Calmodulin and CaMKII modulate ENaC activity by regulating the association of MARCKS and the cytoskeleton with the apical membrane

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Alli AA, Bao HF, Liu BC, Yu L, Aldrugh S, Montgomery DS, Ma HP, Eaton DC. Calmodulin and CaMKII modulate ENaC activity by regulating the association of MARCKS and the cytoskeleton with the apical membrane. Am J Physiol Renal Physiol 309: F456–F463, 2015. First published July 1, 2015; doi:10.1152/ajprenal.00631.2014.—Phosphatidylinositol bisphosphate (PIP2) regulates epithelial sodium channel (ENaC) open probability. In turn, myristoylated alanine-rich C kinase substrate (MARCKS) protein or MARCKS-like protein 1 (MLP-1) at the plasma membrane regulates the delivery of PIP2 to ENaC. MARCKS and MLP-1 are regulated by changes in cytosolic calcium; increasing calcium promotes dissociation of MARCKS from the membrane, but the calcium-regulatory mechanisms are unclear. However, it is known that increased intracellular calcium can activate calmodulin and we show that inhibition of calmodulin with calmidazolium increases ENaC activity presumably by regulating MARCKS and MLP-1. Activated calmodulin can regulate MARCKS and MLP-1 in two ways. Calmodulin can bind to the effector domain of MARCKS or MLP-1, inactivating both proteins by causing their dissociation from the membrane. Mutations in MARCKS that prevent calmodulin association prevent dissociation of MARCKS from the membrane. Calmodulin also activates CaM kinase II (CaMKII). An inhibitor of CaMKII (KN93) increases ENaC activity, MARCKS association with ENaC, and promotes MARCKS movement to a membrane fraction. CaMKII phosphorylates filamin. Filamin is an essential component of the cytoskeleton and promotes association of ENaC, MARCKS, and MLP-1. Disruption of the cytoskeleton with cytochalasin E reduces ENaC activity. CaMKII phosphorylation of filamin disrupts the cytoskeleton and the association of MARCKS, MLP-1, and ENaC, thereby reducing ENaC open probability. Taken together, these findings suggest calmodulin and CaMKII modulate ENaC activity by destabilizing the association between the actin cytoskeleton, ENaC, and MARCKS, or MLP-1 at the apical membrane.

