Novel Immunosuppressive Agents and Regimens

The calcineurin inhibitors (CNI) cyclosporine A (CsA) and tacrolimus (TAC) have significantly improved short-term outcomes after organ transplantation by strikingly minimizing the incidence of acute graft rejection. However, long-term graft and patient survival have not increased markedly (32). The evidence suggests that CNI toxicity (e.g., nephrotoxicity and cardiovascular morbidity) plays an important, albeit not unique, role in the failure to improve long-term outcomes in kidney transplantation (44). Thus, in recent years, researchers have focused on developing novel immunosuppressive strategies that could reduce CNI exposure. A meta-analysis of randomized clinical trials, providing data from >11,000 de novo renal transplant recipients, compared CNI-sparing with standard CNI-based regimens (61). Complete CNI avoidance, achieved by combining a mammalian target of rapamycin inhibitor with mycophenolate mofetil (MMF), resulted in an increased incidence of graft failure and poorer tolerability than standard immunosuppressive therapy with CNI. In contrast, minimizing of CNI in the setting of induction therapy with depleting or nondepleting antibodies, such as thymoglobulin, alemtuzumab, or basiliximab, combined with maintenance immunosuppressive agents, mainly MMF with or without steroids, was associated with a reduced incidence of graft failure and better renal function compared with standard-dose CNI exposure, without increasing the risk of acute cellular rejection (61).

Recently, however, it has been argued that regimens entailing CNI minimization or withdrawal may inadvertently increase the risk of antibody-mediated injury, thereby accelerating renal allograft loss (62). Indeed, increasing evidence indicates that inadequate immunosuppression (due to either noncompliance or immunosuppressive-sparing regimens) is the driving force behind the development of de novo donor-specific anti-human leukocyte antigen (HLA) antibodies (DSA), which are known to threaten long-term graft longevity (14). In relation to this, two studies have documented that converting from CsA to everolimus 3-5 mo after kidney transplantation resulted in a higher rate of de novo DSA and/or antibody-mediated rejection compared with continuing with CsA (36, 52). Note that de novo DSA development in the context of CNI minimization may also be affected by the type of induction therapy adopted. In particular, alemtuzumab treatment was associated with a higher incidence of de novo DSA than the combination of basiliximab and low-dose rabbit anti-thymocyte globulin in kidney transplant recipients given low-dose maintenance immunosuppressive drugs, including the CNI CsA (66).

In the past few years, increasing efforts have also been made to design novel biological agents (i.e., fusion proteins and antibodies) able to selectively target components of the host immune system, so as to avoid or limit the systemic toxic effects of CNI. This approach has led to the development of a second-generation cytotoxic T lymphocyte-associated protein (CTLA)4-Ig, belatacept, a receptor fusion protein consisting of the extracellular binding domain of CTLA4 linked to a modified human Ig (IgG1) Fe domain (33). Belatacept selectively inhibits CD28-mediated costimulation of T cells by binding to costimulatory ligands CD80/CD86 on antigen-presenting cells, a critical step for alloreactive T cell activation (20, 72). In 2011, the United States Food and Drug Administration approved belatacept as a first-line immunosuppressive agent based on the results of two pivotal phase III clinical trials that compared two belatacept-based regimens (more-intensive and less-intensive intravenous treatment schedules) with CsA in de novo kidney transplant patients who were also given basili-
imab, MMF, and steroids (16, 70). In both studies, belatacept-treated groups exhibited similar graft and patient survival rates compared with the CsA arm (16, 70), along with better renal function over a 5-yr posttransplantation followup (8, 53). Intriguingly, de novo DSA development was less frequent in patients that received belatacept, who also had better cardiovascular and metabolic risk profiles (69, 71). Despite these benefits, at 12 mo posttransplant, belatacept treatment was associated with higher rates of acute cellular rejection, mostly of high Banff grades, and posttransplant lymphoproliferative disorders (PTLD), especially in patients seronegative for the Epstein-Barr virus (16, 70). This may eventually make it necessary to limit CNI toxicity, another costimulatory blocker, the fully humanized anti-CD40 monoclonal antibody ASKP1240, is currently being investigated in clinical trials. The preliminary results of a phase II study showed that in de novo kidney transplant patients that received basiliximab and steroids, ASKP1240 in combination with low-dose TAC prevented acute graft rejection at 6 mo posttransplantation as effectively as standard-dose TAC alone (22). However, the efficacy of this new biological agent was not confirmed when TAC was avoided and ASKP1240 was given in combination with MMF, as confirmed by the high rate of acute cellular rejection with this immunosuppressive regimen (22). The higher risk of acute rejection episodes with belatacept or the combination ASKP1240 and MMF compared with CNI-based regimens may be at least partially explained by the inability of these agents to inhibit memory T cells, which are less dependent on costimulation for reactivation (29). Accordingly, the clinical success of costimulatory blockade may ultimately require a complementary immunosuppressive strategy taming memory T cell subsets.

