Repeated administration of low-dose cisplatin in mice induces fibrosis

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Acute kidney injury (AKI), the rapid loss of kidney function, has many medical complications, a high mortality rate, and limited therapeutic interventions beyond palliative care (45). One of the most prominent causes of AKI is pharmaceutical-induced nephrotoxicity, which accounts for 19% of all cases of AKI (44). Cisplatin is a potent chemotherapeutic used for the treatment of many cancers but induces AKI in 30% of patients even in the absence of comorbidities such as advanced age or preexisting kidney diseases (35). Clinically, cisplatin is given at low doses in multiple rounds to try to avoid nephrotoxic side effects, but even with this precaution in place, AKI still occurs (3). Blood urea nitrogen (BUN) and serum creatinine (SCr) are clinical tests used to monitor kidney function but are highly insensitive (9, 10). Elevated BUN or SCr levels in patients during the course of cisplatin treatment requires that the dose of cisplatin be lowered or delayed or, alternatively, that the patient be switched to a potentially less effective chemotherapeutic that lacks nephrotoxic side effects (3). None of these options are favorable and can result in a less efficacious cancer treatment.

Until recently, it was assumed that patients that survive AKI and don’t require dialysis are able to achieve full recovery of kidney function (23). However, recent large-scale longitudinal studies that assessed the impact of AKI on long-term renal function have indicated that patients that had AKI are more likely to develop CKD and end-stage renal disease (ESRD) than patients with no history of AKI. Furthermore, the occurrence of CKD/ESRD was directly proportional to the level and frequency of AKI experienced by these patients (1, 11, 12, 16, 28–30, 42). Additionally, the incidence of AKI and CKD/ESRD has significantly increased in the past decade (16, 27, 47), as the overall age of our population is also increasing (18). This is of importance because kidney function declines with normal aging even in the absence of obvious kidney disease (2). The majority of individuals receiving nephrotoxic chemotherapeutics are middle aged or older and already have increased exposure to renal stressors, enhanced susceptibility to injury, and decreased ability to repair after injury. With improved diagnosis and treatment of cancers, there is also increased long-term patient survival. Hence, the percentage of cancer survivors expected to develop CKD/ESRD will increase, placing a major burden on patient quality of life and our healthcare system.

Gaining an understanding of the cellular processes involved in the development of CKD after cisplatin-induced AKI would be useful for developing renoprotective agents. Unfortunately, the standard dosing mouse model of AKI does not allow for long-term studies. In the standard dosing model, a single high dose of cisplatin (>20 mg/kg) is administered once to 8- to 10-wk-old mice. This dose is lethal to the mouse beyond 72 h, and, as a result, examining the AKI to CKD transition is not possible. Furthermore, this model does not accurately represent the repeated dosing regimen used in the clinic. Thus, there is a need for a more clinically relevant mouse model that enables the study of the cellular processes involved in the progression of cisplatin-induced AKI to CKD.

We have developed a model of cisplatin-induced kidney injury that reflects the repeated low dosing of cisplatin used in the clinic and allows for detailed analysis of repair, recovery, and long-term kidney function. Here, mice were treated with 7 mg/kg cisplatin once a week for 4 wk and euthanized 3 days...
after the last injection. We compared this dosing regimen with mice that were treated according to the standard dosing model of AKI (one dose of 25 mg/kg cisplatin and euthanized 3 days later). We analyzed markers of kidney function and injury, inflammatory cytokines and chemokines, indicators of endoplasmic reticulum (ER) stress and cell death, and profibrotic indicators. While the standard dosing model of AKI and the repeated dosing model have similarities in their effects on kidney function, kidney damage is less severe in the repeated dosing model, enabling mice to survive beyond the 24-day course of treatment. Data also indicated that interstitial fibrosis occurred in the repeated dosing model but not in the standard dosing model. Data suggested that the increased incidence of CKD after cisplatin-induced AKI may be a result of repeated injury leading to fibrosis.