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The amiloride-sensitive epithelial sodium channel (ENaC) plays a vital role in the regulation of total body fluid homeostasis and control of blood pressure; therefore, regulation of ENaC is critical for normal health. There are two modes of ENaC regulation: altering the density of channels in the membrane by changing membrane insertion or retrieval (40, 43) or altering the activity of individual channels by changing their open probability (1, 12, 21, 41). In particular, the open probability of ENaC is regulated by interaction with anionic lipids like phosphatidylinositol bisphosphate (PIP2) (1, 21, 41). The problem with understanding how this mode of regulation could occur is that PIP2 is a rare lipid in the membrane and ENaC is a relatively rare protein. If PIP2-ENaC interaction depended upon random lateral diffusion of the two molecules in the apical membrane, the channel would open only rarely. In fact, ENaC is typically open about half the time. This implies that the local concentration of PIP2 near ENaC is higher than would be expected based on the average density in the membrane generally. We have previously shown this can be accomplished by myristoylated alanine-rich C-kinase substrate (MARCKS) or MARCKS-like protein (MLP), which binds ENaC and electrostatically sequesters and concentrates PIP2 near ENaC (1). The effector domain of MARCKS contains multiple serine residues that are subject to phosphorylation by PKC, is responsible for binding anionic phospholipids, and harbors a calmodulin binding sequence (14, 35). MARCKS and MLP-1 are regulated by changes in cytosolic calcium; increasing calcium promotes dissociation of MARCKS from the membrane, but the mechanisms by which calcium promotes dissociation from the membrane are unclear (23). However, it is known that increased intracellular calcium can activate calmodulin. Activated calmodulin could regulate MARCKS and MLP-1 in two ways. Activated calmodulin can bind to the effector domains of MARCKS or MLP-1, inactivating both proteins, allowing their dissociation from the membrane (14). Mutations in MARCKS that prevent calmodulin association prevent dissociation of MARCKS from the membrane (14). Alternatively, calmodulin (CaM) also activates CaM kinase II (CaMKII). CaMKII is known to play an important role in the brain, but this multifunctional kinase is also expressed in other tissues, including the kidney. Timmins et al. (38) reported the major isoform of CaMKII expressed in murine kidneys was CaMKIIδ. Rothschild et al. (32) showed that only CaMKIIδ1 is expressed in zebrafish kidneys. The β-isoform of CaMKII has been best characterized for its role in actin cytoskeletal assembly. Hoffman and coworkers (16) recently determined isoform specificity of CaMKII, including the γ-isoform of CaMKII in mediating the inhibition of actin polymerization and modulation of the actin cytoskeleton (16). Disruption of the cytoskeleton with cytochalasin E reduces ENaC activity (29). Filamin is among the many different actin binding proteins that have been reported to bind and regulate the cell surface expression of transmembrane proteins. For example, filamin A has been shown to regulate the surface expression of BKCa channels (20), the CFTR (37), Kv4.2 potassium channels (27), calcium-sensing receptors (CaR) (42), the inwardly rectifying potassium channel Kir2.1 (33), and ENaC (39). Furthermore, filamins function as scaffolding proteins that are capable of connecting the actin cytoskeleton to various cytoplasmic signaling proteins. The association of ENaC with the cytoskeletal network is believed to help maintain its expression at the apical membrane and prevent its downregulation by endocytosis.
There have been multiple studies performed to show the cytoskeleton is involved in the regulation of ENaC (8, 17, 19, 29). The present study corroborates our previous reports of ENaC being regulated by MARCKS and the actin cytoskeleton. Here, we show ENaC activity is sensitive to calcium-calmodulin and CaMKII. We focus on the effect of this pathway on MARCKS protein expression and posttranslational changes in the actin cytoskeletal protein filamin A to elucidate the mechanism by which ENaC is regulated.

MATERIALS AND METHODS

Cell culture and treatments. Xenopus 2F3 cells were maintained as previously described (1, 4, 29). In some experiments, cells were cultured in media containing normal calcium (1.05 mM CaCl₂) or high calcium (4 mM CaCl₂) before being lysed in mammalian protein extraction reagent (MPER; Thermo Scientific, Rockford, IL) supplemented with Halt protease and phosphatase inhibitors (Thermo Scientific). All animal protocols are approved by the Emory University Institutional Care and Use Committee.

Site-directed mutagenesis. Lysine residues 155 and 164 within the effector domain of MARCKS were mutated to glutamic acid residues by a QuikChange site-directed mutagenesis kit (Stratagene, La Jolla, CA). PAGE purified oligonucleotides (Integrated DNA Technologies) used for site-directed mutagenesis were 5’-AGG CTT TCA TTC GAA AAA TCT TTT GAG TTC AGT GGT TTC-3’ and its complement 3’-GAA ACC ACT CAA TTC AAA AGA TTT TTC GAA TGA AAA ACG CTT-5’. The underlined nucleotides correspond to the mutated bases. Constructs were confirmed by DNA sequencing.

Immunoprecipitation. Cells were lysed in MPER and 200 μg of total protein freshly supplemented with protease and phosphatase inhibitors (Thermo Scientific) was incubated with a 1:250 dilution of prewashed MARCKS polyclonal antibody at 4°C for 4 h with end-over-end mixing. Complexes were incubated with a 1:10 dilution of prewashed 50% protein G agarose (Millipore, Billerica, MA) at 4°C for 6 h with end-over-end mixing. The beads were washed four times with ice-cold MPER, and the bound proteins were eluted in 1× SDS sample buffer and analyzed by SDS-PAGE, Western blotting, or Coomassie staining.