**Cell-Based Therapy for Immune Tolerance Induction**

To date, transplantation tolerance has been intentionally achieved in a small number of kidney transplant recipients undergoing donor hematopoietic stem cell transplantation under a nonmyeloablative conditioning regimen (Table 1). These encouraging clinical studies are based on early experiments in

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**Table 1. Clinical studies of tolerance induction to kidney transplantation by donor HSC transplantation**

<table>
<thead>
<tr>
<th>Center</th>
<th>Nonmyeloablative Conditioning Regimen and Maintenance immunosuppression (Dose and Days in Respect to Kidney Transplant)</th>
<th>Donor HSCs (HLA Match, Dose, and Day of Cell Transplantation in Respect to Kidney Transplant)</th>
<th>Chimerism</th>
<th>Patient Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Stanford University</td>
<td>• Thymoglobulin: 1.5 mg/kg, days 0 to +5</td>
<td>HLAmatched</td>
<td>Mixed, stable (at least for the first 6 mo)</td>
<td>Total N = 22</td>
</tr>
<tr>
<td></td>
<td>• Total lymphoid irradiation: 10 doses of 80–120 cGy during the first 14 days posttransplant</td>
<td>Granulocyte colony-stimulating factor mobilized enriched CD34+ hematopoietic progenitors (4.3–17.5 × 10^6 cells/kg) mixed with CD3+ T cells (1–10 × 10^6 total cells/kg) at day +11</td>
<td>N = 16 off immunosuppression</td>
<td>N = 3 acute rejection</td>
</tr>
<tr>
<td>Massachusetts General Hospital</td>
<td>• Anti-CD2 monoclonal antibody: days −2, −1, 0, and +1</td>
<td>HLAmismatched</td>
<td>Mixed, transient (2–3 wk posttransplant)</td>
<td>Total N = 10</td>
</tr>
<tr>
<td></td>
<td>• Thymic irradiation: 700 cGy, days −1</td>
<td>Unprocessed bone marrow cells (2–3 × 10^6 cells/kg) at day 0</td>
<td>N = 5 off immunosuppression</td>
<td>N = 3 graft loss for acute humoral rejection (N = 1, 10 days posttransplant), thrombotic microangiopathy complication (N = 1, 7 mo posttransplant), and acute rejection (N = 1, 3 yr)</td>
</tr>
<tr>
<td></td>
<td>• Rituximab: 375 mg/m², days −5, −4, +2 (fourth and fifth patients) and days +5 and +12 (last five patients)</td>
<td></td>
<td>N = 2 belatacept for chronic rejection 5 and 7 yr posttransplant</td>
<td>N = 2 belatacept for chronic rejection 5 and 7 yr posttransplant</td>
</tr>
<tr>
<td></td>
<td>• Steroids: tapered in 10–20 days</td>
<td></td>
<td>N = 2 belatacept for chronic rejection 5 and 7 yr posttransplant</td>
<td>N = 2 belatacept for chronic rejection 5 and 7 yr posttransplant</td>
</tr>
<tr>
<td></td>
<td>• Maintenance: calcineurin inhibitors, tapered and discontinued in 8–14 mo</td>
<td></td>
<td>N = 2 belatacept for chronic rejection 5 and 7 yr posttransplant</td>
<td>N = 2 belatacept for chronic rejection 5 and 7 yr posttransplant</td>
</tr>
<tr>
<td>Northwestern University</td>
<td>• Cyclophosphamide: 50 mg/kg, days −3 and +3</td>
<td>HLAmismatched</td>
<td>Full, durable</td>
<td>Total N = 19</td>
</tr>
<tr>
<td></td>
<td>• Total body irradiation: 200 cGy, days −1</td>
<td>Engineered cellular product enriched for HSC and tolerogenic facilitating cells (2–33 × 10^6 total cells/kg) at day +1</td>
<td>N = 12 off immunosuppression</td>
<td>N = 5 on immunosuppression (transient chimerism)</td>
</tr>
<tr>
<td></td>
<td>• Fludarabine: 30 mg/kg, days −4, −3, and −2</td>
<td></td>
<td>N = 2 graft loss for viral sepsis (N = 1, 3 mo posttransplant) and for infection (N = 1, 9 mo posttransplant)</td>
<td>N = 2 graft loss for viral sepsis (N = 1, 3 mo posttransplant) and for infection (N = 1, 9 mo posttransplant)</td>
</tr>
</tbody>
</table>