### MATERIALS AND METHODS

**Reagents and antibodies.** The following antibodies were purchased from Cell Signaling (Beverly, MA) unless otherwise noted: cleaved caspase-3 (no. 9664), C/EBP homologous protein (CHOP; no. 2895), JNK Signalin (no. 9258), phosphorylated (p-JNK) (no. 4668), transforming growth factor (TGF)-β (no. 37125), fibrotenon (F3648, Sigma-Aldrich, St. Louis, MO), p-SMAD3 (no. 12747), α-tubulin (SC-23948, Santa Cruz Biotechnology, Dallas, TX), and β-actin (SC-47778, Santa Cruz Biotechnology). Cisplatin (P4394, Sigma-Aldrich) was used for experiments comparing the effects of a high single dose (euthanization 3 days later) with the new repeated dosing model. Pharmaceutical grade cisplatin (purchased directly from the University of Louisville hospital pharmacy) was used for experiments comparing the effects of single versus repeated injury from cisplatin. Similar effects were observed for both sources of cisplatin.

**Animals.** FVB/n mice were purchased from The Jackson Laboratory (Bar Harbor, ME). All mice were maintained on a 12:12-h light-dark cycle and provided food and water ad libitum. Animals were maintained under standard laboratory conditions. All animal procedures were approved by the Institutional Animal Care and Use Committee of the University of Louisville and followed the guidelines of the American Veterinary Medical Association. Cisplatin at 25 mg/kg in PBS (200 μl/animal) was administered by an intraperitoneal injection. Seventy-two hours after cisplatin injection, these mice were euthanized. Another cohort of mice received either 7 or 9 mg/kg cisplatin administered by an intraperitoneal injection once a week for 4 wk. For these survival experiments, mice were monitored for weight loss and signs of discomfort/distress on a daily basis. Mice exhibiting a weight loss of 20% or more total body weight or high levels of discomfort and stress were euthanized. Mice that survived the course of treatment were euthanized 72 h after the fourth injection of cisplatin. Serum was prepared and stored at −80°C. The kidneys were flash frozen in liquid nitrogen or fixed in 10% formalin.

**BUN and Scr determination.** BUN (DIUR-500) and SCr (C7548-120) were determined using kits from Bioassay Systems (Hayward, CA) and Point Scientific (Canton, MI), respectively, following the manufacturers’ instructions. For SCr, this specific assay kit uses a two-reagent enzymatic assay system to eliminate interference by endogenous creatine and ascorbic acid.

**Protein quantification.** Western blot analysis, and ELISAs. Homogenates were made from the kidney cortex by homogenization in cell extraction buffer [10 mM Tris-HCl (pH 8.0), 150 mM NaCl,1 mM imidazole, 1 mM magnesium acetate, 20 mM EGTA, and 10 mM β-mercaptoethanol] containing a Complete Protease Inhibitor Cocktail Tablet and Phosphatase Inhibitor Cocktail Tablets (Roche, Indianapolis, IN). Homogenates were centrifuged at 15,000 g for 10 min at 4°C. Supernatants were removed, aliquoted, and stored at −80°C until use. Protein concentrations were determined using Bradford Reagent (Bio-Rad, Hercules, CA). Kidney homogenate (40 μg) was separated on 4–12% gradient Tris-glycine-SDS polyacrylamide gels and transferred to polyvinylidene difluoride membranes, which were blocked with 5% (wt/vol) dried milk in Tris-buffered saline containing 0.1% Tween 20 (TBST) for 1 h. Membranes were incubated with 1:5,000 dilutions of primary antibody overnight at 4°C. The next morning, membranes were washed three times for 5 min each with TBST containing 5% (wt/vol) dried milk. After incubation for 1 h at room temperature with secondary antibodies conjugated with horse-radish peroxidase [1:50,000 in TBST containing 1.25% (wt/vol) dried milk], membranes proteins were detected by chemiluminescence substrate. ELISAs for kidney injury molecule (KIM)-1 (DY1817, R&D Systems, Minneapolis, MN) and neutrophil gelatinase-associated lipocalin (NGAL; DY1857, R&D Systems) were performed on the urine as directed by the manufacturer.