SDS-PAGE. Western blotting, and densitometric analysis. Cells were washed once with 1× PBS before being scraped in MPER containing protease and phosphatase inhibitors (Thermo Scientific). The cell lysate was collected and then passed five times through a 23-gauge needle with a syringe before being incubated on ice for 1 h. The BCA protein assay (Thermo Scientific) was performed according to the manufacturer’s instructions to determine protein concentration. Samples were prepared in Laemmli sample buffer (Bio-Rad, Hercules, CA) containing 5% β-mercaptoethanol. One hundred micrograms of protein was resolved on 4–20% Tris-HCl polyacrylamide precast gels using Tris glycine/SDS buffer (Bio-Rad) on a Criterion or Protean Western blotting system (Bio-Rad). The separated proteins were electrophoretically transferred onto Hybond C-extra nitrocellulose membranes (GE Healthcare, Piscataway, NJ) using prechilled Tris glycine buffer on a Criterion or Protean Western blotting system (Bio-Rad). The membranes were then blocked in 5% (wt/vol) dry milk in 1× TBS (Bio-Rad) for 1 h at room temperature. Western blotting was performed by first incubating the blocked membranes with rabbit polyclonal primary antibodies (anti-phospho-α-filamin A; phospho Ser2152, Cell Signaling Technology), anti-filamin A (Cell Signaling Technology), anti-CaMKII (Cell Signaling Technology), anti-PKC-α (Cell Signaling Technology), anti-MARCKS (Abcam), and anti-ENaC-α (1), β (1), and γ (22) prepared at a 1:1,000 dilution in 5% BSA-1× TBS for 8 h at 4°C. The membranes were washed three times at 5-min intervals with 1× TBS before being incubated with horseradish peroxidase-conjugated goat anti-rabbit secondary anti-

body at a dilution of 1:3,000 prepared in blocking solution. The membranes were washed four times at 4-min intervals with 1× TBS and incubated with SuperSignal Dura Chemiluminescent Substrate (Pierce) according to the manufacturer’s instructions. The membranes were subjected to exposure using a Kodak Gel Logic 2200 Imager and Molecular Imaging software system (Carestream Health, Rochester, NY). Precision Plus Protein Standards (Bio-Rad) were used as markers to determine the relative molecular masses of the immunoreactive bands.

Recombinant protein production. Recombinant proteins were expressed and purified as previously described (1–4).

Single-channel patch-clamp studies. Micropipettes were pulled from filamented borosilicate glass capillaries (TW-150F, World Precision Instruments, Sarasota, FL) using a two-stage vertical puller (Narishige, Tokyo, Japan) and had a resistance of 6–10 MΩ. 2F3 cells were cultured to confluence on glutaraldehyde-fixed, collagen-coated Millipore-CM filters (Millipore) mounted on the bottom of Lucite rings. At room temperature, cells were visualized with Hoffman modulation optics (Modulation Optics), and, after the pipette tip contacted the cell surface, negative pressure was applied to obtain a seal resistance of 10–20 GΩ. Physiological amphibian saline titrated with 0.1 N NaOH or HCl to a pH of 7.3–7.4 and consisting of (in mM) 95 NaCl, 3.4 KCl, 0.8 CaCl₂, 0.8 MgCl₂, and 10 HEPES was used for the patch pipette and extracellular bath solutions. A cell-attached patch configuration was used, and voltages are given as the negative of the patch pipette potential. Positive potentials represent depolarizations, while negative potentials represent hyperpolarizations of the cell membrane away from the resting potential. The product of the number of functional channels and the open probability was calculated using pCLAMP 10 software (Molecular Devices) and represents a measurement of ENaC activity within a patch.