HSCs, hematopoietic stem cells; MMF, mychophenolate mofetil; CsA, cyclosporine A.
mice (24) and later studies in patients with hematological malignancies (63, 64) showing that inducing mixed chimerism (coexistence of hematopoietic cells of both recipient and donor origin) through transplantation of donor bone marrow in recipients undergoing myeloablative conditioning achieved long-term tolerance toward a subsequent kidney transplant from the same bone marrow donor. Since then, similar tolerogenic protocols have been applied in clinics with noncologenous transplant recipients of living donor kidneys. These protocols have been adapted to design less toxic conditioning regimens capable of ablating the recipient bone marrow, eventually allowing various levels of chimerism to occur. Three investigator groups have recently reported the initial results of small clinical trials for transplant tolerance induction using donor hematopoietic stem cells in HLA-matched and mismatched kidney transplantation (Table 1).

The Stanford University study included 22 kidney transplant patients that received peripheral blood CD34+ stem cell infusions and kidney transplants from HLA-matched donors (57–59). Weaning maintenance immunosuppression (CsA/MMF) was based on the presence of durable mixed chimerism. Nineteen of twenty-two patients successfully developed persistent mixed chimerism after the peculiar nonmyeloablative conditioning regimen of the Stanford protocol (Table 1), and sixteen patients were successfully weaned off immunosuppressive drugs. Kidney graft tolerance was achieved independently of whether mixed chimerism, obtained in the first 6 mo posttransplantation, persisted or vanished during or after CsA discontinuation (58, 60). This finding questions about the role of persistent mixed chimerism in promoting graft tolerance, at least in the setting of the Stanford protocol. When applied to kidney transplant patients with HLA-mismatched donors (n = 6), this protocol failed to induce persistent mixed chimerism. Two of the six patients met the criteria for immunosuppressive drug withdrawal [transient chimerism, absence of clinical rejection episodes, and graft-versus-host disease (GVHD) and donor reactivity in a mixed lymphocyte reaction] and were withdrawn from immunosuppression at 12 mo after kidney transplantation. However, both patients developed rejection episodes 3.5 and 5.5 mo after drug discontinuation (40, 60). A more recent attempt in HLA haplotype-matched (3/6 HLA mismatches) recipients (n = 10) applied a dose escalation approach of CD3+ T cells (from 3–50 million) to the same total lymphoid irradiation and antithymocyte globulin conditioning regimen (60). Chimerism lasting at least 12 mo was observed in the two patients given the highest number of CD34+ cells (15–22 × 10^6 cells/kg) and in the three patients given the highest CD3+ T cell dose (50 × 10^6 cells/kg). In the two patients given the highest CD34+ cell dose, MMF was successfully discontinued and TAC tapering was in progress (60). This revised regimen is currently being tested in a larger cohort of patients to assess its efficacy in inducing persistent chimerism and immunosuppressive drug withdrawal in HLA-mismatched patients (60).

At Massachusetts General Hospital, Kawai and coworkers treated 10 patients with combined kidney and bone marrow transplantation from HLA-haploidentical (3/6 mismatches) related donors under nonmyeloablative conditioning (26, 28). Over time, the protocol was implemented with multiple rituximab doses to prevent the development of DSA, as observed in the initial few patients enrolled (28). Although donor mixed chimerism was completely lost in all patients within the first 21 days after combined cell and kidney transplantation, graft tolerance was achieved in 7 of 10 subjects, 5 of whom remained off immunosuppression long term (27, 28). However, all patients developed engraftment syndrome, characterized by severe but transient renal dysfunction during the first week after cell transplantation (19). Therefore, a modified version of the conditioning protocol with total body irradiation replacing cyclophosphamide has begun in an effort to prevent the engraftment syndrome (27).

Recent comparisons of outcomes of kidney transplant patients who achieved tolerance either by the Stanford (60) or Massachusetts General Hospital (27) protocols demonstrated that tolerant patients had improved graft survival versus patients on standard immunosuppression (60) and reduced incidence of immunosuppression-related complications such as hypertension, hyperlipidemia, de novo diabetes, malignancies, and infections (27).