**Gene expression.** Total RNA was isolated using RNA-STAT 60 (TEL-TEST, Friendswood, TX) combined with mini-bead-beater glass beads and a Mini Bead Beater machine (Cole-Palmer, Vernon Hills, IL) following the manufacturer’s protocol. cDNA was made from 1 μg total RNA using High-Capacity Reverse Transcriptase (Life Technologies, Grand Island, NY) following the manufacturer’s instructions. Gene-specific cDNAs were quantified using real-time PCR and predesigned TaqMan assays. TNF-α (Mm00443258_m1), IL-6 (Mm00461901_m1), IL-1β (Mm00343228_m1), chemokine (C-C motif) ligand (CXCL1) (Mm04207460_m1), monocytic chemotactic protein (MCP)-1 (Mm00441242_m1), plasminogen activator inhibitor (PAI)-1 (Mm00435860_m1), α-smooth muscle actin (α-SMA; Mm1546133_m1), bone morphogenetic protein (BMP)-7 (Mm01432102_m1), collagen type IV (Col4a1; Mm00488051_m1), cyclooxygenase 2 (COX-2; Mm00750399_m1), and growth factor (FGF)-2 (Mm00481269_m1) were purchased from Life Technologies and used in combination with 2× Gene Expression Master Mix (Life Technologies).

**Histology.** Kidney sections (5 μm thick) from cisplatin-treated and untreated animals were stained with hematoxylin and eosin as well as periodic acid-Schiff, and the degree of morphological changes was determined by light microscopy in a blinded fashion by a renal pathologist. The following measures were chosen as an indication of morphological damage to the kidney after treatment with vehicle or cisplatin: proximal tubular necrosis, loss of the brush border, proximal tubule degradation, tubular casts, presence of inflammatory cells, and interstitial fibrosis. These measures were evaluated on a scale from 0 to 4, which ranged from not present (0) to mild (1), moderate (2), severe (3), and very severe (4).

**Immunohistochemistry.** Kidney sections (5 μm thick) were rehydrated in Histoclear followed by an ethanol gradient. Antigen retrieval was performed in citric acid buffer (pH 6.0) at 95°C in a steamer for 30 min. Endogenous peroxidases were inhibited with 3% hydrogen peroxide and dual endogenous enzyme blocker (Dako) for 10 min followed by two 5-min PBS washes. Slides were then blocked with avidin for 10 min followed by a PBS wash and then biotin for 10 min followed by a wash in PBS (Dako). Slides were further blocked with 5% normal goat serum in 0.1% TBST for 1 h at room temperature. α-SMA primary rabbit antibody (Abcam) was added to slides at a concentration of 0.5 μg/ml and allowed to incubate at 4°C overnight. Slides were rinsed with PBS for 5 min, three times. Biotinylated goat anti-rabbit IgG antibody (1: 25,000, BA-1000, Vector Laboratories) was added to each section and incubated for 30 min at room temperature. Slides were rinsed twice with PBS (5 min each). Vector ABC reagent (PK-7100, Vector Laboratories) was added to each section and incubated for 30 min at room temperature. Slides were rinsed two times with PBS followed by the addition of 100 μl of DAB substrate for 5-7 min to detect horseradish peroxidase (SK-4800, Vector Laboratories). Slides were rinsed in distilled water for 5 min, counterstained with modified Mayer’s hematoxylin (no. 72804, Thermo Scientific).
were euthanized 3 days after the fourth treatment (day 24) due to moribund status. Mice treated with the 7 mg/kg standard dosing regimen were euthanized 3 days after cisplatin mg/kg) once a week for 4 wk. All mice subjected to the model, 25 mg/kg) with mice given a dose of cisplatin (7 or 9 mg/kg) once a week for 4 wk (repeated dosing model) or with 25 mg/kg cisplatin given once (standard dosing model). Mice were monitored daily for weight loss and changes in overall well-being and were euthanized when moribund in accordance to Institutional Animal Care and Use Committee guidelines. At day 24, surviving mice were euthanized and analyzed.

