2 trafficking by a PDE2-mediated reduction in cAMP and subsequently protein kinase A levels. Thus ANP may initiate Ca²⁺ transients in isolated cortical thick ascending limbs. This activity appeared to be mediated by NPR-C (27).

ANP-dependent NPR-C activation is known to activate phospholipase C and release NO. The functional implication in ANP-mediated NO release remains to be determined.

**Collecting duct.** In the collecting duct system, corin was identified within the apical plasma membrane and apical vesicles (89). The signal intensity increased toward the medulla. Like in the nephron segments, local ANP mRNA and protein expression was detected. Moreover, all components of the ANP-induced signaling cascade could be identified within the inner medullary collecting duct (IMCD). Immunohistochemical studies localized NPR-A to the apical membranes of the IMCD (45), and the cGMP-stimulated phosphodiesterase 5 (PDE5) and protein kinase G II (PKGII) in the cytosol (89, 104).

It is well established that the medullary collecting duct is the main site of ANP action in promoting sodium excretion (122). The current data indicate that ANP regulates Na⁺ reabsorption at both the apical and basolateral sides in the IMCD. As in the nephron segments, sodium reabsorption in principal cells of the IMCD consists of passive Na⁺ entry through the apical membrane and active pumping into the peritubular space by the basolateral Na⁺⁺-K⁺-ATPase. The epithelial sodium channel (ENaC) is the main sodium channel for sodium entry and is characterized by low conductance (4–5 pS) with high specificity for Na⁺ over K⁺ (20-fold), which is inhibited by amiloride (18, 102).

Current data on cGMP/PKGII-induced ENaC activation are less conclusive. In single patch-clamp studies with *Xenopus* 2F3 cells, ANP and 8-pCPT-cGMP were shown to decrease the open probability of ENaC (49). Long-term ANP treatment, however, induced a translocation of ENaC into the apical membrane presumably to prevent sustained natriuresis after prolonged ANP exposure (110).

In addition to ENaC, another type of sodium channel exists in principal cells of the collecting duct, i.e., the CNG (24, 71, 72). CNGs have a higher conductance (28 pS), transport Na⁺ and K⁺ with similar affinities and are inhibited by cGMP, amiloride, and diltiazem (24, 71, 72). It has been shown that cGMP inhibits these channels by PKG-dependent channel phosphorylation and allosteric changes (71). It is possible, therefore, that CNGs may serve as another ANP target in promoting sodium excretion (Fig. 2).

At the basolateral site of IMCD, ANP is known to inhibit Na⁺⁺-K⁺-ATPase activity through PKGII-induced phosphorylation and in a cGMP-dependent manner (Fig. 2) (15, 44, 98).
Moreover, ANP was shown to modulate transepithelial water transport in collecting duct cells. In isolated rat and rabbit cortical collecting ducts, short-term ANP treatment reduces vasopressin-stimulated water permeability (33, 79). Furthermore, in vasopressin-pretreated cells ANP markedly decreased the kinetics of cell swelling, which was mimicked by 8-bromo-cGMP and blunted by PKGII inhibition. ANP also reduced vasopressin-induced phosphorylation of aquaporin-2 (AQP2) at position S256 (64). However, in untreated collecting duct cells, long-term ANP treatment induced an AQP2 translocation into the apical membrane followed by increased phosphorylation, which may increase transepithelial water flux (14, 110). Thus, similarly as shown for ENaC, prolonged ANP actions favor volume retention.

Another mechanism of ANP-dependent natriuresis has been proposed recently and may involve the inhibition of flow-activated Ca\(^{2+}\) entry into collecting duct cells (Fig. 2) (39). The transient receptor potential vanilloid 4 (TRPV4) and the transient receptor potential polycystin 2 (TRPP2) were shown to form a heteromeric channel complex in cilia, a crucial structure for flow sensation. It was found that ANP, cGMP, and PKGII inhibited Ca\(^{2+}\) entry through their action on heteromeric TRPV4-P2 channels. PKGII was shown to phosphorylate the channel complex on TRPP2T719A and TRPP2S827A, thereby inhibiting channel complex formation and preventing the flow-induced Ca\(^{2+}\) entry into cortical collecting duct cells. These findings are intriguing. Further studies are needed to elucidate the influence of intracellular Ca\(^{2+}\) levels on sodium absorption.

**Regulation of ANP Cascade Components**

ANP, corin, and NPR-A expression patterns are mainly regulated by varying their transcription rates. The involved transcription factors as well as factors modulating their action have been identified in past years.