Northwestern University has also reported successful induction of allograft tolerance in HLA-mismatched kidney transplant recipients by almost completely replacing the recipient hematopoietic cells with donor cells (full chimerism) (34, 35). The tolerogenic protocol includes transplanting the cellular product enriched with hematopoietic stem cells and "t tolerogenic facilitating" cells. "Facilitating" cells are a CD8+ T cell receptor (TCR)– bone marrow–derived mixed cell population that favor the engraftment of bone marrow without promoting GVHD (75). The mechanism through which these facilitating cells promote bone marrow engraftment appears to involve the development of regulatory T cells (Tregs) as well as IL-10–producing type 1 Tregs (75). Twelve of nineteen patients achieved durable chimerism and are off immunosuppressive drugs. Five other patients did not develop stable donor cell engraftment and are continuing with immunosuppressive drug therapy, and two subjects experienced allograft loss. The investigators reported that no GVHD has occurred for up to 5 yr despite very high levels of donor chimerism (35). Evaluation of immune reconstitution and immunocompetence in chimeric subjects revealed that most of these individuals retained memory for hepatitis B, measles, mumps, rubella, and varicella, suggesting that mixed, rather than full chimerism, was present in these transplant patients. In addition, chimeric recipients developed protective immunity in response to pneumococcal vaccination, providing evidence for immunocompetence (35).

Collectively, these studies documented the feasibility and efficacy of cell-based therapy to induce tolerance in organ transplant recipients. However, challenges related to the procedure and clarification of the mechanism(s) involved in the protolerogenic process must still be addressed before these tolerogenic cell-based protocols will be part of current kidney transplantation management. Indeed, cell-based therapies with bone marrow or hematopoietic stem cells, with or without facilitating cells, require transplant conditioning regimens to promote cell engraftment and chimerism (Table 1). These conditioning protocols have been modified to improve the clinical outcomes, but this has not eliminated their inherent dangers and the likelihood of infections and GVHD. This shortcoming must be balanced with the short- and long-term risks/benefits of current pharmacological immunosuppressive therapies in kidney transplantation. Furthermore, whether inducing chimerism is the cornerstone for achieving transplantation tolerance or an epiphenomenon remains unclear. According to the above studies, graft tolerance can be achieved...
after inducing either mixed (60) or almost full (35) chimerism or even with very low and transient (37) chimerism, yet it is not clear why some chimeras become tolerant to the donor kidney and some do not. In mouse models of mixed chimerism, the lifelong contribution of donor antigen-presenting cells that mediate the central deletion of donor-reactive T cells in the thymus is the major mechanism of long-term tolerance of the transplanted organs (55). The transient nature of chimerism in human patients (2, 37) and the rapid expansion of peripheral T cells after bone marrow or hematopoietic stem cell transplantation (35, 57, 58) would, however, suggest that peripheral rather than intrathymic mechanisms play a role in immune tolerance induction, at least immediately after organ transplantation. It has recently been proposed that in the long term, extrathymic deletion of donor-reactive CD4+ and CD8+ T cell clones plays a role in maintaining tolerance (42).

Although hematopoietic stem cell-based therapies hold promise for kidney transplantation, alternative cell-based treatments that do not require peritransplant conditioning regimens are being investigated for potential use in immunotherapy in solid organ transplantation.

Many immune cells with regulatory properties, such as tolerogenic dendritic cells, regulatory macrophages, Tregs, and mesenchymal stromal cells (MSCs) have been proposed for cell therapy (Table 2) (73). Among them, Foxp3+ Tregs, either naturally occurring or peripherally induced (25), and MSCs (7) have been particularly in the spotlight.

Tregs can exert dominant tolerance to alloantigens in vivo by inducing regulatory properties in alloreactive T cells in animal models (11) and may establish a tolerant state that could obviate the need for immunosuppressive drugs (65). The clinical use of regulatory cells in organ transplantation is currently being explored in several ongoing trials (Table 2) (67). Since Tregs are small in number in the peripheral circulation, they required ex vivo expansion before infusion into patients. Either expanded naturally occurring Tregs or induced Tregs from naïve CD4+ T cells stimulated by donor alloantigens are being proposed as cellular therapy to control the unwanted alloim-