RESULTS

Effects of dosing regimens on mouse survival. The current model used to study cisplatin-induced AKI does not allow for the analysis of long-term effects on kidney function, nor does it recapitulate the repeated nature of the dosing regimen of cisplatin in the clinic. We hypothesized that administration of a low dose of cisplatin once a week for several weeks would be more clinically relevant and allow for the analysis of long-term effects on kidney function. To test this, we compared survival of mice given a single high dose of cisplatin (standard dosing model, 25 mg/kg) with mice given a dose of cisplatin (7 or 9 mg/kg) once a week for 4 wk. All mice subjected to the standard dosing regimen were euthanized 3 days after cisplatin injection due to moribund status. Mice treated with the 7 mg/kg repeated dosing regimen survived until day 24 (Fig. 1). Statistical analyses comparing survival curves of 7- and 9 mg/kg-treated mice revealed no statistical significance, but there was statistical significance between repeated dosing and standard dosing survival curves (Fig. 1). These data indicate that the repeated dosing model with 7 mg/kg enables the survival of all treated mice for long-term studies of kidney function.

Effects of dosing regimens on kidney injury and function. To assess the impact of repeated dosing on the kidney, we measured markers of kidney function and injury in the serum and urine of mice, respectively. BUN levels of mice treated with the standard dosing model were significantly increased at 72 h posttreatment (Fig. 2A). In the repeated dosing model, BUN also increased, but not significantly (Fig. 2A). SCr levels were significantly increased for both the repeated and standard dosing models, but SCr levels were higher in the standard dosing model (Fig. 2A). Urinary KIM-1 and NGAL levels were examined, as they are more sensitive biomarkers of AKI than

Sirius red/fast green staining. Kidney sections (5 μm thick) were rehydrated in Histoclear followed by an ethanol gradient. Slides were then dipped into a Coplin jar containing PBS with 0.1% Tween 20 and incubated for 5 min. Slides were washed with distilled water twice for 5 min each and then incubated in saturated picric acid containing 0.1% Sirius red and 0.1% fast green. Sirius red/direct red 80 (catalog no. 365548) and fast green FCF (catalog no. F7258) were from Sigma, whereas picric acid [saturated ~1.2% (wt/vol)] was from Ricca Chemicals (catalog no. S860-32). Slides were washed with 5% glacial acetic water until the water ran clear. Tissue samples were then dehydrated and fixed using Permunt (F-SP15-100, Fisher Scientific). Positive staining for Sirius red was quantified using Metamorph Image Analysis software, and the percentage of positive pixels was calculated as follows: [threshold area/total area − acellular area].

Statistical analysis. Data are expressed as means ± SE for all experiments. Multiple comparisons of normally distributed data were analyzed by one-way ANOVA, as appropriate, and group means were compared using Tukey posttests. Single comparisons were analyzed by Student’s t-test where appropriate. For statistical analysis of the survival curve, a log-rank (Mantel-Cox) test was used. The criterion for statistical differences was P < 0.05, P < 0.01, P < 0.001, and P < 0.0001.

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Fig. 2. Comparison of kidney function and injury between the standard dosing model and repeated dosing model. Eight-week-old male FVB mice were injected (intraperitoneally) with saline vehicle or 25 mg/kg cisplatin given once (standard dosing model) or with cisplatin (7 mg/kg) once a week for 4 wk (repeated dosing model). Animals were euthanized 72 h after the last injection. A: levels of blood urea nitrogen (BUN) and serum creatinine (SCr) measured in the serum. B: kidney injury molecule (KIM)-1 and neutrophil gelatinase-associated lipocalin (NGAL) measured in the urine. Data are expressed as means ± SE; n = 10. Statistical significance was determined by Student’s t-test. ***P < 0.01; ****P < 0.001.

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fibrosis (Fig. 5, increase in the presence of inflammatory cells and interstitial increase in cast formation (Fig. 5) in both models. Interest-

D

fibrosis in the repeated dosing model but not in the standard dosing model. Since pathology of kidney sections revealed tubulointerstitial damage. Blinded analysis by a certified pathologist indicated that tubular necrosis was significantly higher in the standard dosing model compared with the repeated dosing model (Fig. 4). These data suggest similar effects of both dosing regimens on inflammatory cytokines and chemokines.