Detergent extraction and sucrose density gradient centrifugation. Cells were lysed in freshly prepared ice-cold 1% Brij 96/TNE buffer (10 mM Tris-HCl, pH 7.5, 150 mM NaCl, 5 mM EDTA, 2 mM Na vanadate, and protease and phosphatase inhibitor cocktail). The cell lysate was collected and passed five times through a 23-gauge needle with a syringe before being incubated on ice for 1 h. Lysates were centrifuged at 10,000 rpm for 5 min at 4°C. Five hundred microliters of the supernatant was mixed with an equal volume of freshly prepared 80% sucrose in TNE (without Brij 96) and then transferred to a 13 × 23-mm Beckman centrifuge tube. Eighty percent microsomes were freshly prepared 35% sucrose in TNE were carefully layered on top of the mixture followed by 500 μl of freshly prepared 5% sucrose. The sucrose gradient was then subject to ultracentrifugation in a SW50.1 rotor (Beckman) at 34,000 rpm for 16 h at 4°C. Fifteen fractions of 250-μl volume were carefully collected from the top to the bottom of the tube and then analyzed by Western blotting using flotillin and μ2 antibodies as protein markers.

Measurements of transepithelial voltage, resistance, and current. Measurements were made from confluent cells using an epithelial voltmeter (EVOM, World Precision Instruments) under sterile conditions. Transepithelial current was calculated by Ohm’s law and expressed as microamperes per square centimeter. To calculate amiloride-sensitive sodium current, the difference in total current and amiloride-insensitive current was determined after the addition of amiloride (0.5 μM) to the apical side of the inserts. Statistical analysis. A P value of <0.05 was considered significant. One-way ANOVA was performed for the comparison of more than one group.

RESULTS

Calmodulin inhibition augments endogenous ENaC activity in Xenopus 2F3 cells. We previously demonstrated a role for MARCKS and the actin cytoskeleton in the regulation of ENaC activity in Xenopus 2F3 cells (1, 29). Calmodulin is a calcium binding protein that is known to regulate the function of
MARCKS (14) and organization of the actin cytoskeleton (11). Here, we treated Xenopus 2F3 cells with a pharmacological inhibitor of calmodulin, calmidazolium, and performed single-channel patch-clamp recordings to determine whether ENaC activity is sensitive to calmodulin. Single-channel patch-clamp studies showed a significant increase in amiloride-sensitive ENaC current after application of calmidazolium, in which the activity was rescued after inducing calcium mobilization by application of ionomycin (Fig. 1A). Consistent with these results, analysis of single-channel patch-clamp data showed a statistically significant increase in the open probability of ENaC in response to calmidazolium and decrease in the open probability of the channels after ionomycin treatment (Fig. 1B).

Calmodulin directly binds to MARCKS and CaMKII but not to ENaC in a calcium-dependent manner. To determine whether the calmodulin effect on ENaC was in part due to direct binding between the two proteins and to confirm calmodulin binding to MARCKS and CaMKII in Xenopus 2F3 cells, we performed glutathione-S-transferase (GST) pulldown assays. Purified GST-calmodulin immobilized on a glutathione Sepharose support was used as the bait protein, and a crude cellular lysate from Xenopus 2F3 cells was used as the source of prey proteins, including ENaC, MARCKS, and CaMKII. GST pulldown assays showed calmodulin does not directly bind to ENaC, but does bind to MARCKS and CaMKII in a calcium-dependent manner (Fig. 2).

The calmodulin-dependent regulation of MARCKS involves lysine residues 155 and 164. The myristoylation domain of MARCKS inserts itself hydrophobically within the inner leaflet of the plasma membrane, and the MARCKS effector domain interacts electrostatically with acidic lipids (5). Both domains of MARCKS are required for sufficient association with the
The effector domain of MARCKS plays multiple roles in the regulation of the protein because it also contains the calmodulin binding site. It is plausible that mutating the calmodulin binding site or introducing negative charges within the effector domain of MARCKS would alter calmodulin in the membrane. We made a single construct consisting of lysine residues 155 and 164 being mutated to glutamic acid residues by site-directed mutagenesis to evaluate both of these assumptions. First, we cotransfected green fluorescent protein (GFP)-calmodulin with MARCK-cyan fluorescent protein (CFP) and investigated the effect of calcium augmentation on GFP-calmodulin translocation. We observed rapid translocation of GFP-calmodulin to the membrane after application of exogenous calcium (Fig. 3).