GATA transcription factor family members, particularly GATA-4 and -6, and T box factor 5, have been shown to play an important role in ANP gene expression in cardiac myocytes. GATA-4 functionally synergizes with transcription factors GATA-6, MEF-2, dHAND, SRF, Nkx2.5, and YY1 (48). The phosphorylation of GATA-4 by p38 MAPK was shown to increase the binding affinity of GATA-4 to the ANP promoter, thereby promoting ANP gene expression (48). Other factors modulating ANP gene expression include α-adrenergic agonists, endothelin, prostaglandin F\(_2\alpha\), growth factors, vitamin D, retinoids, glucocorticoids, mechanical strain, and hypoxia (48).

In the corin gene promoter, similar conserved binding sites for Tbx5, GATA, Nkx2.5, and Krüppel-like transcription factors have been identified (34). GATA-4 appears to be the major transcription factor in the heart accounting for increased corin and ANP gene expression under hypertrophic conditions (115). Transcription factors controlling renal corin and ANP expression patterns as well as factors modulating promoter activity in the kidney remain to be determined.

NPR-A gene expression is regulated primarily by Sp1, possessing three binding sites in the promoter region, as determined by mutational analysis (48). A number of factors have been reported to regulate NPR-A gene promoter activity, which includes vitamin D, angiotensin II, endothelin, osmotic stimuli, endothelial NOS, p38 MAPK (48), and serum and glucocorticoid-inducible kinase 1 (23). Interestingly, ANP was demonstrated to negatively regulate NPR-A promoter activity and gene expression via a cGMP-dependent mechanism involving the cGMP response element-binding protein (CREBP) (74). CREBP is widely distributed in human tissues with high abundance in the heart and was found at low levels in the kidney.

Phosphorylation is another important mechanism in regulating NPR-A activity (99, 120). Binding of ANP to NPR-A induces a homologous desensitization of NPR-A, which correlates with a complex phosphorylation pattern of the receptor. Under steady-state conditions, NPR-A is highly phosphorylated at multiple sites on its intracellular domain, including Ser473, Ser487, Ser497, Thr500, Thr502, Thr506, Ser510, and Thr513. NPR-A-activated desensitization was accompanied by increased phosphorylation at Ser487, whereas at the other sites dephosphorylation occurred. Performing functional analysis of Ser487 using site-directed mutagenesis revealed that phosphorylation at this site blunts NPR-A activation but also prevents further desensitization (99, 119). It is believed that ANP-
mediated NPR-A desensitization is one of the reasons for the receptor’s resistance in the presence of high circulating ANP levels.

**Mechanisms in ANP Resistance**

In many edematous disease states, ANP-induced signaling components may be improperly regulated and thereby prevent the natriuretic action of ANP. In fact, ANP resistance is a hallmark of diseases such as CHF (22), liver cirrhosis (68, 77), and nephrotic syndrome (87, 107, 108), which are commonly associated with sodium retention. In these diseases, cumulative sodium retention often leads to edema and ascites. As an underlying mechanism, a maladaptive renal response to compensate for edematous disorders has been identified, and reduced ANP response and dysregulated ANP-induced signaling were found (25, 65, 81, 91, 95).

In animal models of and patients with CHF, liver cirrhosis, and proteinuric kidney disease, attenuated or reduced urine flow rate and decreased urinary sodium excretion are common (12, 22, 31, 32, 46, 57, 58, 61, 69, 73, 113). Increased circulating ANP levels often correlated with the severity of the disease (16, 68, 87, 100). Under these pathological conditions, the renal response to pharmacological ANP infusion was markedly attenuated (22, 83, 87, 88, 107). In fact, the lack of a proper renal response to ANP is a pathological feature, but the underlying mechanism remains controversially discussed. It has been suggested that ANP resistance may be due to decreased ANP availability and/or NPR-A desensitization at the renal site.

To date, reduced uterine, cardiac, and renal corin levels and/or activity have been found in animal models of and patients with preeclampsia, heart failure, and nephrotic syndrome (56, 89, 109). These findings suggest that corin deficiency may impair the ANP signaling pathway, contributing to the ANP resistance in these diseases. In addition, there is growing evidence that PDE5 upregulation may also play a role in ANP resistance. PDE5 is known to degrade cGMP. In chronic cardiovascular diseases, liver cirrhosis, or nephrotic syndrome, increased PDE5 levels have been associated with decreased cGMP levels, which may contribute to ANP resistance and sodium retention in patients (1, 3, 87, 107).