Effect of dosing regimen on activation of ER stress and cell death proteins. Cell death and ER stress are characteristic of cisplatin-induced AKI. It is known that inhibition or deletion of key players in pathways of apoptosis or ER stress protects the kidney from cisplatin-induced injury in the standard dosing model (22, 25, 39). Therefore, we assessed cellular markers of ER stress and cell death proteins in both models. JNK phosphorylation and activation are associated with ER stress-induced apoptosis (37, 49). We found that p-JNK was elevated in both models (Fig. 4). CHOP is also associated with ER stress and was also activated in both models (Fig. 4) (14). However, cleaved caspase-3, a marker of apoptosis, was not increased in the repeated dosing model (Fig. 4). These data suggest that while both models show similar trends in activation of ER stress proteins, there may be less cell death activation in the repeated dosing model.

Effect of dosing regimen evident in tissue pathology. The standard dosing model of cisplatin-induced AKI is associated with changes in kidney pathology. We compared kidney pathology of the standard and repeated dosing models by examining tubular necrosis, loss of brush borders, tubule dilation, cast formation, presence of inflammatory cells, and interstitial fibrosis, all of which are indicative of kidney injury and damage. Blinded analysis by a certified pathologist indicated that tubular necrosis was significantly higher in the standard dosing model compared with the repeated dosing model (Fig. 5A). In contrast, there was a significant loss of brush borders (Fig. 5B), an increase in tubular dilation (Fig. 5C), and an increase in cast formation (Fig. 5D) in both models. Interestingly, only the repeated dosing model displayed a significant increase in the presence of inflammatory cells and interstitial fibrosis (Fig. 5, E and F). These data demonstrate that there are key differences in kidney pathology between the standard and repeated dosing models.

Fibrotic markers and fibrosis in the repeated dosing model. Since pathology of kidney sections revealed tubulointerstitial fibrosis in the repeated dosing model but not in the standard model, we examined known markers of fibrosis in this model. After kidney injury, TGF-β is released from immune cells (32, 37). TGF-β can then signal through its receptor, leading to the phosphorylation of SMAD3, thereby activating pathways that increase extracellular matrix protein deposition, particularly fibronectin (8, 40). BMP-7 is also a member of the TGF-β superfamily and works to counteract the profibrotic activity of TGF-β (33). TGF-β, p-SMAD3, and fibronectin were all increased at the protein level in the repeated dosing model (Fig.
DISCUSSION

Treatment of human cancers with cisplatin often leads to nephrotoxic side effects that are cumulative and dose dependent. AKI occurs in some individuals even after one low dose of cisplatin, and multiple episodes of AKI can cause CKD (13). Whereas Kobayashi et al. (31) performed repeated cisplatin dosing in mice to determine circadian changes related to drug administration, in the present study, we developed a model to study the effects of multiple “hits” of cisplatin-induced AKI and the subsequent development of CKD. The standard dosing model of cisplatin-induced AKI has limitations that cannot be

Comparison of a single low dose of cisplatin (7VVV) with the repeated dosing model. To determine if fibrosis is a result of a single low dose of cisplatin or rather repeated injury from several low doses, mice were administered a low dose of cisplatin (7 mg/kg) followed by three weekly vehicle injections (7VVV) and compared with mice that received four weekly cisplatin injections (7777). BUN and SCr both increased significantly after repeated dosing (Fig. 6B). CDKN2A encodes for p16, and increased expression of CDKN2A is associated with cell cycle arrest and cellular senescence (49). mRNA expression of CDKN2A was significantly increased after repeated dosing (Fig. 6B). Furthermore, collagen deposition as the result of extracellular matrix production can be quantified with Sirius red and fast green staining and found that collagen levels increased in the kidneys after repeated dosing of cisplatin (Fig. 6C). α-SMA is a marker of myofibroblasts, which are known to deposit collagen (31, 34). α-SMA immunohistochemistry indicated increased myofibroblasts after repeated dosing of cisplatin (Fig. 6D). Taken together, these data indicate that there are alterations in key mediators of kidney fibrosis in the repeated dosing model.