CaMKII inhibition augments endogenous ENaC activity in Xenopus 2F3 cells. The subcellular translocation of calmodulin and its effect on ENaC is dependent on calcium. The calcium-calmodulin complex regulates the activity of the serine/threonine-specific CaMKII. To determine whether CaMKII is involved in modulating ENaC activity, transepithelial resistance and voltage were recorded from Xenopus 2F3 cells and used to calculate amiloride-sensitive transepithelial current. Amiloride-sensitive transepithelial current increased in a time-dependent manner in response to application of the pharmacological inhibitor of CaMKII, KN93 (Fig. 4). Next, single-channel patch-clamp recordings were performed to corroborate a role for CaMKII in modulating ENaC activity and to determine whether CaMKII affects the open probability of ENaC. There was a statistically signif-

![CaM-GFP and MARCKS-CFP](image)

**A**

Control

10mM CaCl₂

**B**

Mutant

10mM CaCl₂

Fig. 3. Confocal microscopy and site-directed mutagenesis analysis of calcium-CaM-induced MARCKS translocation. A: Xenopus 2F3 cells were cotransfected with CaM-green fluorescent protein (GFP) and MARCK-cyan fluorescent protein (CFP). Forty-eight hours after transfection, cells were imaged for calmodulin-GFP and MARCKS-CFP. A moderate amount of calmodulin-GFP and MARCKS-CFP is expressed at the apical membrane in cells exposed to basal levels of calcium. An increase in CaM translocation to the apical membrane and MARCKS displacement from the membrane is shown for cells challenged with high levels of calcium (10 mM). B: Xenopus 2F3 cells were cotransfected with CaM-GFP and a mutant MARCKS-CFP (lysine residues 155 and 164 mutated to glutamic acid residues). Forty-eight hours after transfection, cells were imaged for calmodulin-GFP and MARCKS-CFP. Similar to A, a moderate amount of calmodulin-GFP and mutant MARCKS-CFP is expressed at the apical membrane in cells exposed to basal levels of calcium. An increase in CaM translocation to the apical membrane in response to calcium is shown, but the displacement of MARCKS from the membrane as seen in A is abrogated with the mutant MARCKS. All images shown represent the apical membrane and are the first of a series of images taken from the top to the bottom of the inset. Phase-contrast images show transfection efficiency and confluence of cells.

![Calibration graph](image)

![Experiment graph](image)

Fig. 4. Amiloride-sensitive transepithelial current measurements in Xenopus 2F3 cells after inhibition of CaMKII with KN93. Xenopus 2F3 cells grown on permeable supports were maintained in culture for 10 days to allow for the formation of tight junctions and the generation of measurable voltages and resistances across the monolayers. The pharmacological inhibitor of CaMKII, KN93 (0.5 μM) was applied to the apical side of Xenopus 2F3 cells at time 0 after recording of baseline voltage and resistance. Transepithelial voltages and resistances were used to calculate the current across the monolayers. A transient increase in transepithelial current was observed after 1 h of application of KN93. The increase in transepithelial current was sustained for an additional hour. At the end of the experiment, amiloride (0.5 μM) was applied to the apical side of the cells as a control (not shown).
significant increase in the open probability of ENaC after application of the CaMKII inhibitor (Fig. 5).

MARCKS and MLP-1 protein expression is sensitive to CaMKII inhibition. We previously reported a shift in MARCKS protein expression from light to heavy fractions with cytochalasin E treatment (29). In those studies, we used a recombinant antibody against the entire MARCKS protein to detect MARCKS and MLP-1 expression. To determine whether both MARCKS and MLP-1 protein expression are also sensitive to CaMKII, we performed similar studies in which we treated Xenopus 2F3 cells with the CaMKII inhibitor KN93 before performing sucrose density gradient and Western blot analysis of the different fractions. We found total protein expression levels of both MARCKS (Fig. 6, top bands) and MLP-1 (bottom bands) did not change significantly, but instead the expression of each protein shifted from heavy to light fractions in response to CaMKII inhibition (Fig. 6).