Table 2. Regulatory cell-based therapy in kidney or liver transplant patients

<table>
<thead>
<tr>
<th>Study Identifier</th>
<th>Center</th>
<th>Regulatory Cells</th>
<th>Indication</th>
</tr>
</thead>
<tbody>
<tr>
<td>NCT02091232</td>
<td>Massachusetts General Hospital, Boston, MA</td>
<td>Tregs</td>
<td>Living donor kidney transplant</td>
</tr>
<tr>
<td>NCT02129881</td>
<td>Guy’s Hospital, London and Oxford Transplant Centre, Oxford, United Kingdom</td>
<td>Polyclonal Tregs</td>
<td>Living donor kidney transplant</td>
</tr>
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<td>NCT02371434</td>
<td>Charité University Medicine, Berlin, Germany</td>
<td>Polyclonal Tregs</td>
<td>Living donor kidney transplant</td>
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<tr>
<td>NCT02248001</td>
<td>University of California, San Francisco, CA</td>
<td>Polyclonal Tregs</td>
<td>Living donor kidney transplant</td>
</tr>
<tr>
<td>NCT02145325</td>
<td>Northwestern University, Chicago, IL</td>
<td>Polyclonal Tregs</td>
<td>Living donor kidney transplant</td>
</tr>
<tr>
<td>NCT02088931</td>
<td>University of California, San Francisco, CA</td>
<td>Polyclonal Tregs</td>
<td>Kidney transplantation</td>
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<tr>
<td>Planned</td>
<td>Ospedale San Raffaele, Milan, Italy</td>
<td>Antigen-specific type 1 Tregs (T10 cells)</td>
<td>Living donor kidney transplant</td>
</tr>
<tr>
<td>NCT02188719</td>
<td>University of California, San Francisco, CA, and Mayo Clinic, Rochester, MN</td>
<td>Donor alloantigen reactive Tregs</td>
<td>Living donor liver transplant</td>
</tr>
<tr>
<td>NCT2474199</td>
<td>University of California, San Francisco, CA, and Mayo Clinic, Rochester, MN</td>
<td>Donor alloantigen reactive Tregs</td>
<td>Living donor liver transplant</td>
</tr>
<tr>
<td>NCT02166177</td>
<td>Guy’s and St Thomas’ National Health Service Foundation Trust, London, United Kingdom</td>
<td>Autologous Treg product TR002</td>
<td>Liver transplant</td>
</tr>
<tr>
<td>NCT01624077</td>
<td>Nanjing Medical University, Jiangsu, China</td>
<td>Donor alloantigen-specific Tregs</td>
<td>Liver transplant</td>
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</table>

Dendritic cells

<table>
<thead>
<tr>
<th>Study Identifier</th>
<th>Center</th>
<th>Regulatory Cells</th>
<th>Indication</th>
</tr>
</thead>
<tbody>
<tr>
<td>NCT02252055</td>
<td>Nantes University Hospital, Nantes, France</td>
<td>Autologous tolerogenic dendritic cells</td>
<td>Living donor kidney transplant</td>
</tr>
<tr>
<td>NCT02085629</td>
<td>University of Regensburg, Regensburg, Germany</td>
<td>Donor-derived regulatory macrophages</td>
<td>Living donor kidney transplant</td>
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<tr>
<td>NCT02012153</td>
<td>Mario Negri Institute, Bergamo, Italy</td>
<td>Autologous MSCs</td>
<td>Living donor kidney transplant</td>
</tr>
<tr>
<td>NCT02409940</td>
<td>Institute of Medical Education and Research, Chandigarh, India</td>
<td>Autologous MSCs</td>
<td>Living donor kidney transplant</td>
</tr>
<tr>
<td>NCT02490020</td>
<td>Zhuijiang Hospital, Guangdong, China</td>
<td>MSCs</td>
<td>Kidney transplant</td>
</tr>
<tr>
<td>NCT01429038</td>
<td>University Hospital of Liege, Liege, Belgium</td>
<td>Third-party MSCs</td>
<td>Kidney and liver transplant</td>
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<tr>
<td>NCT02492490</td>
<td>Fuzhou General Hospital, Fujian, China</td>
<td>Third-party MSCs</td>
<td>Kidney transplant</td>
</tr>
<tr>
<td>NCT02387151</td>
<td>Leiden University Medical Center, Leiden, The Netherlands</td>
<td>Third-party MSCs</td>
<td>Kidney transplant</td>
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<tr>
<td>NCT02492308</td>
<td>Fuzhou General Hospital, Fujian, China</td>
<td>Autologous stromal-vascular fraction-derived MSCs</td>
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<td>NCT01841632</td>
<td>University Hospital Regensburg and Athersys Incorporated, Regensburg, Germany</td>
<td>Third-party multipotent progenitor cells</td>
<td>Liver transplant</td>
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</tbody>
</table>