The renal origin of volume retention has been located in the collecting duct, which involves increased activation of the Na\(^+\)-K\(^+\)-ATPase (2, 36, 73) and augmented expression and apical membrane targeting of ENaC subunits (60, 61, 123). In liver cirrhosis, this increase was shown to occur early in disease progression and was reduced subsequently, probably as a counterregulatory mechanism (62). It remains unclear whether Na\(^+\)-K\(^+\)-2Cl\(^-\) and the Na\(^+\)-Cl\(^-\) cotransporter contribute to volume retention under these disease conditions (60, 97, 121).

In rats, long-term ANP infusion induced AQP2 and ENaC trafficking into the apical membrane (110). Similarly, ANP- and cGMP-stimulated mpkCCD\(_{14}\) cells led to increased membrane insertion of AQP2 (13). It is possible that in edematous conditions with high circulating ANP levels, the augmented membrane availability of ENaC and AQP2 may promote sodium and water retention and may therefore, at least partially, be accountable for the accompanied formation of edema and/or hypertension.

Corin is the rate-limiting enzyme for ANP activation. Recently, reduced corin levels were found in kidneys of proteinuric rats (89) and patients with chronic kidney disease (43). So far, no data have been reported regarding the renal expression levels of the ANP-induced signaling components in patients with or animal models of CHF, liver cirrhosis, and preeclampsia. Analysis of kidneys from corin knockout mice revealed a paradoxical increase in medullary PKGII, PDE5, and the ENaC \(\beta\)-subunit, whereas ENaC \(\alpha\)- and \(\gamma\)-subunits as well as Na\(^+\)-K\(^+\)-ATPase expression levels remained unchanged (89). Corin knockout mice developed salt-sensitive hypertension, which was caused by impaired sodium excretion and increased water retention (111). This phenotype was corrected by amiloride treatment; treatment with amlodipine, a Ca\(^{2+}\) channel antib}-
tor, or losartan, an AT1 receptor blocker, had no such effect. These data suggest a potential role of ENaC in inducing salt-sensitive hypertension in coron knockout mice. Similarly, kidneys of proteinuric rats had increased levels of PKGII and PDE5 in the medullary collecting duct (Fig. 3). It remains unclear how corin reduction and/or the increase in PKGII and PDE5 lead to ENaC activation. Further studies are needed to elucidate the underlying molecular mechanism.

Summary and Perspectives

Corin and ANP play an important role in regulating body fluid and electrolyte homeostasis. Components of the ANP signaling cascade are expressed in the glomerulus and along the nephron to exert natriuretic, diuretic, and renoprotective effects. The main natriuretic and diuretic ANP actions occur in the medullary collecting duct via its receptor NPR-A. Dysregulation of the ANP-induced signaling cascade may impair sodium and water excretion. In this context, reduced corin activity, as the rate-limiting enzyme for ANP production, and altered expression levels of ANP signaling components have been found in animal models and patients with ANP resistance and sodium and water retention. There are many open questions which need to be addressed in the future. For example, what is the importance of the renal vs. the cardiac corin-ANP system on renal ANP-induced natriuresis and diuresis? What are the molecular mechanisms of long-term ANP treatment leading to augmented ENaC and AQP2 membrane availability? How is the expression level of corin and ANP signaling components in CHF, liver cirrhosis, or preeclampsia? What is the time course of expression levels of corin and ANP-induced signaling components in edematous diseases? How may dysregulated ANP signaling cascade components contribute to the ANP resistance commonly observed in patients with edematous diseases? Further studies to answer these questions will help our understanding of the role of corin and ANP in sodium homeostasis and renal pathophysiolo

GRANTS

Support from the Deutsche Forschungsgemeinschaft (FOR 667 to F. Theilig), Swiss National Foundation (31003A_138517 to F. Theilig) and National Institutes of Health Grants (HL089298 and HD064634 to Q. Wu) are gratefully acknowledged.

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

Author contributions: F.T. provided conception and design of research; F.T. prepared figures; F.T. drafted manuscript; F.T. and Q.W. edited and revised manuscript; F.T. and Q.W. approved final version of manuscript.

REFERENCES


AJP-Renal Physiol • doi:10.1152/ajprenal.00164.2014 • www.ajprenal.org

Downloaded from http://ajprenal.physiology.org/ by 10.220.33.1 on August 27, 2017


46. Ibebuogu UN, Gladysheva IP, Houng AK, Reed GL. Decompensated heart failure is associated with reduced corin levels and decreased cleavage of pro-atrial natriuretic peptide. *Circ Heart Fail* 4: 114–120, 2011.