Calcium attenuates binding between ENaC subunits and filamin. Although the interaction between ENaC and filamin has been reported, the molecular determinants governing the interaction are largely unknown. Here, we investigated the role of calcium in the regulation of ENaC activity. To elucidate the mechanism for this novel regulation, we investigated the effect of calcium on the association between ENaC subunits and filamin. There is a strong association between filamin and ENaC-α, β, and γ in Xenopus 2F3 cells cultured in basal levels of calcium (Fig. 8). The association between each ENaC subunit and filamin was attenuated in Xenopus 2F3 cells cultured in high levels of calcium (Fig. 8). We also investigated the association between PKC-α and filamin, since we previously showed MARCKS promotes the PIP2-dependent regulation of ENaC. PKC is known to regulate the function of MARCKS, and here we show PKC-α interacts with filamin in a calcium-dependent manner similar to ENaC and filamin (Fig. 8).

CaMKII inhibition and disruption of the cytoskeleton decrease phospho-filamin A. Filamin A plays a role in linking the cytoskeleton to the plasma membrane. Wang et al. (39) showed filamin binds ENaC and decreases its activity. As indicated by studies by Bourguignon et al. (7), filamin may be a substrate of CaMKII. Ohta and Hartwig (25) showed filamin is phosphorylated in vitro at serine residues by CaMKII. Therefore, we wanted to determine whether CaMKII contributes to the phosphorylation of filamin in Xenopus 2F3 cells. We also wanted to determine whether disruption of the actin cytoskeleton affects filamin phosphorylation. We treated Xenopus 2F3 cells with either the cytoskeleton disrupter cytochalasin E or the CaMKII inhibitor KN93 and then performed SDS-PAGE and Western blot analysis, while probing for phospho- and total filamin protein. As shown in Fig. 7, both cytochalasin E and KN93 treatment attenuated the amount of phospho-filamin.

![Fig. 5. Single-channel patch-clamp recordings measuring the effect of the CaMKII inhibitor KN93 on ENaC P₀ in Xenopus 2F3 cells. A: representative single-channel patch-clamp recordings illustrating inhibition of CaMKII with the pharmacological inhibitor KN93 (0.5 μM) increases the activity of ENaC. B: summary line graph showing the P₀ of ENaC increases after inhibiting CaMKII with KN93. Values are means ± SE. *P < 0.05.](image)

![Fig. 6. Sucrose density gradient and Western blot (WB) analysis of MARCKS and MARCKS-like protein 1 (MLP-1) in response to CaMKII inhibition in Xenopus 2F3 cells. Western blot analysis probing for MARCKS and MLP-1 shows the total amount of MARCKS and MLP-1 protein did not change, but the protein density shifted from heavy to light fractions. A greater amount of MARCKS and MLP-1 protein appear in lanes 1 and 4 with KN93 treatment (0.5 μM) compared with the vehicle (MOCK) treatment.](image)

![Fig. 7. WB analysis of filamin and phospho-filamin in response to CaMKII inhibition and actin cytoskeleton disruption. Protein expression for phospho-filamin (bottom blot) decreased after CaMKII inhibition with KN93 or actin cytoskeleton disruption with cytochalasin E. Similar blots were probed for total filamin protein expression (top blot) as a control.](image)
cells were cultured in either normal (1.05 mM CaCl₂) or high calcium (4 mM CaCl₂) for 8 h before being lysed in the presence of protease and phosphatase filamin and ENaC PKC-inhibitors. The lysate was used to immunoprecipitate ENaC subunits and cells treated with high levels of calcium.