A–F: Standard Repeated

<table>
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<tr>
<th>Treatment (mg/kg)</th>
<th>Standard</th>
<th>Repeated</th>
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<tbody>
<tr>
<td>tubule necrosis</td>
<td>0</td>
<td>0</td>
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<tr>
<td>loss of brush borders</td>
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<td>collagen deposition</td>
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<td>interstitial fibrosis</td>
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Fig. 5. Qualitative analysis of kidney pathology indexes. Renal histological changes were assessed on hematoxylin and eosin- and periodic acid-Schiff-stained kidney sections (5 μm thick). Eight-week-old male FVB mice were injected (intraperitoneally) with saline vehicle or cisplatin (7 mg/kg) once a week for 4 wk (repeated dosing model) or with 25 mg/kg cisplatin given once (standard dosing model). Animals were euthanized 72 h after the last injection.

A: tubular necrosis. B: loss of proximal tubule brush borders. C: proximal tubule dilation. D: proximal tubule cast formation. E: presence of inflammatory cells. F: interstitial fibrosis. In A–F, scoring of the sections was performed in a blinded manner by a renal pathologist (J. Megyesi) using a scale of 0-4 (where 0 = not present, 1 = mild, 2 = moderate, 3 = severe, and 4 = very severe renal histological changes in proximal tubules). Data are means ± SE; n = 5–10. Statistical significance was determined by one-way ANOVA followed by Tukey’s multiple-comparison test. ***P < 0.001.
overcome for studying the AKI to CKD progression. For one, a single high-dose regimen is not clinically relevant. Patients are administered multiple low doses of cisplatin over an extended period of time. Second, the standard dosing model cannot be aged out to study long-term effects associated with repeated AKI, namely, fibrosis, the underlying pathology of CKD (7). In the present study, we report a model for studying the nephrotoxic effects of cisplatin that mimics the repeated administration of cisplatin given clinically and allows for analysis of long-term effects on kidney function. Data obtained from this model indicated that repeated cisplatin injury induces interstitial fibrosis and suggest that targeting fibrotic mediators may prevent both short- and long-term renal side effects of cisplatin.

The standard dosing model of cisplatin induces high levels of kidney injury and cell death through apoptosis and necrosis. This, in turn, results in a rapid loss of kidney function. With the repeated dosing model, cleaved caspase-3 as a measure of apoptosis is low, and pathology reveals a low level of tubular necrosis. This translates to lower injury levels and a smaller decline in overall kidney function. These lower levels of injury and the maintenance of kidney function with the repeated dosing model may be key to explaining how mice treated with multiple low doses of cisplatin are able to survive for 24 days and perhaps even beyond that.

In the standard dosing model of cisplatin-induced AKI there is a strong inflammatory response, which involves TNF-α elevation and elevation of its downstream targets. In the repeated dosing model, a similar inflammatory response is observed. However, IL-6 expression is not as elevated in the repeated dosing model. IL-6 plays a role in mounting an effective immune response and has been indicated in AKI. Particularly, IL-6 expression is correlated with the onset and severity of AKI and has been indicated as a potential urinary and plasma biomarker of AKI (19, 36, 50). The low levels of IL-6 mRNA measured in the repeated dosing model may be indicative of a less severe form of AKI and help explain why less injury is occurring in this model.

Pathology indicates that there is a significant increase in infiltrating immune cells in the repeated dosing model, despite less injury. It has been shown that rapid increase in the macrophage population results in the development of fibrosis (48). Macrophages are known to play a major role in mounting an effective repair response postinjury and have also been indicated in maladaptive repair (24). Whereas M2 macrophages play a role in normal repair, an increase in M1 macrophage population results in the development of fibrosis (48). Macrophages are known to play a major role in mounting an effective repair response postinjury and have also been indicated in maladaptive repair (24). Future studies of this model will focus on identifying the type of infiltrating immune cells, whether the M1 to M2 transition is indicative of a less severe form of AKI and help explain why less injury is occurring in this model.

The repeated administration of low-dose cisplatin induces fibrosis, and this is a physiologically relevant process that could be targeted therapeutically. Grbic et al. (26) have shown that kidney function can recover after a single round of injury induced by diphtheria toxin in transgenic mice.
expressing the diphtheria toxin receptor in proximal tubule cells. However, repeated injury in this model culminated in fibrosis, as determined by increased levels of TGF-β1, fibronectin, and Col1α1 (26). In our model of repeated cisplatin dosing, we also found increased protein levels of TGF-β and fibronectin and increased mRNA expression of Col1α1.