Tregs, regulatory T cells; MSCs, mesenchymal stromal cells.
Tolerant patients showed an increased number of circulating B cells and overexpression of B cell-associated genes in the peripheral blood and urine (4, 45, 48, 56). In particular, tolerant patients showed specific expansion of transitional, naïve B cells (45, 48) and of B cells that express an inhibitory phenotype, such as FcγRIIB (a receptor transducing inhibitory signals) and B cell scaffold protein with ankyrin repeats-1 (which negatively modulates B cell-CD40-mediated AKT activation) (48, 56). Mechanistically, transitional B cells from operationally tolerant patients secreted high level of IL-10 after in vitro polyclonal stimulation (45) and expressed a higher level of microRNA-142 3p, the forced expression of which in the Raji B cell line upregulates the expression of numerous B cell immune response genes, including those previously identified in operationally tolerant patients (12). Moreover, B cells from tolerant patients did not fully differentiate into plasma cells (10) and suppressed in vitro effector T cell function by a contact- and granzyme B-dependent pathway (9). These findings suggest that B cells may actively regulate the immune response to the transplanted kidney promoting T cell unresponsiveness to donor alloantigens. Indeed, despite a normal phenotype (41), circulating T cells from tolerant patients showed decreased alloreactivity (23), which, however, does not involve an active Treg-mediated immunoregulation (23, 38). Until recently, the role of Tregs in spontaneous tolerance has remained ill defined. Initial studies showed that the number of circulating Tregs and their ex vivo regulatory properties were not significantly modified in operationally tolerant patients compared with those with stable graft function under standard immunosuppression (23, 38). A more recent study extensively characterized circulating Tregs in tolerant patients and found that operational tolerance was characterized by circulating CD4+ T cells with a high level of demethylation of the FoxP3 Treg-demethylated region and by expansion of Foxp3high memory T cells with stronger suppressive properties compared with patients with stable graft function on immunosuppression or with healthy volunteers (3). Interestingly, this study shed new light on the potential role of memory Tregs in long-term graft survival (3).

Overall, these findings suggest that long-term graft acceptance in operationally tolerant patients involves a complex interplay of mechanisms that ultimately lead to emergence of B cells and T cells with suppressive and regulatory functions.

On this line, a recent longitudinal analysis of the B cell tolerance signature in tolerant (spontaneous or induced) patients showed that the increased frequency of transitional and naïve B cells as well as the increased expression of B cell-associated genes persist over time in operationally tolerant patients. Of major interest, a similar overexpression of the most predictive B cell-associated genes was found in kidney transplant patients who achieved tolerance using the Massachusetts General Hospital protocol of combined bone marrow transplantation under the nonmyeloablative conditioning regimen (46), suggesting that spontaneous and induced immune tolerance in kidney transplantation may share similar biomarkers or even mechanisms.

Conclusions

In the past few years, novel immunosuppressive agents for preventing graft rejection in organ transplantation, including kidney transplantation, have been developed and approved.
Unfortunately, the initial promise of significant improvements in the management and outcomes of transplant patients has only been partially fulfilled, and major concerns remain for their safety profile and early as well as long-term efficacy. This calls for more collaborative work between basic and clinical academic research institutions and pharmaceutical companies to better dissect the complex cellular and humoral mechanisms underpinning the long-term inexorable deterioration of graft function that still occurs in a large percentage of organ transplant recipients. It also means there is a need to develop more targeted and safer immunosuppressive agents. In parallel, at this stage, more small clinical studies are needed so that the safety and complexity of the immunomodulatory and pro-tolerogenic properties of the novel cell-based therapies as facilitators of successful transplantation can be appreciated fully and so that preclinical discoveries can translate into tangible benefits for organ transplant recipients. Overall, these considerations may be particularly important for new biological agents, such as cell-based therapies, where despite encouraging initial results, uncertainty about safety and efficacy still exists.

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REFERENCES

Perspective

NOVEL IMMUNOMODULATORY THERAPIES IN KIDNEY TRANSPLANTATION


45. Scandling JD, Buske E, Dejbakhsh-Jones S, Benike C, Sarwal M, Millan MT, Shizuru JA, Lowsky R, Engleman EG, Strober S. Tolerance...