DISCUSSION

Multiple groups have shown sodium absorption in cortical collecting ducts is inhibited by increases in intracellular calcium (26, 31). Studies by Robins and Sandle (31) suggest increases in intracellular calcium inhibit wild-type ENaC but not Liddles-mutated ENaC, and this occurs by the removal of sodium channels from the apical membrane. This is consistent with observations by Plant et al. (28) which showed calcium can mobilize Nedd4 to the membrane. This would presumably decrease the density of the sodium channels at the membrane. Calmodulin has been suggested to play a role in calcium-dependent inhibition of ENaC activity, although the exact mechanism is unclear (31).

We recently reported a role for MARCKS (1) and the actin cytoskeleton (29) in the regulation of ENaC activity in Xenopus 2F3 cells. In this study, we show ENaC activity is modulated by calcium-calmodulin and CaMKII. It is likely that the actions of the calcium-calmodulin complex and CaMKII are linked, and the modulation in ENaC activity that is seen in response to pharmacological inhibition of either calmodulin or CaMKII involves other proteins including MARCKS/MLP-1 and other cytoskeletal-associated proteins. There are multiple mechanisms that could link the regulation of ENaC by calcium-calmodulin. Erk-1/2 phosphorylation is affected by CaMKII activity. Classic work by Illario et al. (18) showed inhibition of CaMKII activity by specific inhibitors inhibits Erk-1/2 phosphorylation by Ras-mediated activation of Raf-1. Accumulating evidence suggests Erk phosphorylation decreases ENaC cell surface expression by facilitating its interaction with the ubiquitin ligase Nedd4 (36).

The CaMKII-dependent activation of Erk1/2 is one mechanism for the regulation of ENaC activity. There is a large body of evidence that links ENaC to the actin cytoskeleton (19, 29, 44). CaMKII has been shown to participate in the reorganization of the actin cytoskeleton. Interestingly, Barros and Marshall (6) provided evidence for the Erk 1/2 MAP kinase pathway in the regulation of the actin cytoskeleton. This study, however, explores a different mechanism by which CaMKII can modulate ENaC activity through the regulation of the actin cytoskeleton. An intact cytoskeleton may stabilize ENaC at the membrane by multiple levels. First, it may potentiate ENaC interaction with MARCKS, which in turn promotes the interaction between ENaC and anionic phospholipids such as PIP₂. We have recently shown MARCKS sequesters PIP₂ and serves as a link between PIP₂ and ENaC to increase channel open probability in Xenopus 2F3 cells (1). Second, the actin cytoskeleton may protect ENaC at the cell surface from clathrin-mediated endocytosis and degradation. The association between multiple cytoskeletal elements, such as MARCKS, spectrin, fodrin, actin, and filamin, may prevent binding of the ubiquitin ligase Nedd4-2 to ENaC if there is overlap between the binding sites. In this study, we investigated the ability of CaMKII to modulate the expression of several different proteins that are associated with the actin cytoskeleton. We compared different cellular fractions, including lipid rafts, cytoplasmic, and subplasma membrane by sucrose density gradient and Western blot analysis. We found that protein expression levels of MARCKS and MLP-1 between these different fractions was altered and the phosphorylation state of filamin was sensitive to CaMKII activity in Xenopus 2F3 cells. The shift in MARCKS/MLP-1 to fractions where ENaC is present in response to KN93 compared with MOCK treatment suggests endogenous CaMKII activity may alter the association between ENaC and MARCKS/MLP-1. The inhibition of endogenous CaMKII activity appears to cause a rearrangement and shift of actin cytoskeleton-associated proteins. Similarly, we observed a stronger association between ENaC subunits and MARCKS after inhibiting endogenous CaMKII activity (data not shown). These findings, in parallel with the rapid increase in ENaC activity we see after application of the CaMKII inhibitor KN93 are consistent with our hypothesis that MARCKS promotes ENaC activity when the actin cytoskeleton is intact. Additionally, the increased expression of MARCKS protein in light sucrose density gradient fractions containing lipid rafts after inhibition of CaMKII is also consistent with the observed increase in ENaC activity.