G2/M cell cycle arrest, cellular senescence, and fibrosis have been indicated in the ischemia-reperfusion and unilateral ureteral obstruction mouse models (49). We found increased

Fig. 7. Comparison of single low dose and repeated dosing. Eight-week-old male FVB mice were injected (intraperitoneally) with saline vehicle or cisplatin (7 mg/kg) once (VVVV and 7VVV, respectively) or with cisplatin (7 mg/kg; 7777) once a week for 4 wk (repeated dosing model). Animals were euthanized 72 h after the last injection. A: levels of BUN and SCr assessed in the serum. B: markers of kidney fibrosis assessed via Western blot analysis. C: mRNA levels of inflammatory cytokines and fibrotic markers in the kidney cortex as measured by qRT-PCR. D: SRFG staining of kidney sections and quantification of staining. E: α-SMA immunohistochemical staining in the kidney and quantitation of staining. Statistical significance was determined by one-way ANOVA followed by Tukey’s multiple-comparison test. *P < 0.05; **P < 0.01; ***P < 0.001.
mRNA expression of CDKN2A, which is suggestive of cellular senescence. Furthermore, p-JNK has been indicated as a downstream target of G2/M cell cycle arrest and is believed to also play a role in promoting renal fibrosis (49). While further studies are needed to determine whether or not G2/M arrest is indeed occurring after repeated cisplatin dosing, we did observe an increase in p-JNK without evidence of apoptosis. Thus, the increase in p-JNK may be indicative of G2/M arrest occurring and, as a result, potentially maladaptive repair. Taken together, these data suggest that potential mechanisms such as senescence and G2/M arrest warrant future indepth investigation in this model to identify the mechanisms by which fibrosis is induced.

Fibrosis plays a major role in our repeated dosing model and is indicative that this model can be used to perform extensive mechanistic studies of the AKI to CKD progression. For example, studies should be completed to determine the type and role for infiltrating immune cells in fibrosis in the kidney. Furthermore, it would also be worthwhile to examine kidney function, injury, fibrosis, and inflammatory markers throughout the course of cisplatin treatment rather than just at the end of the repeated dosing regimen. This would provide further insights into when fibrosis occurs temporally. Along with determination of the temporal timeline by which fibrosis occurs, structural studies looking at remodeling of the extracellular matrix during this process would provide insights into the morphological processes that occur within kidney tissue. Fortunately, Torres et al. (43), through the application of clearing multiphoton microscopy, have gained new, detailed morphological insights into the pathophysiology involved in the AKI to CKD transition. While Torres et al. (43) used a dosing regimen consisting of only two relatively high doses of cisplatin, they found that significant remodeling of the extracellular matrix was occurring, although they did not document major increases in collagen levels.

Fibrosis not only plays a role in CKD but also has been indicated in cancer metastasis (17). Our repeated cisplatin dosing model could be adapted to a cancer model to also look at the fibrosis both in the kidneys and cancer. Pabla et al. (38) have shown that repeated administration of 10 mg/kg cisplatin for 4 wk does alter the tumor size in a cancer xenograft model and also leads to an increase in BUN and SCr levels, indicating loss of kidney function. While they did not look at fibrosis in this model, it would be interesting to determine whether cancer-associated fibrosis occurs with our repeated dosing model. In the present study, we used mice on the FVB background rather than the C57BL/6 mouse strain as mice on the C57BL/6 background are extremely resistant to developing interstitial fibrosis (46). In addition, numerous transgenic mouse models of cancer are available on the FVB background for studying the effects of repeated low-dose cisplatin in mice with cancers that develop in the proper tumor microenvironment. Thus, the data presented in this study serve as the foundation for future studies aiming to determine the impact of repeated cisplatin dosing on the tumor and the kidney. Perhaps by further elucidating the mechanism by which fibrosis occurs, we can find new, desirable targets for development of both renoprotective and cancer therapeutics.

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DISCLOSURES
No conflicts of interest, financial or otherwise, are declared by the author(s).

AUTHOR CONTRIBUTIONS

REFERENCES