![Fig. 8. Immunoprecipitation (IP) and WB analysis showing the effect of calcium on the association between ENaC subunits and filamin.](http://ajprenal.physiology.org/)

![Fig. 9. Proposed model depicting the role of calcium, CaM, and CaMKII on ENaC activity in Xenopus 2F3 cells.](http://ajprenal.physiology.org/)
The effector domain of MARCKS and MLP-1 contains basic amino acid residues that are important for its association with the inner leaflet of the plasma membrane. We wanted to determine whether introducing negative charges within the effector domain would attenuate the association between MARCKS and the membrane. We mutated two lysines residues within the effector domain of MARCKS to glutamic acid residues, with one of these lysine residues residing within the calmodulin binding domain of MARCKS. Although the mutant MARCKS protein was able to translocate to and associate with the apical membrane of Xenopus 2F3 cells, its sensitivity to calcium-calmodulin was reduced. This indicates that introducing two negative charges is not enough to prevent MARCKS from interacting with plasma membrane and that either or both lysine 155 and lysine 164 are essential for the translocation of the protein.

We previously showed disruption of the actin cytoskeleton with cytochalasin E treatment reduces total fodrin protein expression and causes a change in its subcellular expression as demonstrated by sucrose density gradient and Western blot analysis (29). Our findings suggest disruption of the cytoskeletal complex reduces ENaC activity in these cells. In a different mechanism, Harris et al. (13) originally demonstrated fodrin proteolytic cleavage by calcium-dependent protease I is regulated by calmodulin binding. In our proposed model, either of these mechanisms that would presumably disrupt the association between ENaC and fodrin would affect ENaC activity. We did not observe any significant changes in the levels of fodrin protein expression in different sucrose density gradient fractions (data not shown) as we did for MARCKS and MLP-1. However, we did confirm a role for CaMKII in the phosphorylation of filamin in Xenopus 2F3 cells.

Here, we investigated whether proteins that link the cytoskeleton to the plasma membrane, such as filamin, are subject to regulation by CaMKII. Wang et al. (39) showed filamin directly binds ENaC and inhibits its activity. Filamin links actin filaments to membrane glycoproteins and potentiates the interaction between these proteins and various cytoplasmic signaling proteins. Bourguignon et al. (7) showed CaMKII upregulation leads to phosphorylation of filamin, and this reduces its interaction with filamentous actin to promote tumor cell migration. Nakamura et al. (24) provided the first direct evidence for the regulation of filamin by calcium-calmodulin. Biochemical studies revealed filamin is phosphorylated by CaMKII (7). Phosphorylation of filamin by CaMKII attenuates its binding to filamentous actin. Several different kinases have been shown to be able to phosphorylate filamin A and regulate its function as an actin cross-linking protein. Cukier and coworkers (10) showed cyclin B1/cyclin-dependent kinase 1 (Cdk1) binds and phosphorylates filamin A at a serine residue. The association between ENaC and filamin is interesting because this actin binding protein has been shown to link the actin cytoskeleton to β-arrestins (34). Chen et al. (9) reported influenza virus-mediated inhibition of ENaC depends on Src activation, which can be mediated by β-arrestin. This mechanism may involve the ability of Src to activate PLC, which in turn can subsequently lead to the activation of PKC. Our laboratory and others have shown PKC inhibits amiloride-sensitive sodium current.

Taken together, our results provide evidence for a calmodulin- and CaMKII-dependent mechanism of ENaC regulation in cells expressed in the distal nephron. This particular mechanism involves phosphorylation of filamin, destabilization of the actin cytoskeleton, and release of MARCKS/MLP-1 from the membrane (Fig. 9). Our work shows for the first time calmodulin and CaMKII modulate the activity of ENaC at the level of its open probability.

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DISCLOSURES

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AUTHOR CONTRIBUTIONS


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